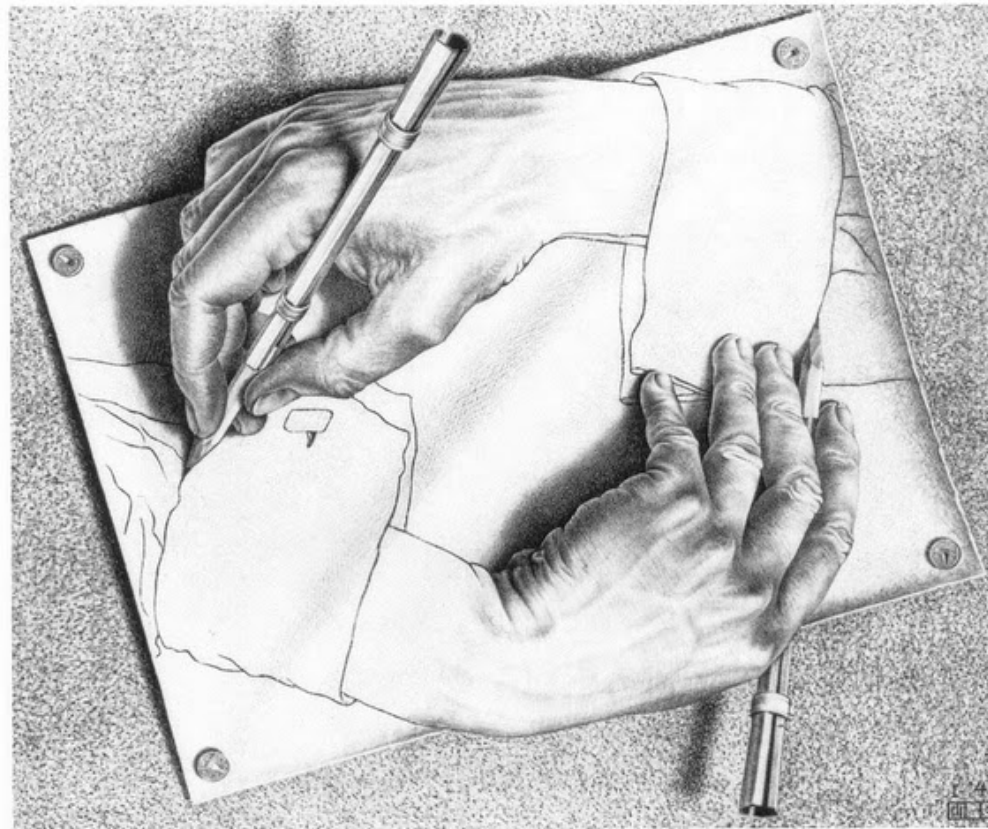


– Not SMRT yet smart – 3D genomics with Hi-C sequencing

Sylvain Foissac, GenPhySE

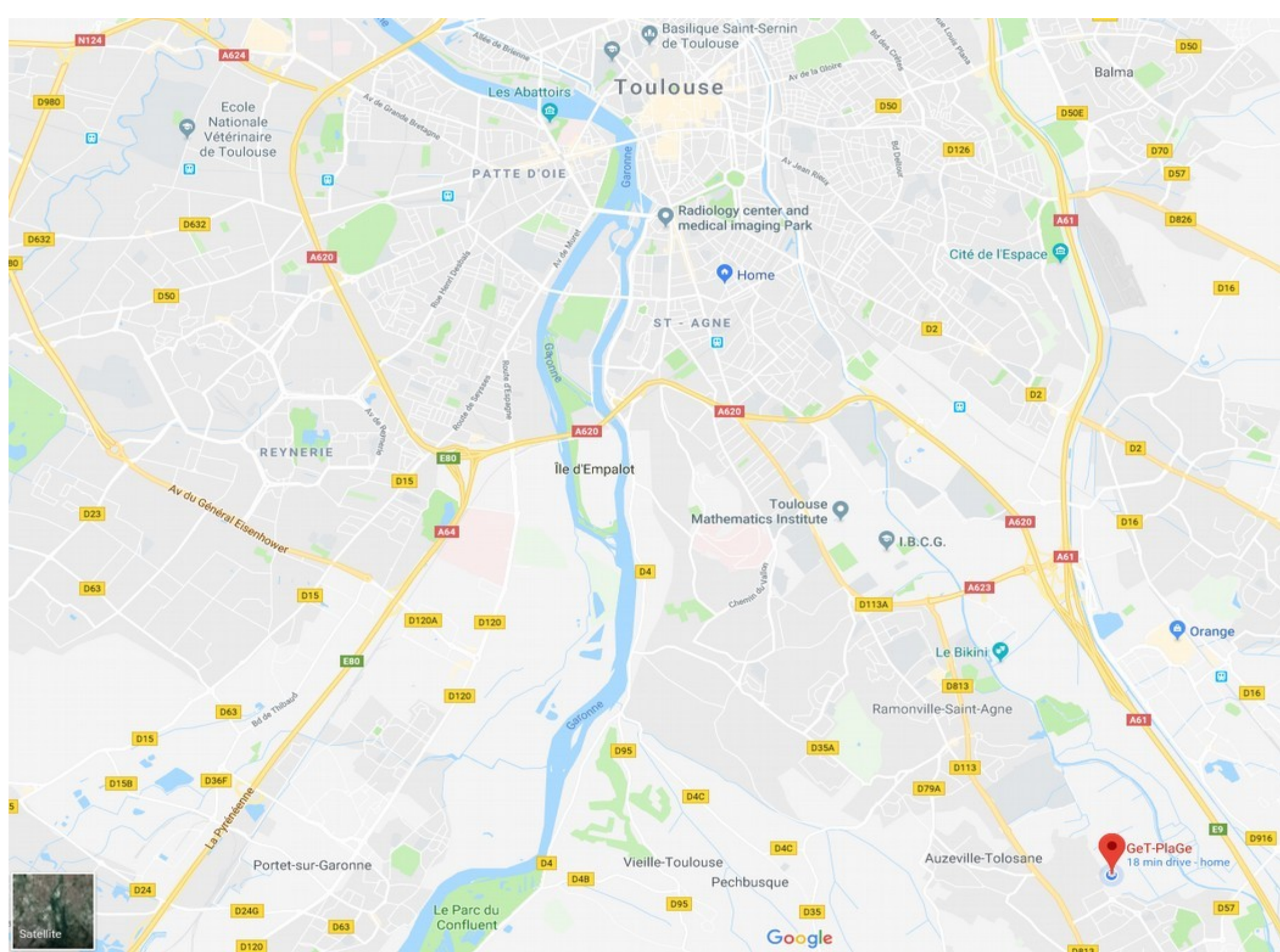
November 28, 2017. Get Genotoul, INRA Toulouse, France

From
structure ...



https://en.wikipedia.org/wiki/M._C._Escher

... to
function





Chapelle des Carmélites

Musée Saint-Raymond

Basilique Saint-Sernin de Toulouse

Syndicat CGT

Sept

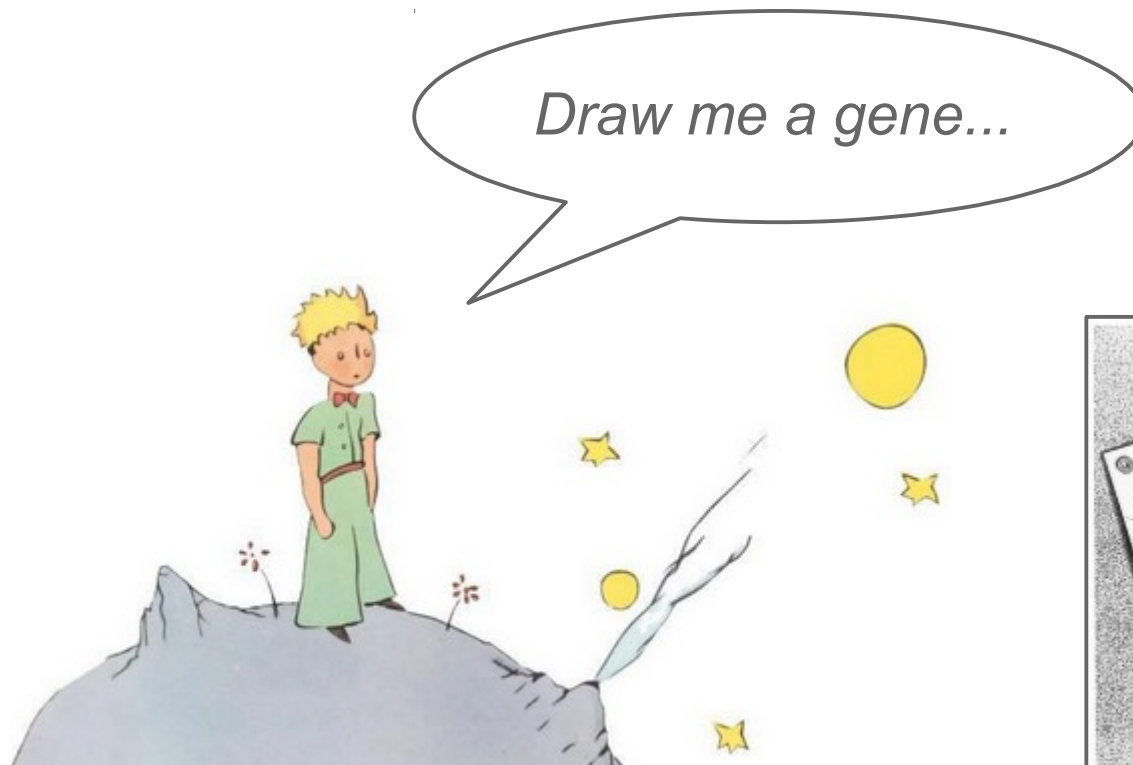
Lycée Général Saint-Sernin

Copy Shop, Corep Lautman

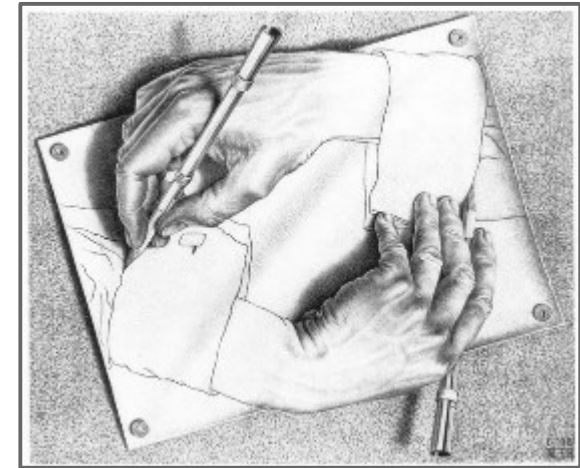
The Geo



Intro: from structure to function



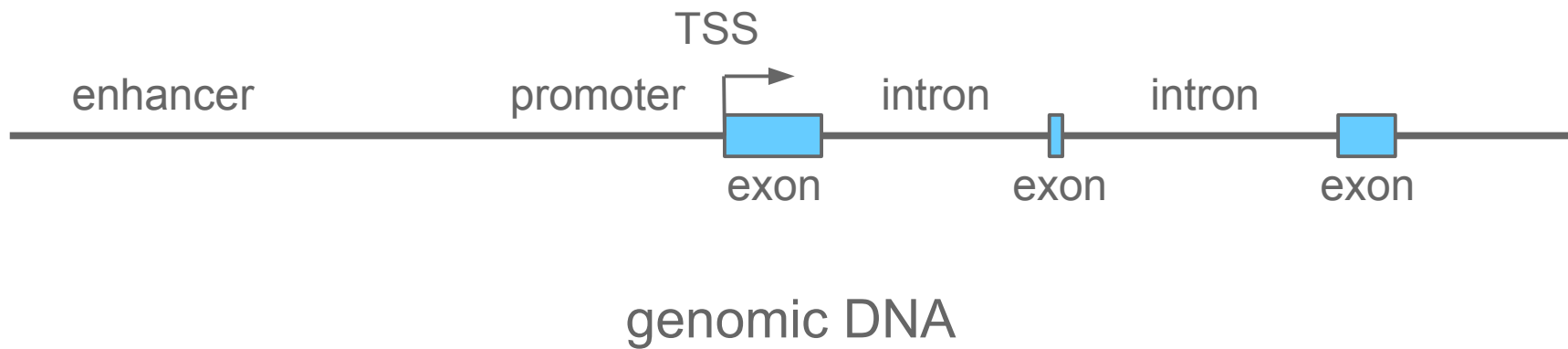
De Saint Exupéry A., 1943



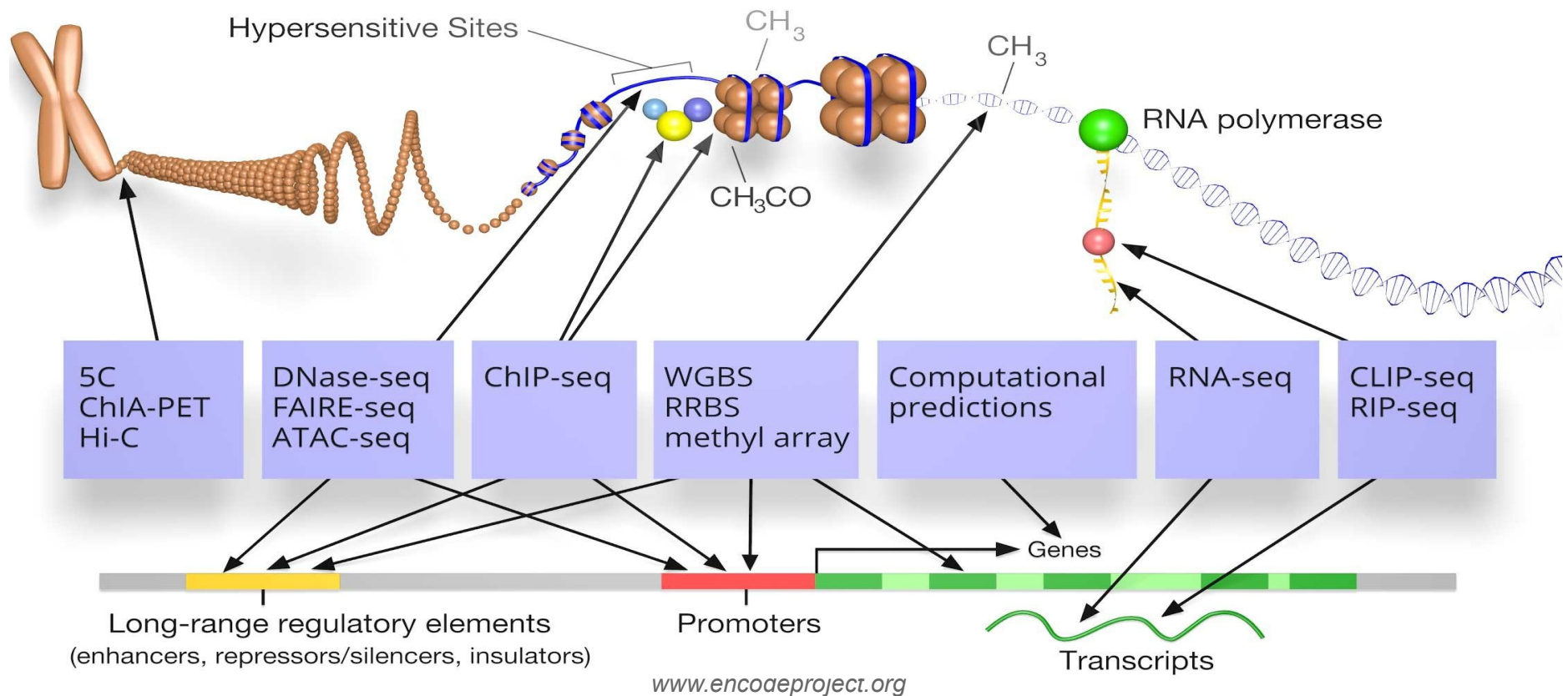
Intro: from structure to function

genomic DNA

Intro: from structure to function



Intro: from structure to function



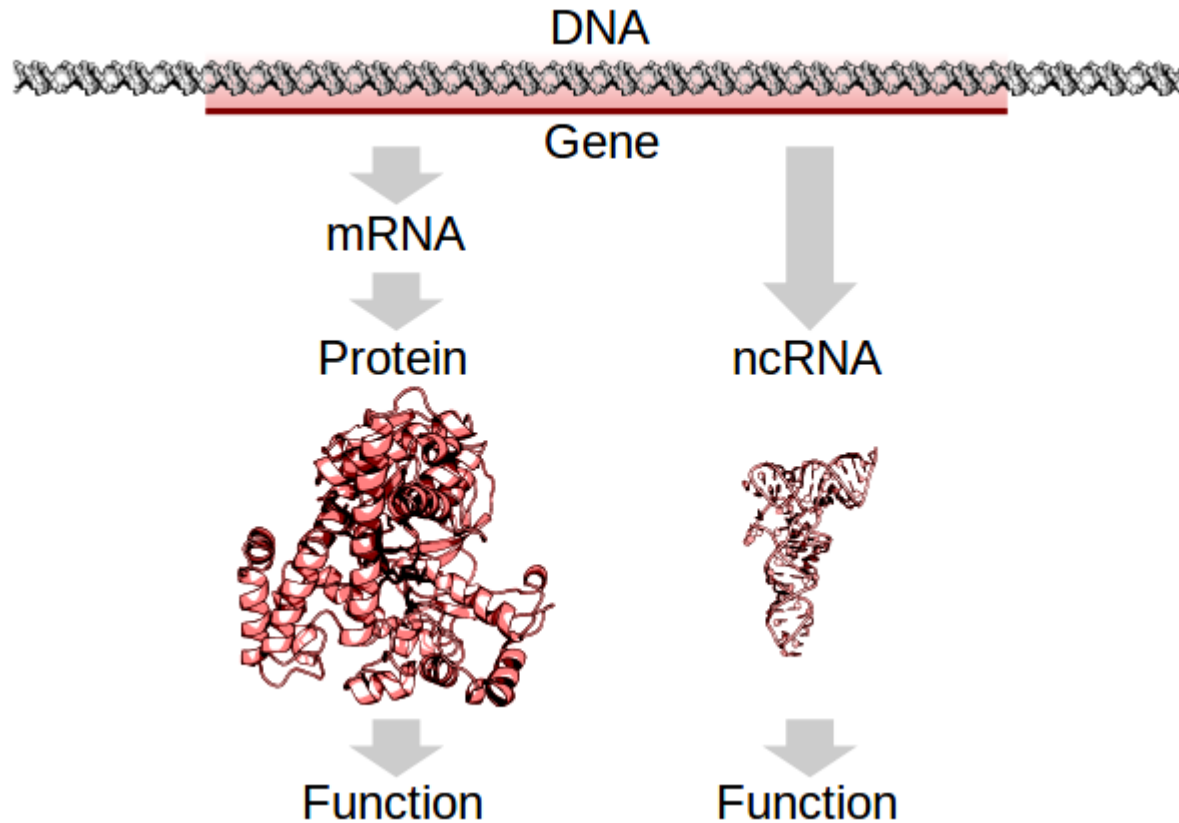
Human:



Livestock:

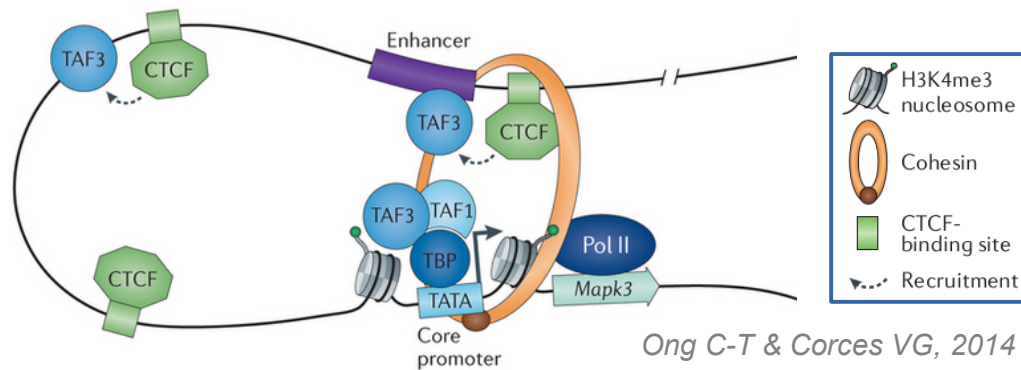
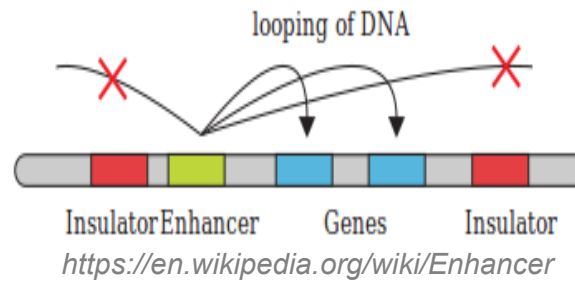


Intro: from structure to function



<https://en.wikipedia.org/wiki/Gene>

Intro: from structure to function



Intro: from structure to function

Cell

From structure to function

Chromosomal Contact Permits Transcription between Coregulated Genes

Stephanie Fanucchi,¹ Youtaro Shibayama,¹ Shaun Burd,¹ Marc S. Weinberg,^{3,4} and Musa M. Mhlanga^{1,2,*}

¹Gene Expression and Biophysics Group, Synthetic Biology Emerging Research Area, Biosciences Unit, Council for Scientific and Industrial Research, Pretoria, Gauteng 0001, South Africa

²Unidade de Biologia e Expressão Genética, Instituto de Medicina Molecular, Faculdade de Medicina, Universidade de Lisboa, Lisboa, 1649-028 Portugal

³Antiviral Gene Therapy Research Unit, Department of Molecular Medicine and Haematology, School of Pathology, Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, Gauteng 2193, South Africa

⁴Department of Molecular and Experimental Medicine, The Scripps Research Institute, La Jolla, CA 92037, USA

*Correspondence: yoda@mhlalab.org

<http://dx.doi.org/10.1016/j.cell.2013.09.051>

SUMMARY

Transcription of coregulated genes occurs in the context of long-range chromosomal contacts that

2012). These highly sensitive assays can nascent mRNA and have revealed the FISH foci in a fraction of the population (2010; Papantonis et al., 2010). This sug

Nucleic Acids Research Advance Access published February 4, 2015

Nucleic Acids Research 2015 1
doi: 10.1093/nar/gkv046

Spatial re-organization of myogenic regulatory sequences temporally controls gene expression

Akihito Harada¹, Chandrashekara Mallappa², Seiji Okada¹, John T. Butler², P. Baker^{2,3}, Jeanne B. Lawrence², Yasuyuki Ohkawa^{1,2,*} and Anthony N. Im

¹Department of Advanced Medical Initiatives, JST-CREST, Faculty of Medicine, Kyushu University, 812-8582, Japan, ²Department of Cell and Developmental Biology, University of Massachusetts

21. G. A. Way, *Nat. Rev. Genet.* 8, 206–216 (2007).
22. S. B. Carroll, *Cell* 134, 25–36 (2008).

and K. Paszkiewicz and the Exeter Sequencing Service facility for genome sequencing services. This work was also supported

TRANSCRIPTION

CTCF establishes discrete functional chromatin domains at the *Hox* clusters during differentiation

Varun Narendra,^{1,2} Pedro P. Rocha,³ Disi An,⁴ Ramya Raviram,³ Jane A. Skok,³ Esteban O. Mazzoni,^{4,*} Danny Reinberg^{1,2,*}

Polycomb and Trithorax group proteins encode the epigenetic memory of cellular position identity by establishing inheritable domains of repressive and active chromatin within the *Hox* clusters. Here we demonstrate that the CTCF-binding factor (CTCF) functions



GENOME
RESEARCH

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Next Genera
USA Congre
27 - 28 October 2015

Spatial enhancer clustering and regulation of enhancer-proximal genes by cohesin

Elizabeth Ing-Simmons^{1,2,7}, Vlad C. Seitan^{1,7}, Andre J. Faure^{3,8}, Paul Flicek^{3,4},

Disruptions of Topological Chromatin Domains Cause Pathogenic Rewiring of Gene-Enhancer Interactions

Dario G. Lupiáñez,^{1,2} Katerina Kraft,^{1,2} Verena Heinrich,² Peter Krawitz,^{1,2} Francesco Brancati,³ Eva Kl Denise Horn,² Hülya Kayserili,⁵ John M. Opitz,⁶ Renata Laxova,⁶ Fernando Santos-Simarro,^{7,8} Brigitte Gilbert-Dussardier,⁹ Lars Wittler,¹⁰ Marina Borschiwer,¹ Stefan A. Haas,¹¹ Marco Osterwalder,¹² Bernd Timmermann,¹³ Jochen Hecht,^{1,14} Malte Spielmann,^{1,2,14} Axel Visel,^{12,15,16} and Stefan Mundlos¹

¹Max Planck Institute for Molecular Genetics, RG Development & Disease, 14195 Berlin, Germany

²Institute for Medical and Human Genetics, Charité Universitätsmedizin Berlin, 13353 Berlin, Germany

³Medical Genetics Unit, Policlinico Tor Vergata University Hospital, 00133 Rome, Italy

Nuclear Aggregation of Olfactory Receptor Genes Governs Their Monogenic Expression

E. Josephine Clowney,¹ Mark A. LeGros,^{2,4} Colleen P. Eirene C. Markenshoff-Papadimitriou,³ Markko Myllys, and Stavros Lomvardas^{1,2,3,*}

¹Program in Biomedical Sciences

²Department of Anatomy

³Program in Neurosciences

University of California, San Francisco, San Francisco, CA 9415

⁴Physical Biosciences Division, Lawrence Berkeley National Lab

Leading Edge
Previews

A CRISPR Connection between Chromatin Topology and Genetic Disorders

Bing Ren^{1,*} and Jesse R. Dixon¹

¹Ludwig Institute for Cancer Research, University of California, San Diego, School of Medicine, 9500 Gilman Drive, La Jolla, CA 92093-0653
*Correspondence: bren@ucsd.edu
<http://dx.doi.org/10.1016/j.cell.2015.04.047>

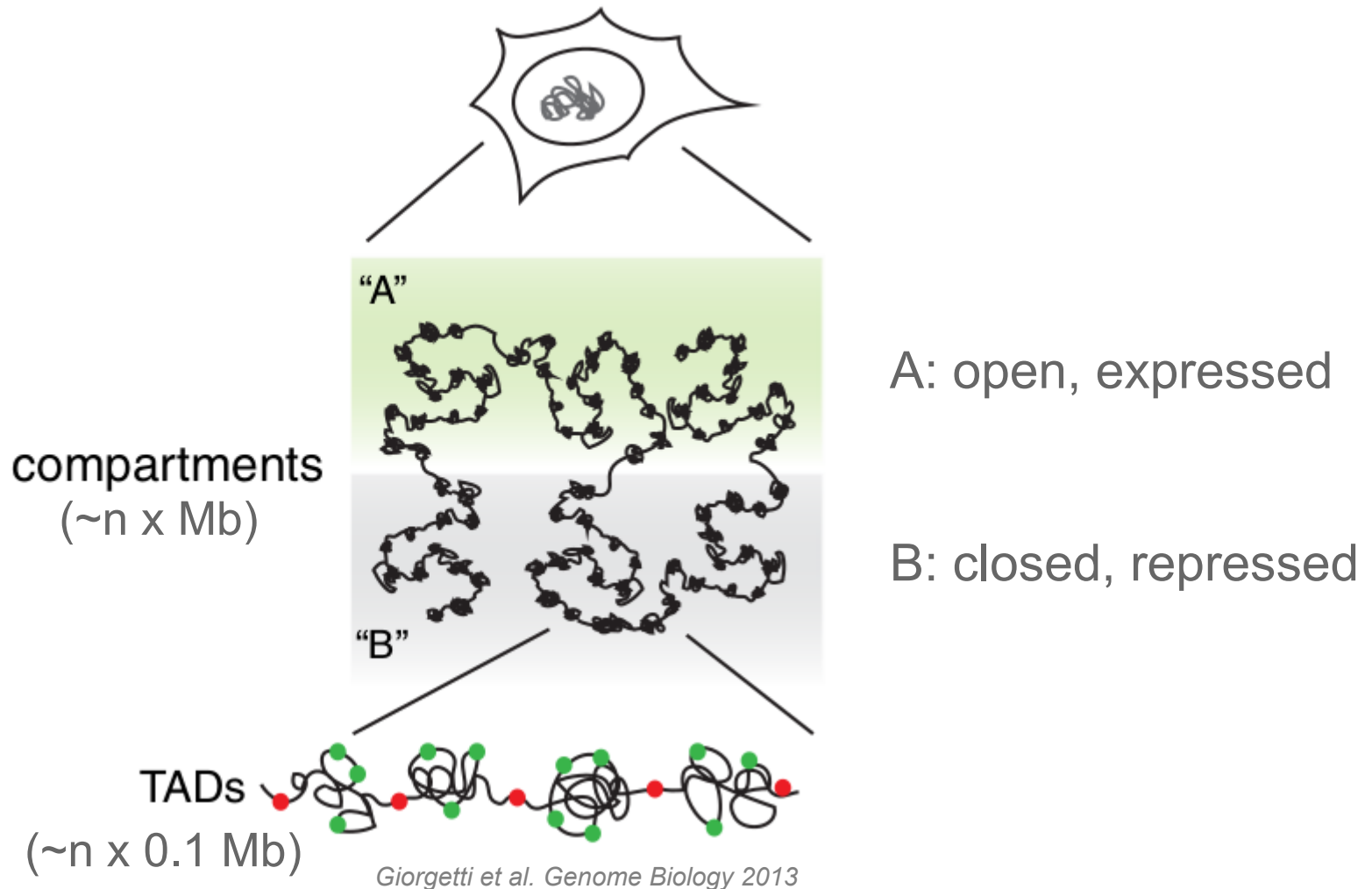
Structural variations are common in the human genome, but their contributions to human diseases have been hard to define. Lupiáñez et al. demonstrate that some structural variants can interrupt chromatin topology, resulting in ectopic enhancer-promoter interactions, altered spatiotemporal gene expression patterns, and developmental disorders.

Structural variations, such as insertion, deletion, duplication, translocation, or inversion of DNA segments, are suggesting that they are stable during development and are not easily disrupted by transcriptional activities of the cell. in the human cases (Figure 1). Remarkably, mutant mice carrying these structural alterations accurately reproduce

Cell

Cell

Intro: from structure to function



Intro: from structure to function

- **How to profile** the 3D structure of the genome in **livestock species**?
- What are the **main features** of this 3D organization?
- Can we **link structure & function**?

The FR-AgENCODE project



*Elisabetta
Giuffra*



*Sylvain
Foissac*

FR-AgENCODE: a French pilot project of the FAANG action
INRA funding: 300KE, SelGen metaprogramme
4 research units, 60+ participants

Goal: improve the functional annotation of animal genomes

The FR-AgENCODE project

Sampling: 40+ tissues

(LIVER, CD4+, CD8+, sperm, plasma, heart, lung, skin, fat, duodenum, ileum, jejunum, cerebellum, frontal lobe, olfactory bulb, trigeminal ganglia, hypothalamus, pancreas, adrenals, kidney, muscle, bone, joints, spleen, lymphatic nodes, peyer's patches, ovary, oocytes, oviduct, uterus, mammary gland, acini, testis, seminal vesicle, etc)



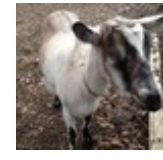
Sus scrofa
(Large White)



Gallus gallus
(White Leghorn)



Bos Taurus
(Holstein)



Capra hircus
(Alpine)



4,115 BioSamples entries available at INRA biorepository

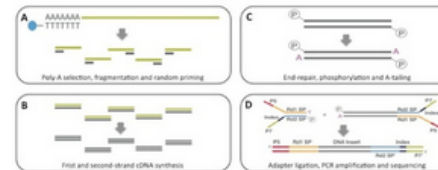


Sequencing assays on 3 target tissues

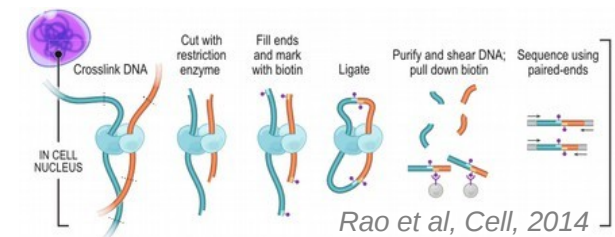
Transcriptome & chromatin structure profiling: polyA+ RNA-seq (130M RP/lib), small RNA-seq (40MR/lib), Hi-C (130M RP/lib), ATAC-seq (50M RP/lib)

~20B reads (3Tb of seq. data)

Illumina Tru-Seq RNA-seq protocol

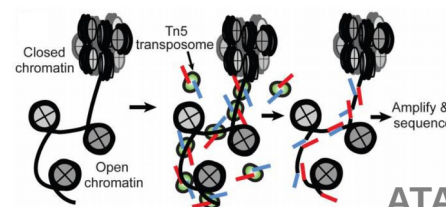


RNA-seq: transcriptome



Rao et al, Cell, 2014

Hi-C: chromosome conformation

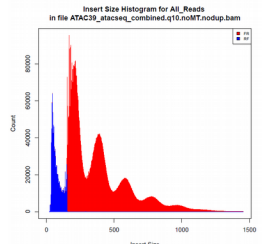
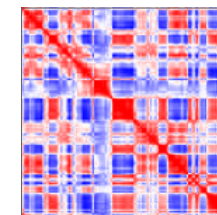
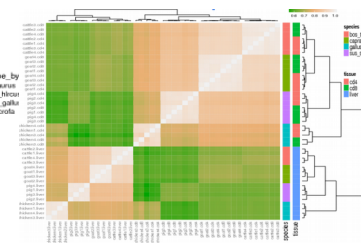
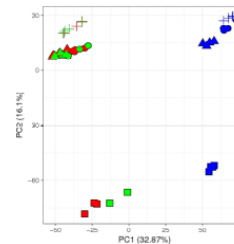


Buenrostro et al, 2013

ATAC-seq: chromatin accessibility

Data analysis

Annotation of genes, transcripts, regulatory regions and topological domains. Comparative and integrative analysis.



FR-AgENCODE data: RNA-seq & ATAC-seq

UCSC Genome Browser on Pig Aug. 2011 (SGSC Sscrofa10.2/susScr3) Assembly

move <<< << < > >> >>> zoom in 1.5x 3x 10x base zoom out 1.5x 3x 10x 100x

chr2:17,145,243-17,328,781 183,539 bp. enter position, gene symbol or search terms

go

chr2

2

Scale

chr2:

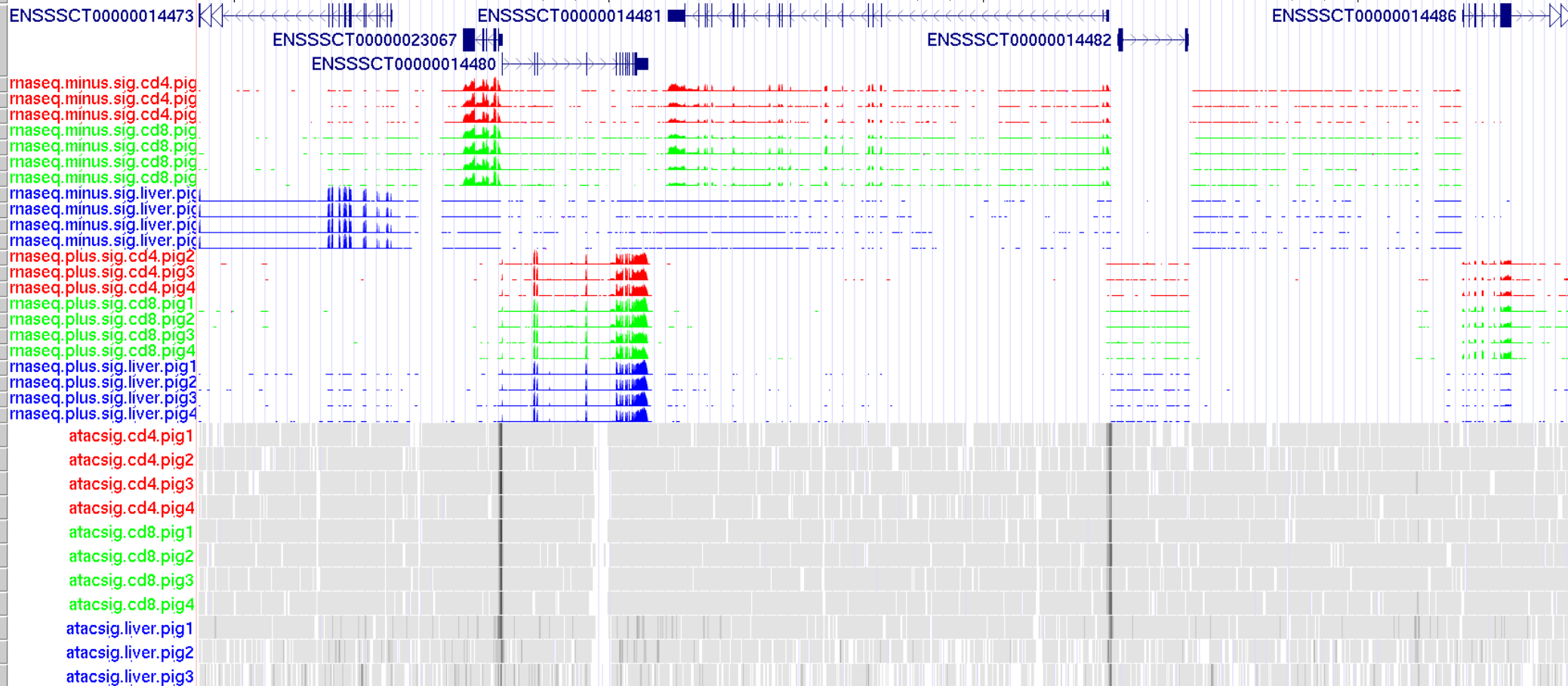
50 kb

susScr3

17,200,000

17,250,000

17,300,000



FR-AgENCODE data: RNA-seq & ATAC-seq

UCSC Genome Browser on Pig Aug. 2011 (SGSC Sscrofa10.2/susScr3) Assembly

move <<< << < > >> >>> zoom in 1.5x 3x 10x base zoom out 1.5x 3x 10x 100x

chr3:102,149,406-102,590,830 441,425 bp. enter position, gene symbol or search terms

go

chr3

3

Scale

chr3:

102,200,000 102,250,000 102,300,000 102,350,000 102,400,000 102,450,000 102,500,000 102,550,000

100 kb

susScr3

ENSSSCT00000009249

ENSSSCT00000009253

ENSSSCT000000021973

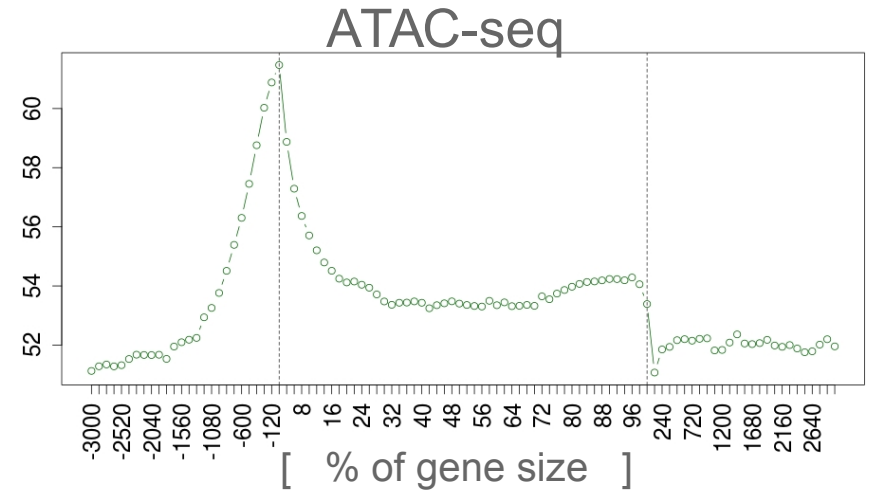
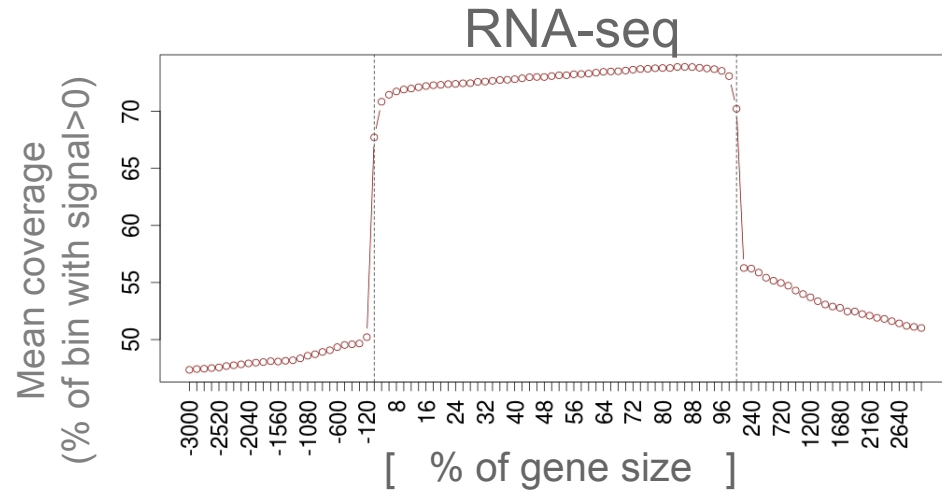
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ENSSSCT00000009251

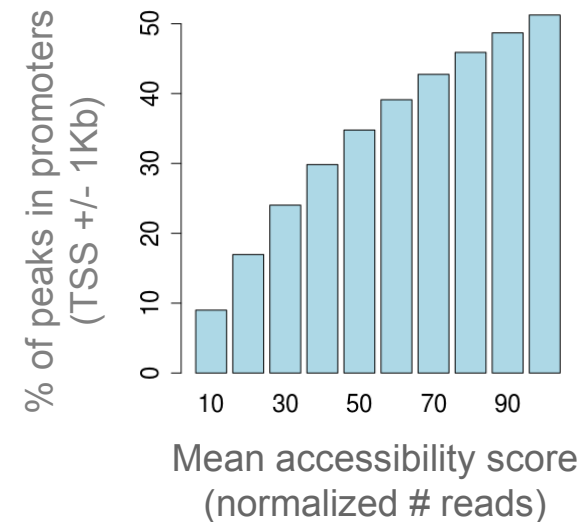
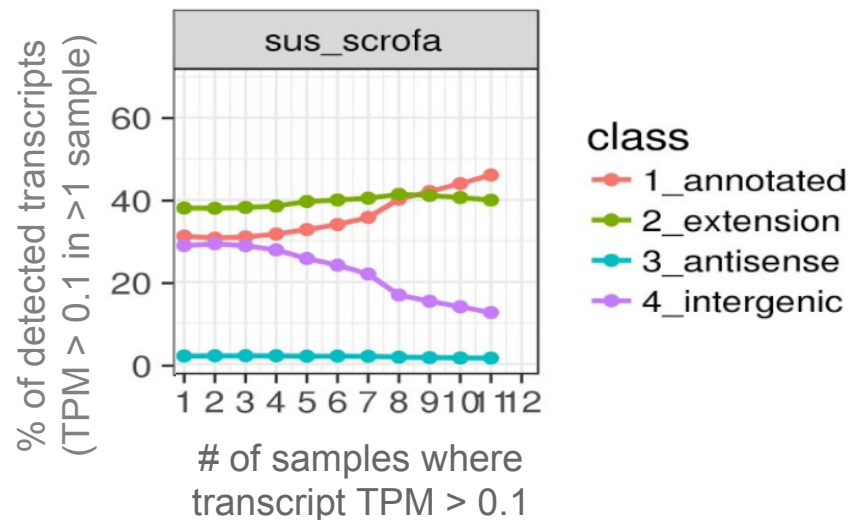
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rnaseq.minus.sig.liver.pig
rnaseq.plus.sig.cd4.pig2
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atcsg.cd8.pig1
atcsg.cd8.pig2
atcsg.cd8.pig3
atcsg.cd8.pig4
atcsg.liver.pig1
atcsg.liver.pig2
atcsg.liver.pig3

FR-AgENCODE results: RNA-seq & ATAC-seq



Relative position to annotated pig genes in bp (+/-3Kb)



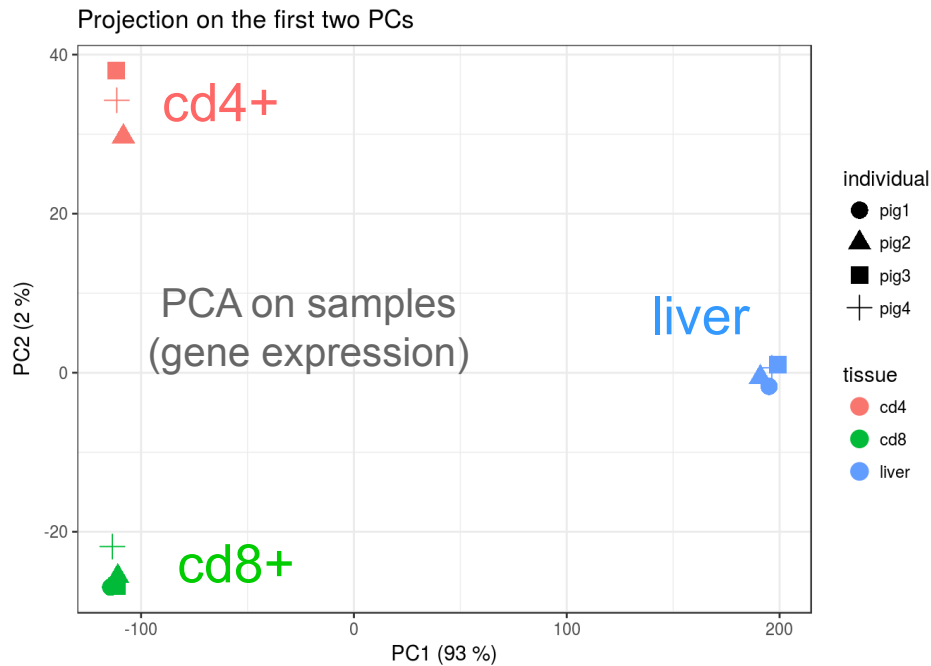
Annotated transcripts are more expressed than novel ones

ATAC-seq reads and peaks accumulate in promoters

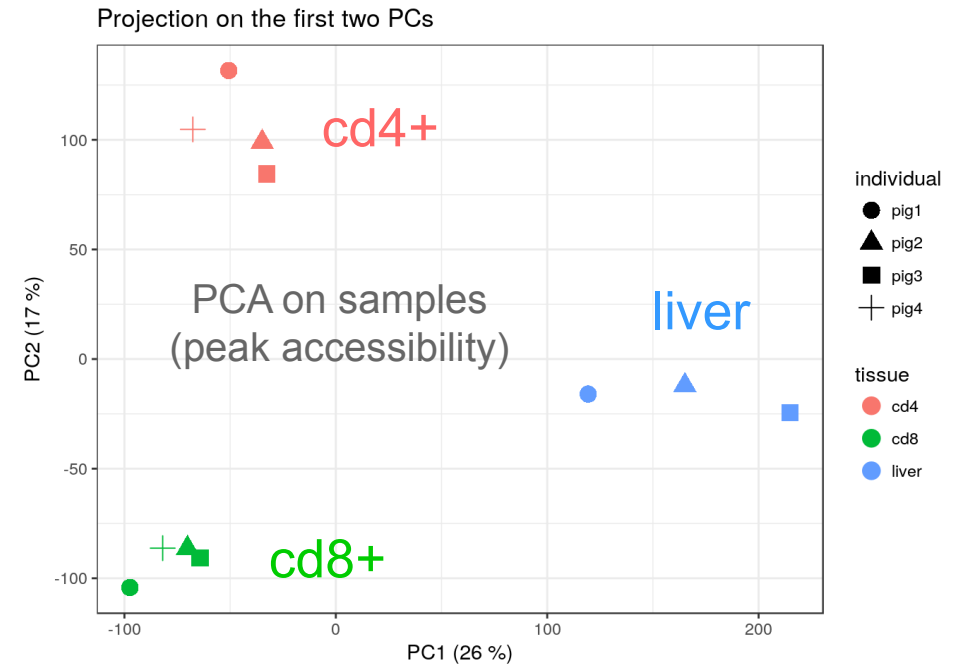
FR-AgENCODE results: RNA-seq & ATAC-seq

Species	Genome / Gene annotation	Annotated Transcripts			Number of novel transcripts detected*	Number of novel lncRNAs
		Total number	Detected*			
			#	% of total		
Bos taurus	UMD 3.1 / Ensembl 84	26,740	16,100	60.2	65,538	7,929
Capra hircus	CHIR_ARS 1 / NCBI	53,266	34,442	64.7	38,197	NA
Gallus gallus	GalGal 5 / Ensembl 87	38,118	22,898	60.1	41,116	3,264
Sus scrofa	SScrofa 10.2 / Ensembl 84	30,585	18,746	61.3	57,978	6,581
* with TPM ≥ 0.1 in ≥ 2 samples						

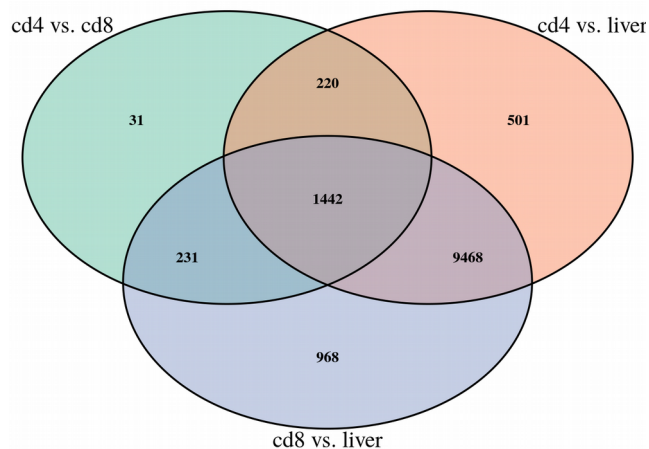
FR-AgENCODE results: RNA-seq & ATAC-seq



Filtering: TPM>0.1 in min 2 samples
Normalization: TMM (edgeR)
N=15,928



Filtering: none
Normalization: LOESS (csaw)
N=120,914



**Differential
gene
expression
(*S. scrofa*)**

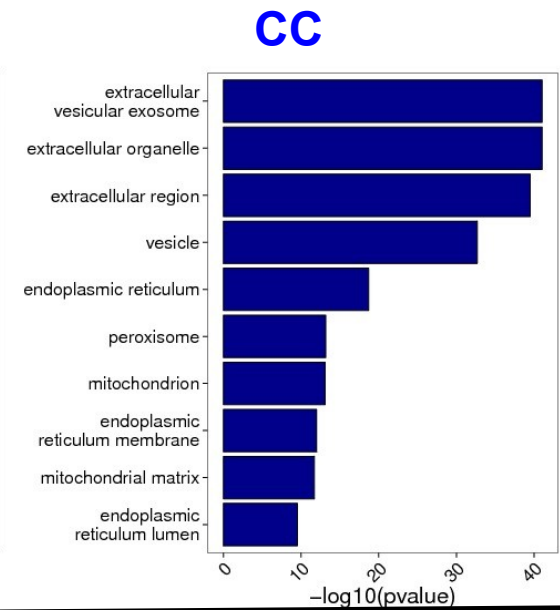
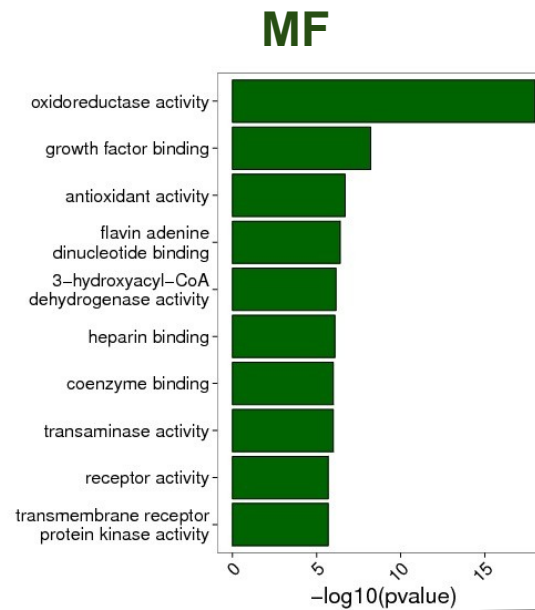
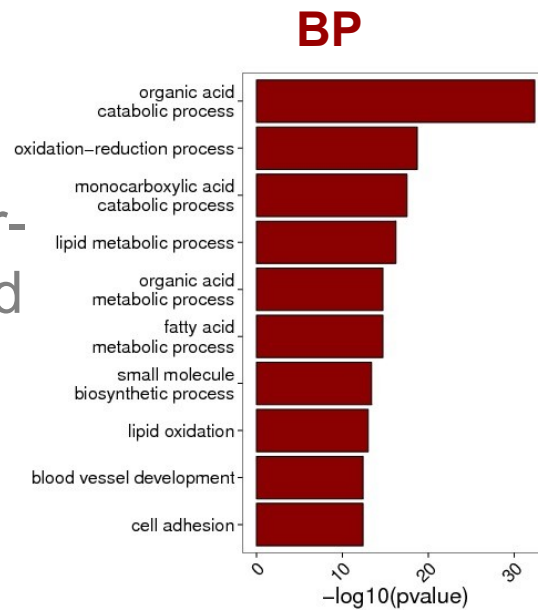
Top- significant GO terms (Biol. Proc.) for
over-expressed genes in liver vs. T cells

oxoacid metabolic process	carboxylic acid biosynthetic process
small molecule catabolic process	carboxylic acid catabolic process
oxidation-reduction process	alpha-amino acid catabolic process
small molecule biosynthetic process	lipid metabolic process

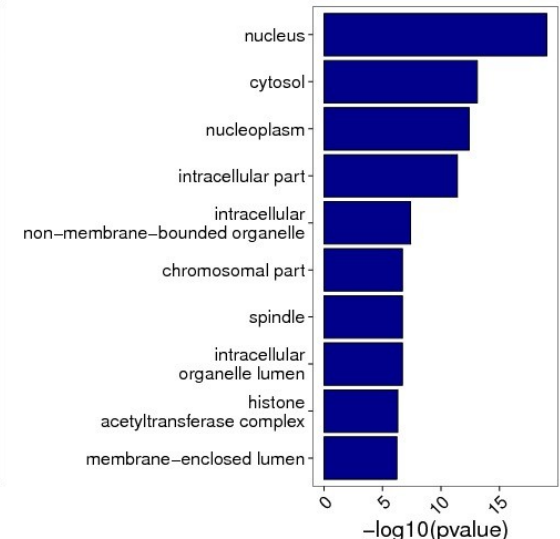
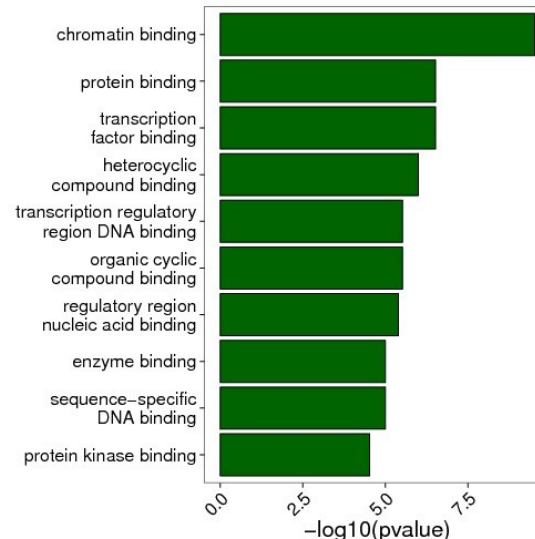
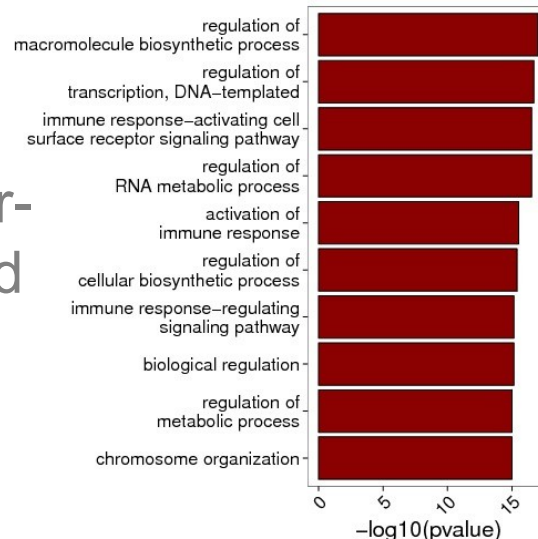
FR-AgENCODE results: RNA-seq & ATAC-seq

Gene Ontology analysis

Liver over-expressed genes

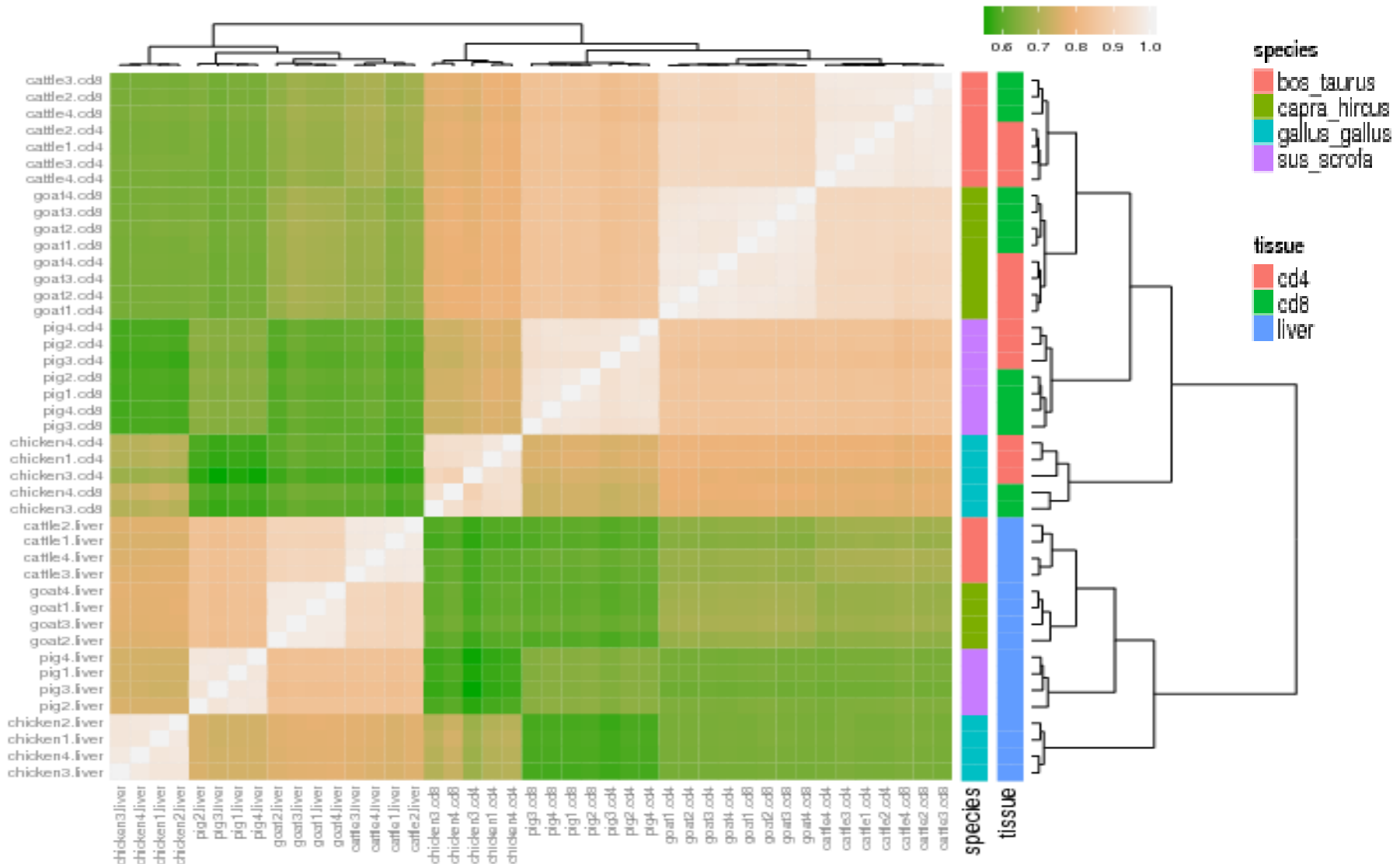


T cell over-expressed genes



FR-AgENCORE results: RNA-seq & ATAC-seq

Expression of orthologous genes: multi-species clustering

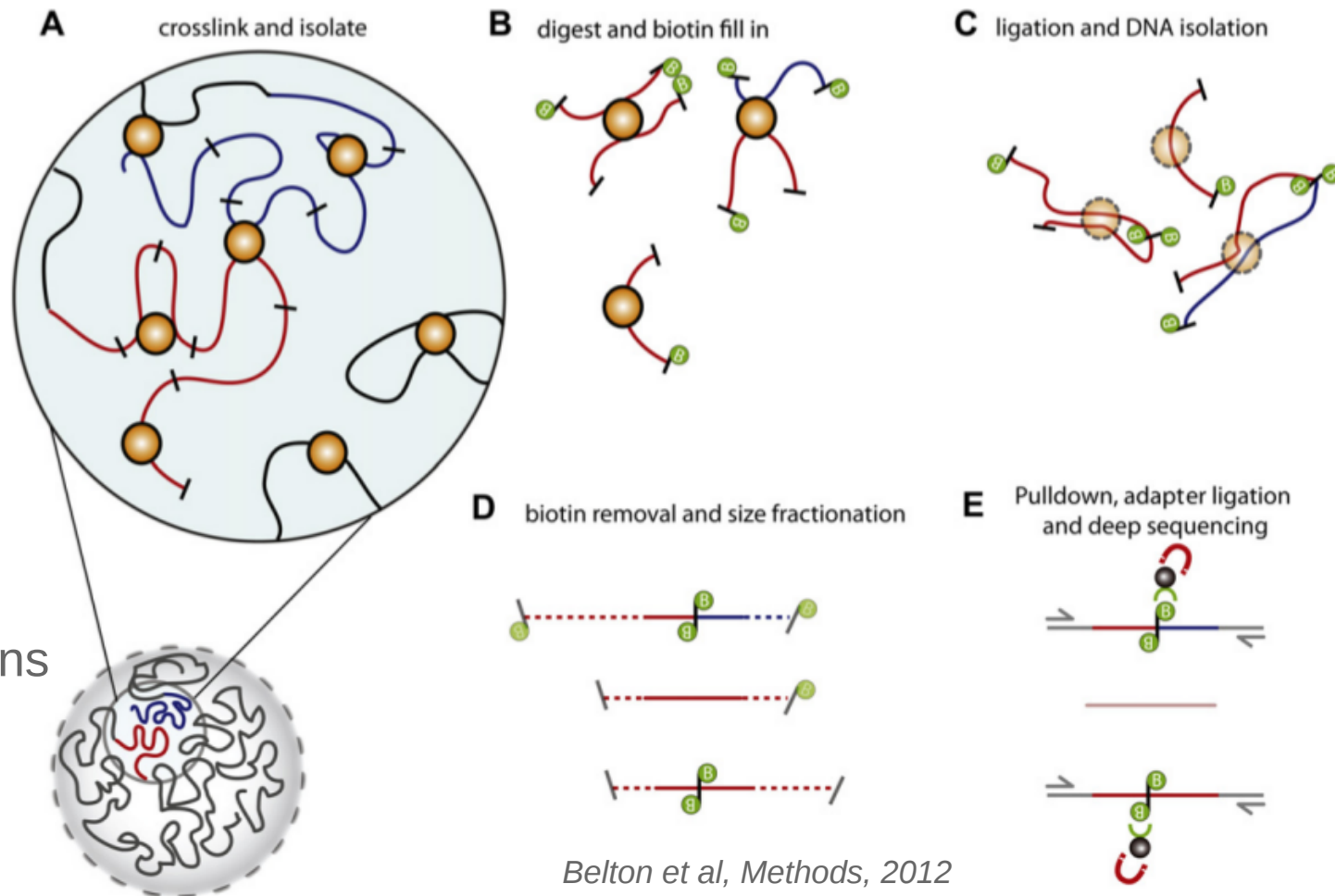


Hi-C : the protocol

Hi-C: chromosome conformation capture by high-throughput sequencing

(Lieberman-Aiden et al, Science, 2009, Rao et al, Cell, 2014)

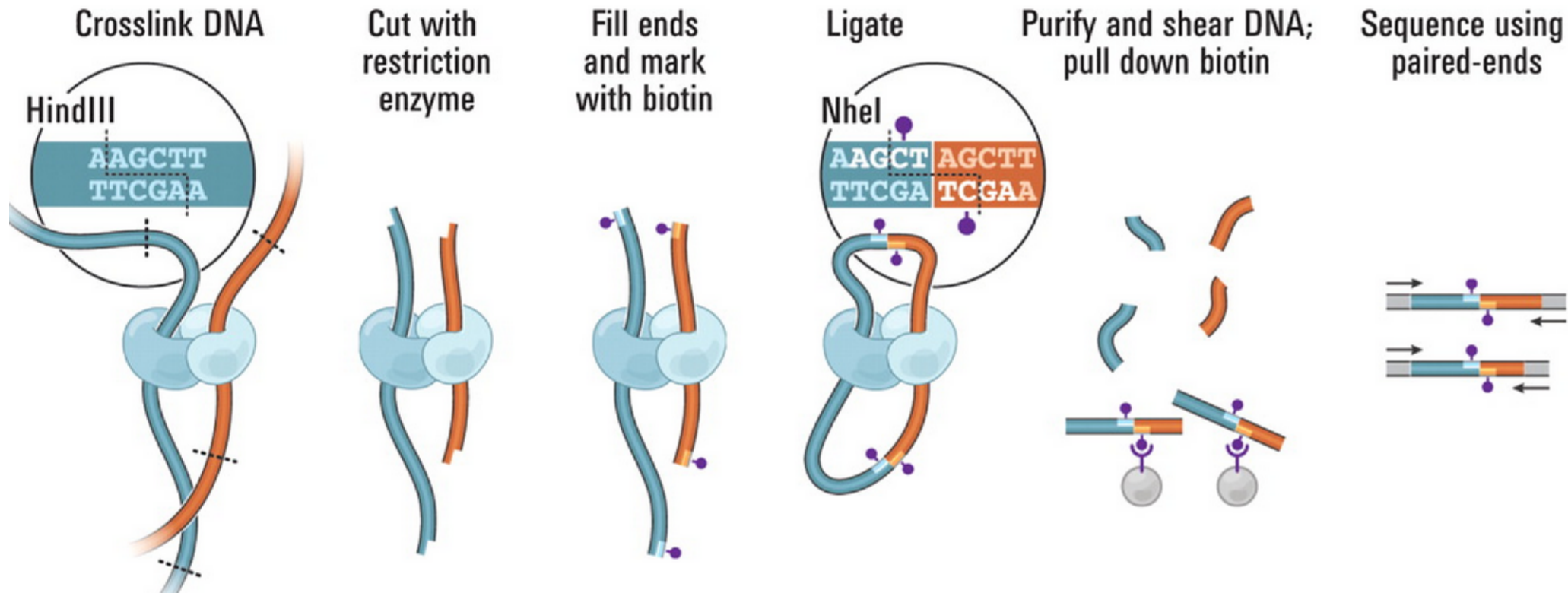
- crosslink DNA (“fixation”)
- cleave genome with restriction enzyme
- biotin-mark and ligate extremities
- fragment, select biotin-marked junctions
- sequence fragments (paired-ends)



Hi-C : the protocol

Hi-C: chromosome conformation capture by high-throughput sequencing

(Lieberman-Aiden et al, Science, 2009, Rao et al, Cell, 2014)



Rao et al, Cell, 2014

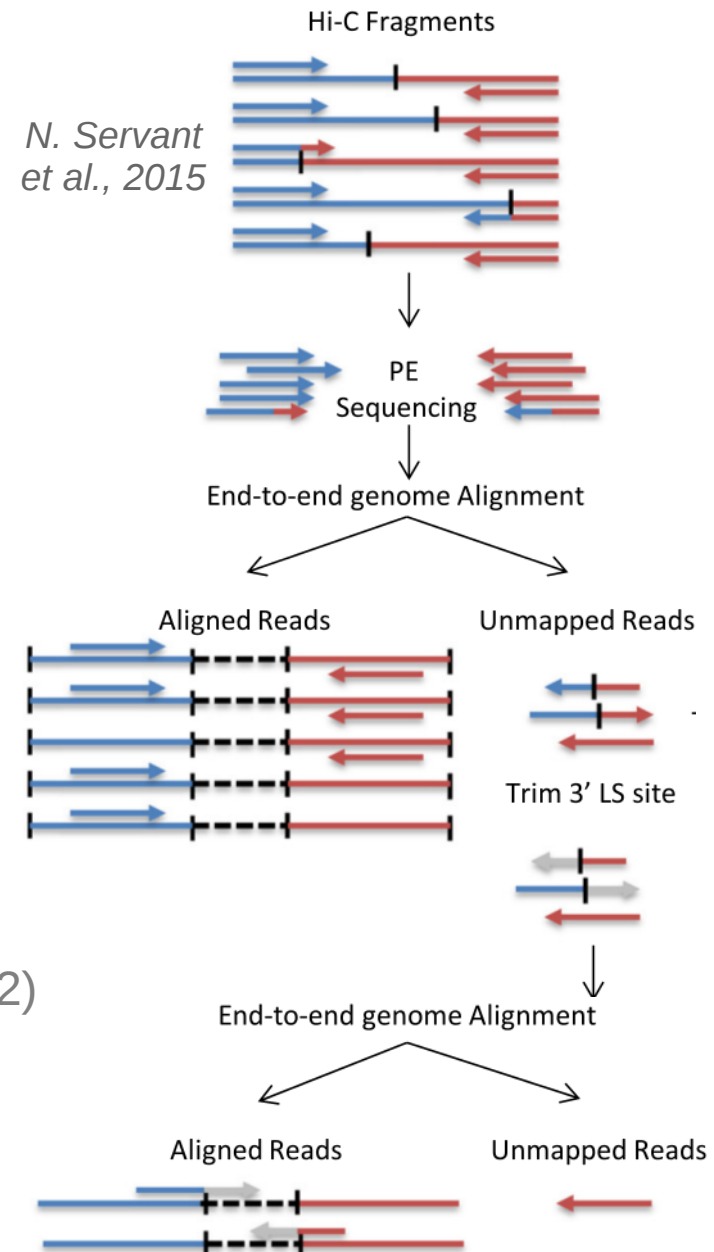
Hi-C : the analysis pipeline

FR-AgENCODE pipeline

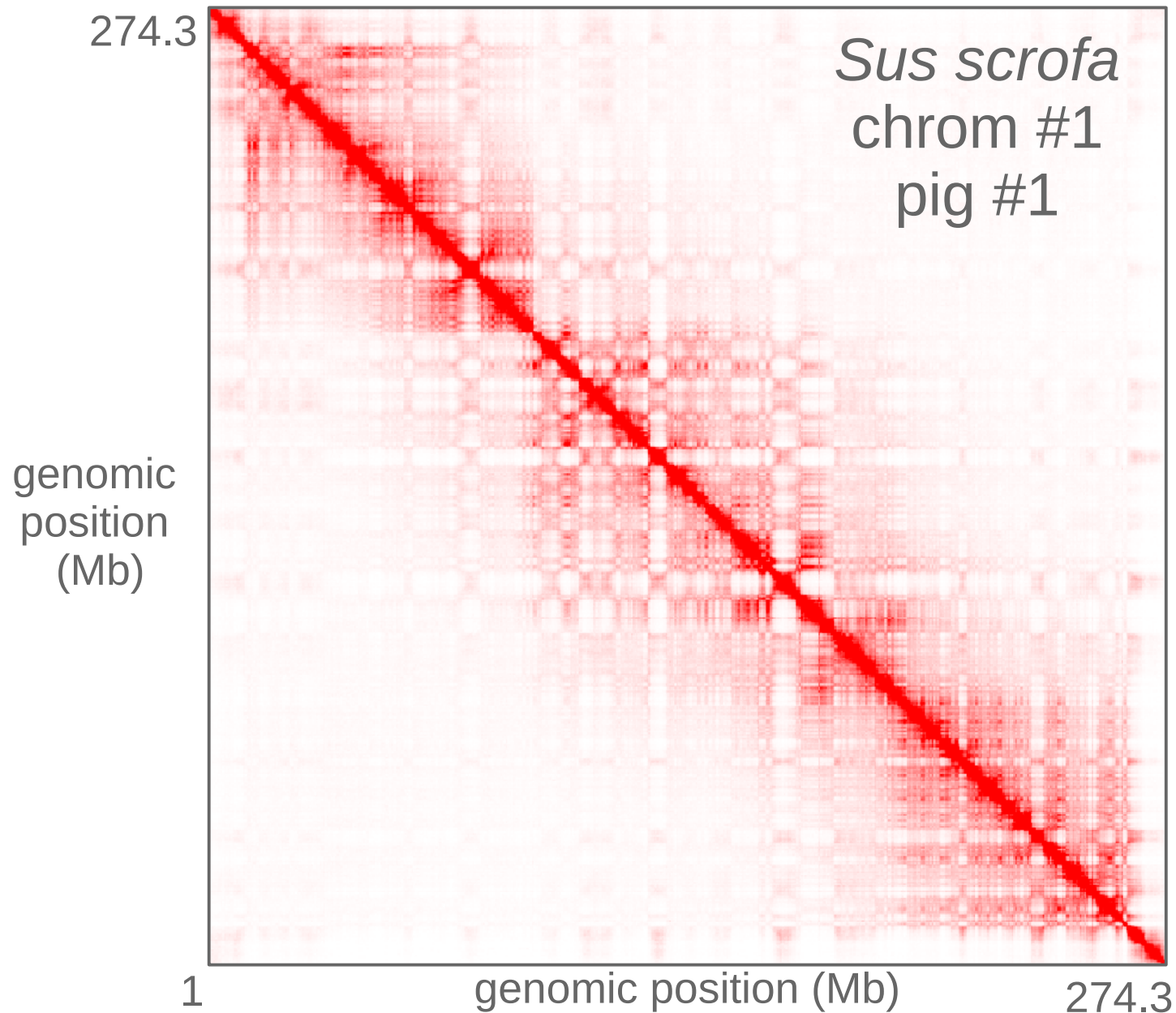
- Trim reads (ligation site)
- **Map on reference genome**
- Discard inconsistent pairs
- **Count reads in pairs of genomic bins & generate contact matrix**
- Normalize contact matrix (non parametric, matrix balancing)
- Generate html report
- **Identify Topological Associated Domains**, *cis* and *trans* interactions
- **Identify A and B compartments**

Software

- HiC-Pro pipeline (Servant et al 2015)
- Bowtie2 mapping (Langmead et al, 2009)
- ICE normalization (Imakaev et al, 2012)
- HiTC display and A/B comp. (Servant et al, 2012)
- HiFive pipeline (Sauria et al, 2015)
- Armatus TAD finding (Filippova et al, 2014)
- Juicebox browser (Durand et al, 2016)



Hi-C results : the interaction matrix



Hi-C results : the interaction matrix

pig1

pig2

Sus scrofa
chr1

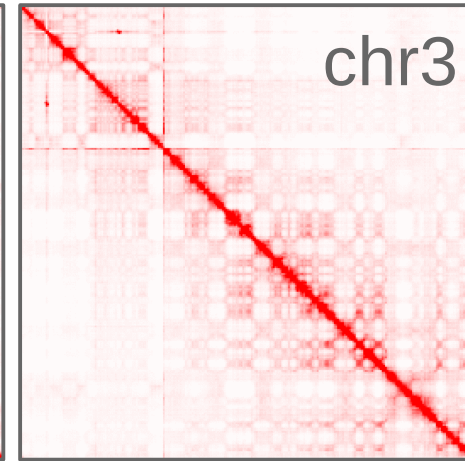
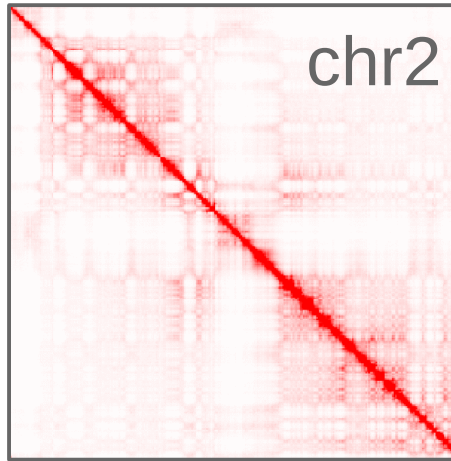
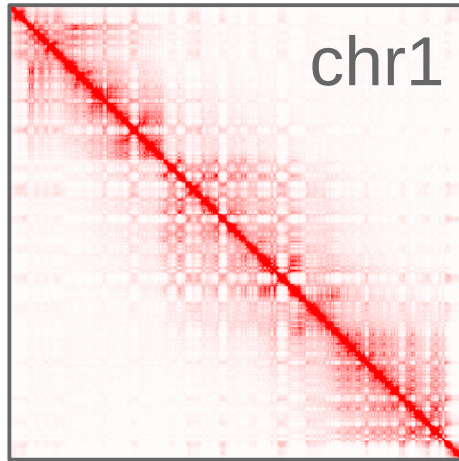
merged

pig3

pig4

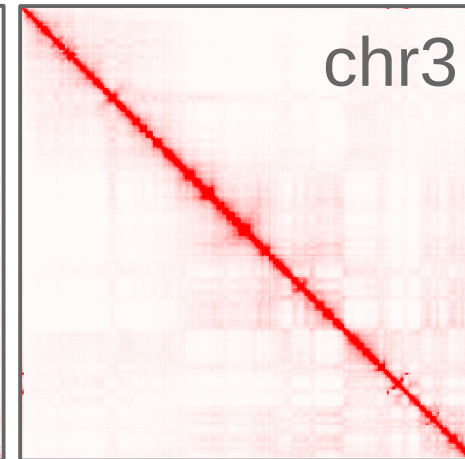
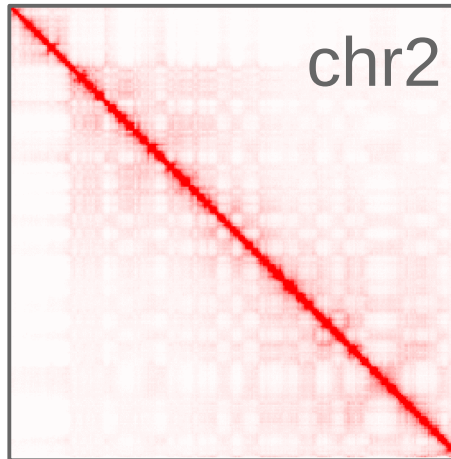
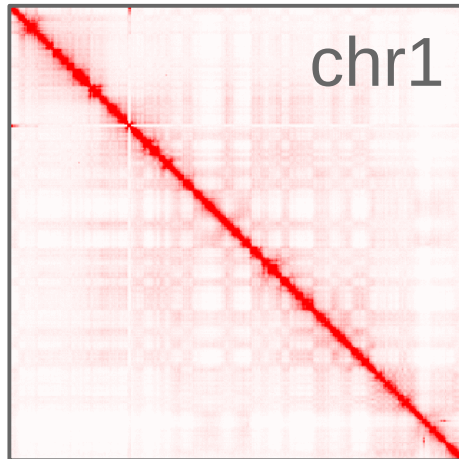
Hi-C results : the interaction matrix

Pig



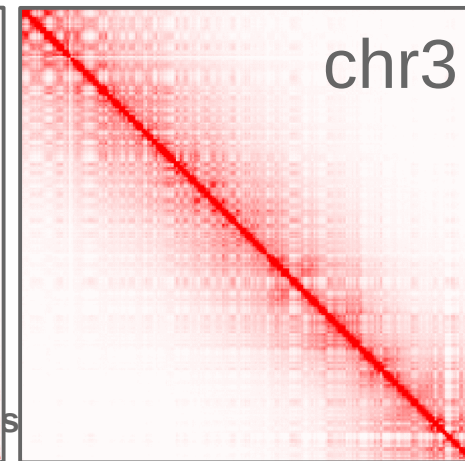
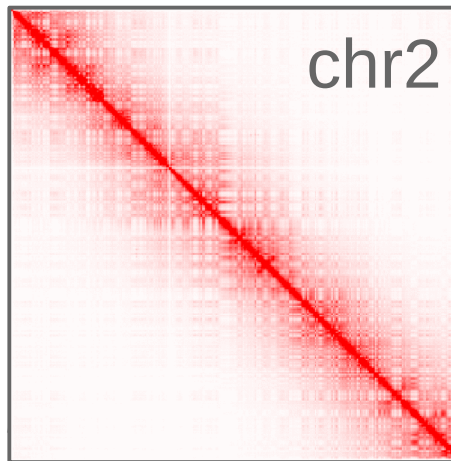
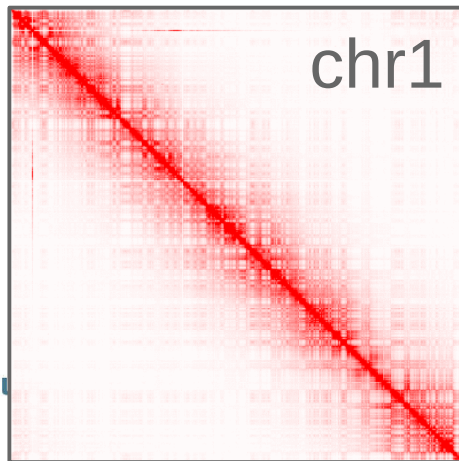
...

Goat



...

Chicken



...

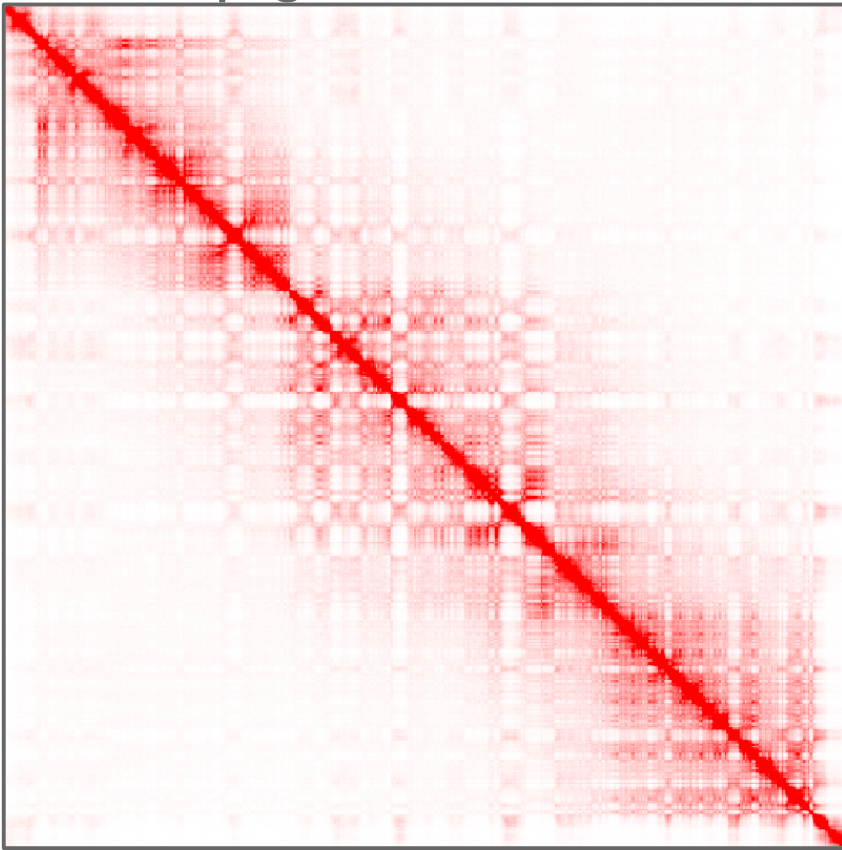
Hi-C results : finding A/B compartments

- Normalization 1: matrix balancing (GC%, repeats, RS density, etc)
- Normalization 2: observed/expected counts (genomic distance)
- Pearson correlation matrix
- Principal Component Analysis => 1st PC sign

Hi-C results : finding A/B compartments

- Normalization 1: “ICE” matrix balancing (GC%, repeats, RS density, etc)

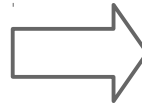
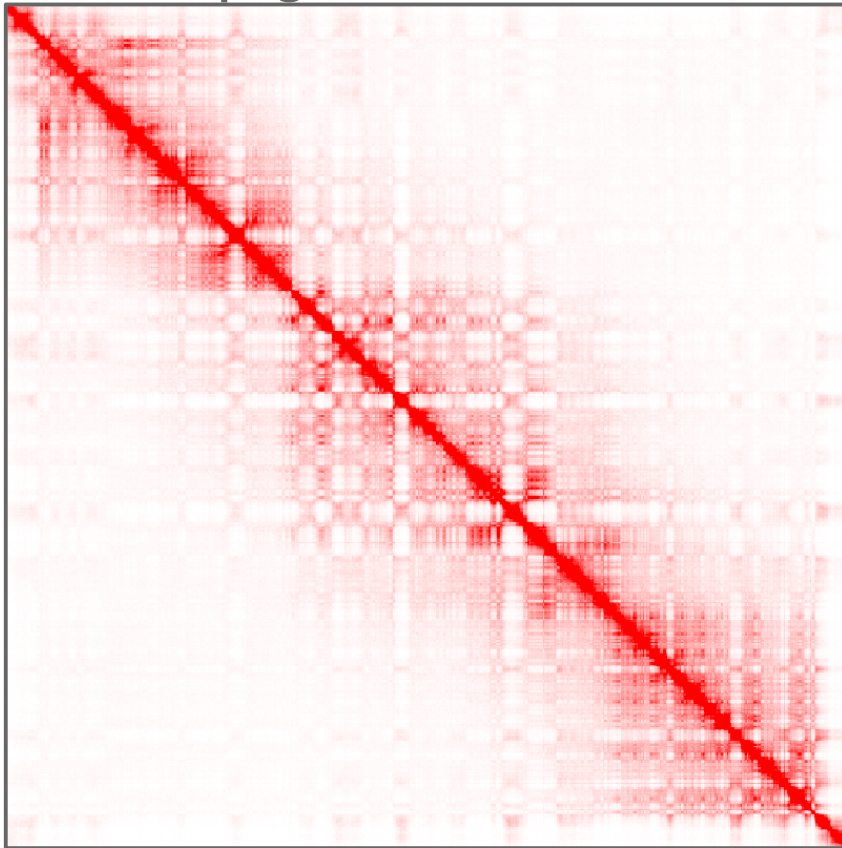
pig, chr1, raw



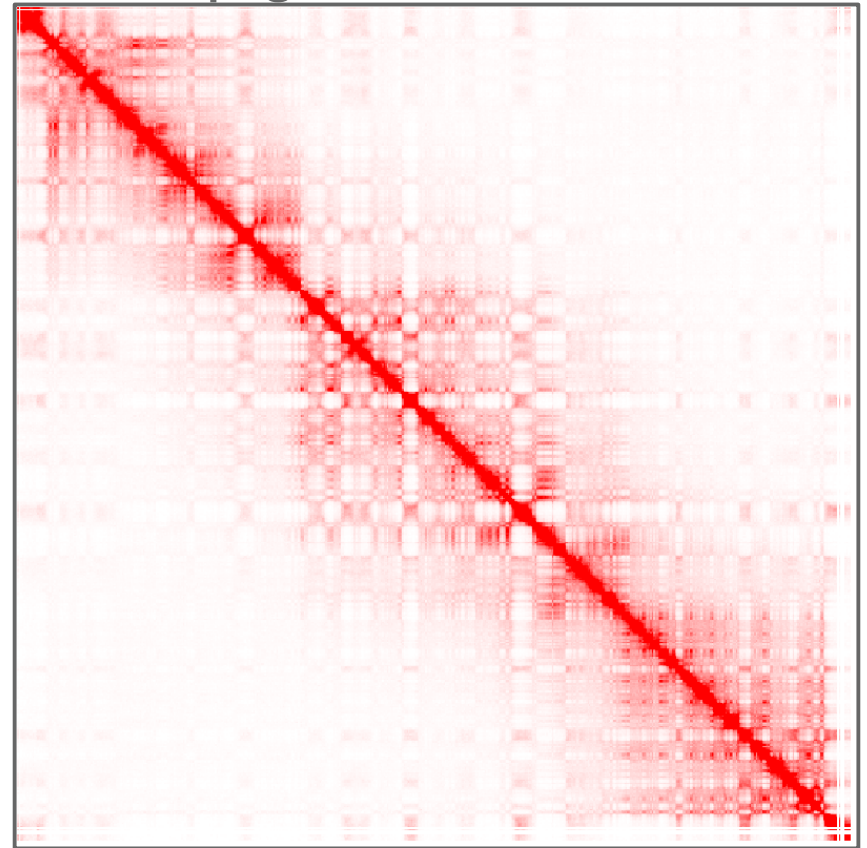
Hi-C results : finding A/B compartments

- Normalization 1: “ICE” matrix balancing (GC%, repeats, RE density, etc)

pig, chr1, raw



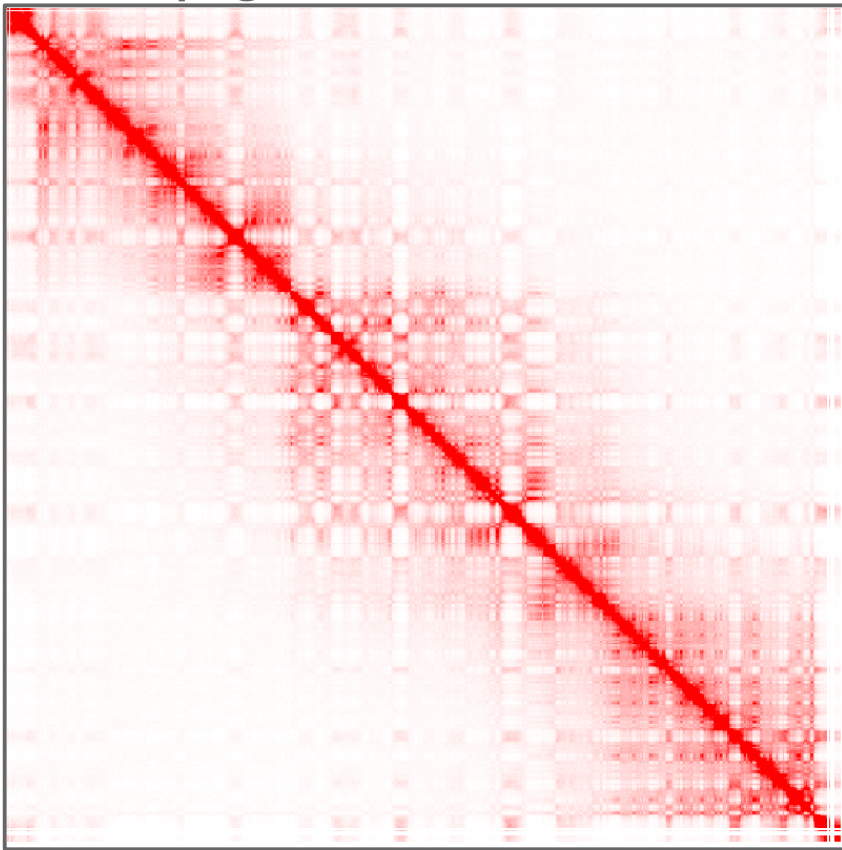
pig, chr1, norm1



Hi-C results : finding A/B compartments

- Normalization 2: observed/expected counts (genomic distance)

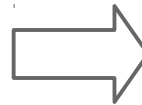
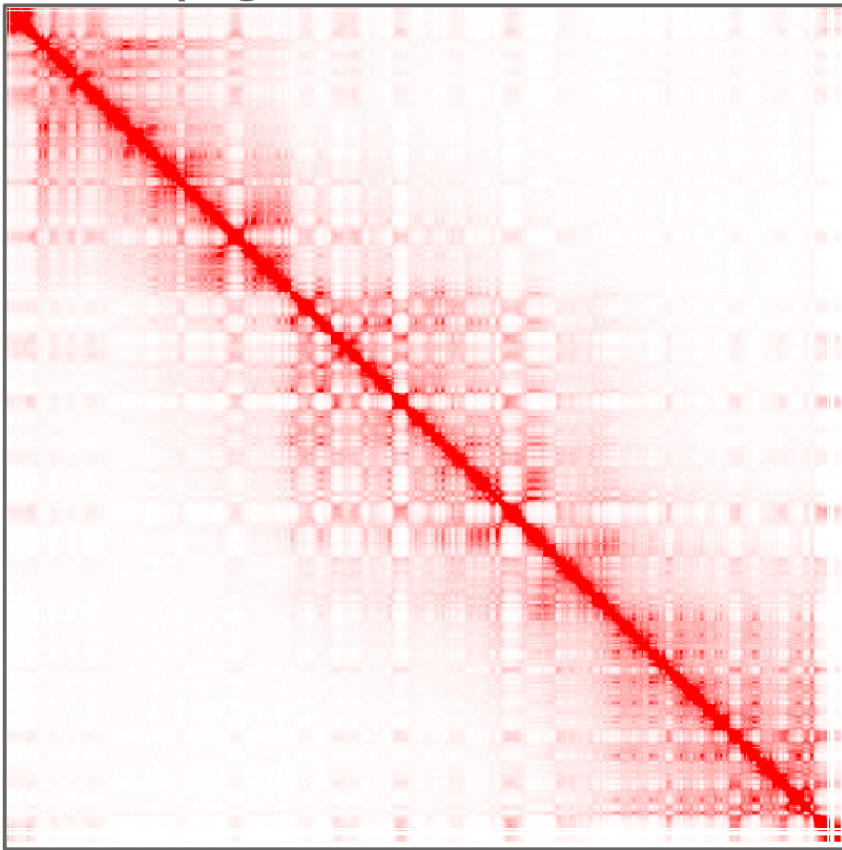
pig, chr1, norm1



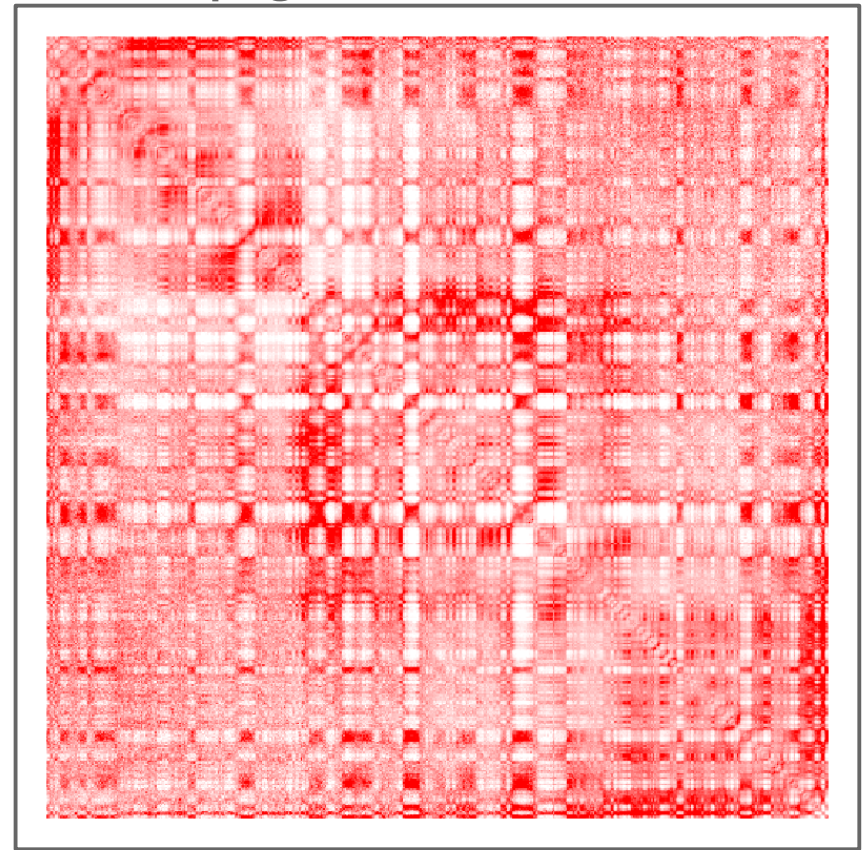
Hi-C results : finding A/B compartments

- Normalization 2: observed/expected counts (genomic distance)

pig, chr1, norm1



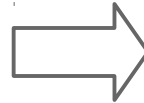
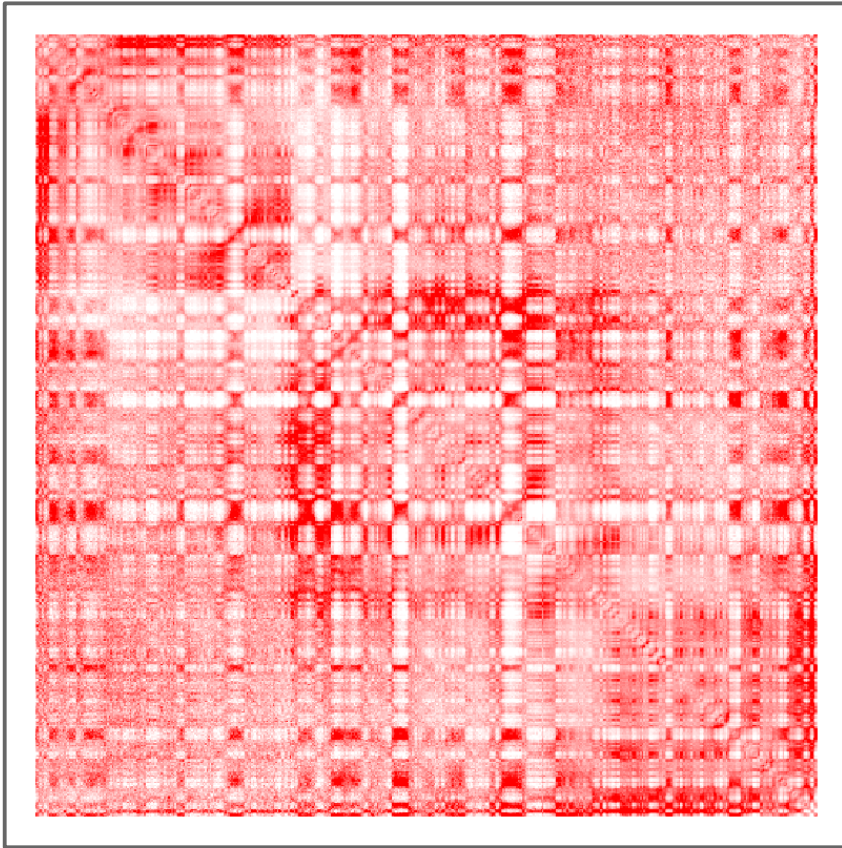
pig, chr1, norm2



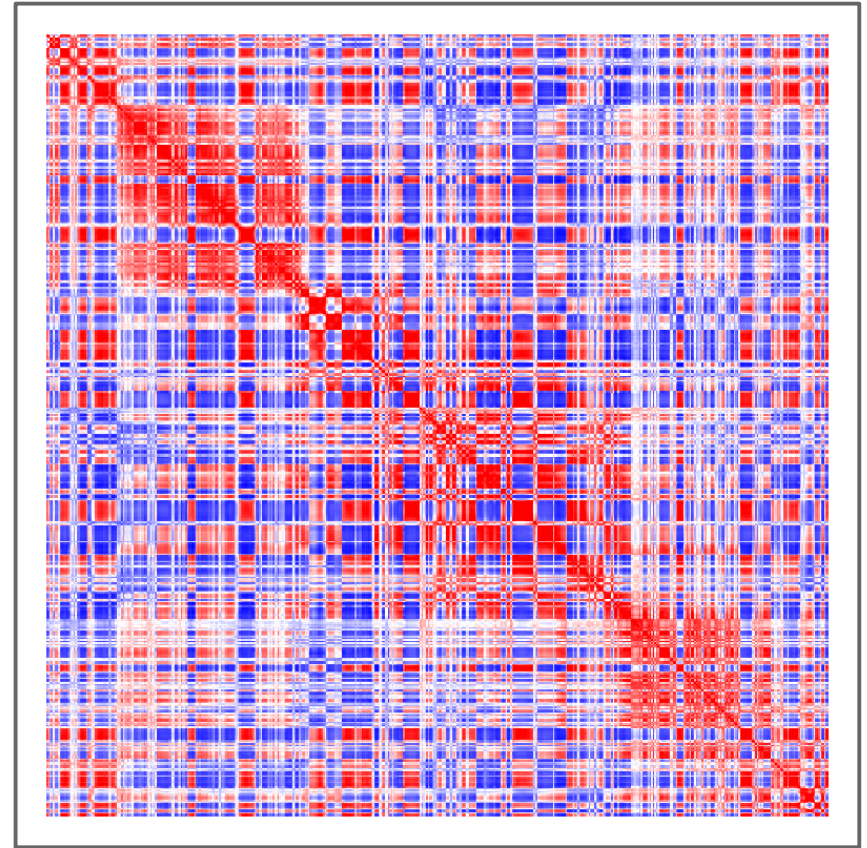
Hi-C results : finding A/B compartments

- Pearson correlation matrix

pig, chr1, norm2



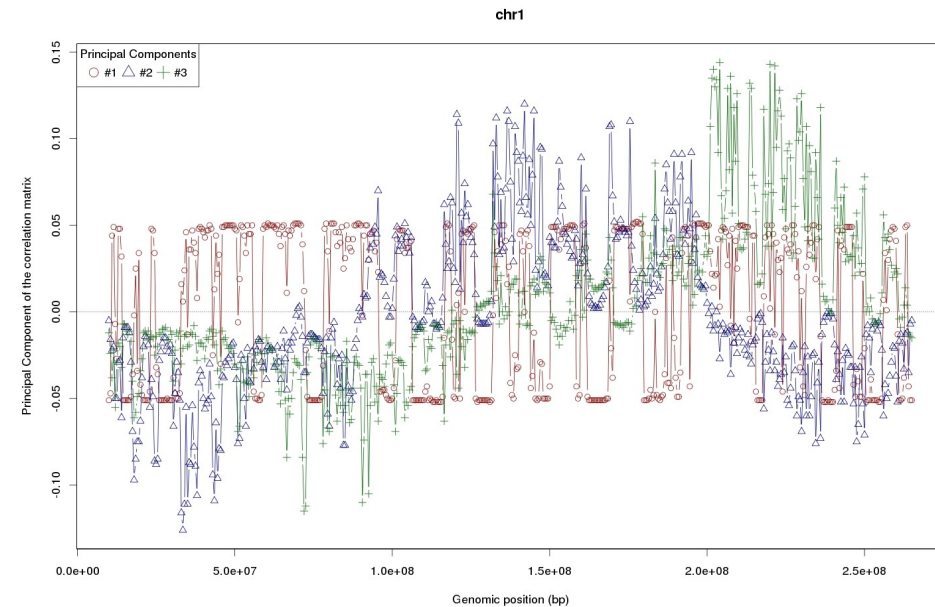
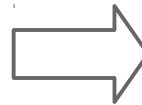
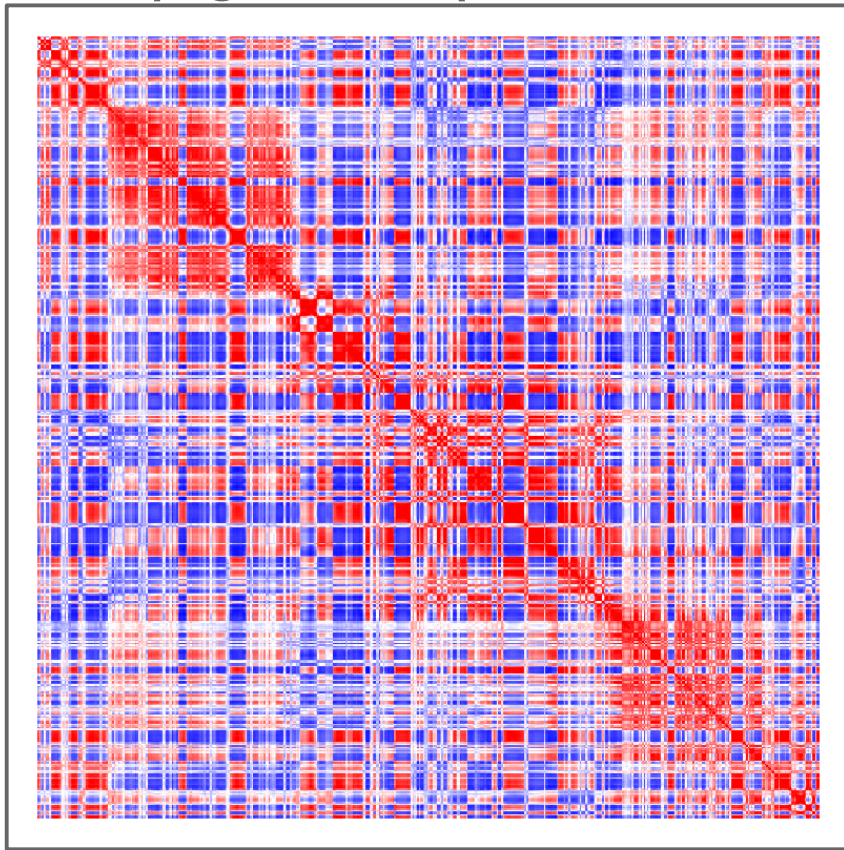
pig, chr1, pearson



Hi-C results : finding A/B compartments

- Principal Component Analysis => 1st PC sign

pig, chr1, pearson



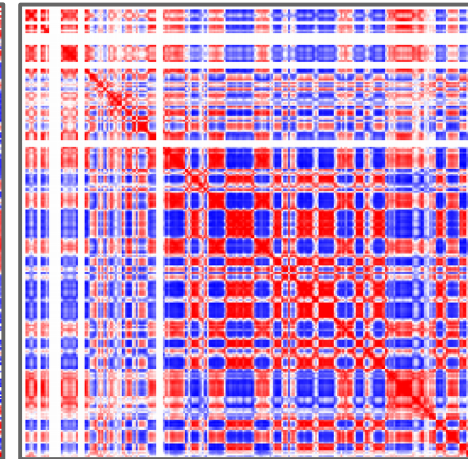
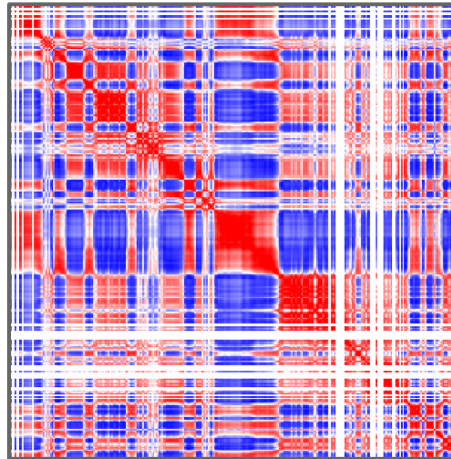
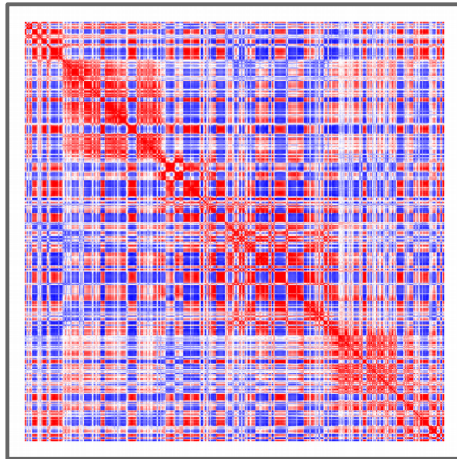
Hi-C results : finding A/B compartments

chr1

chr2

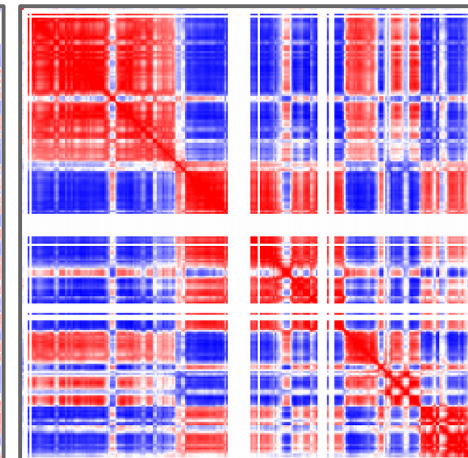
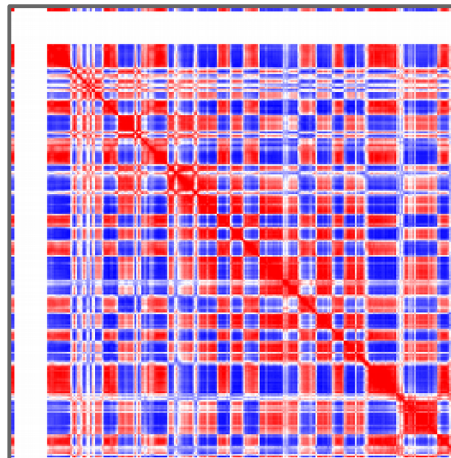
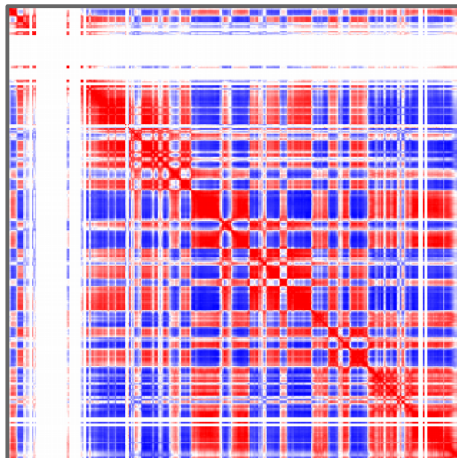
chr3

Pig



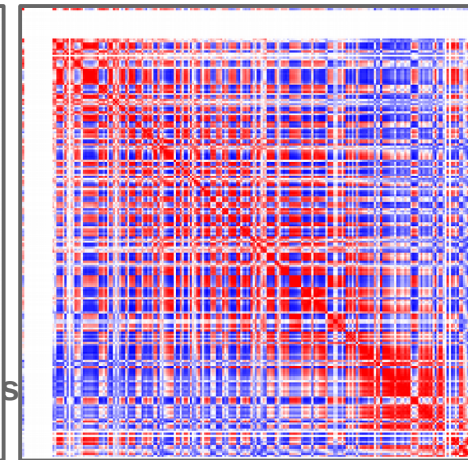
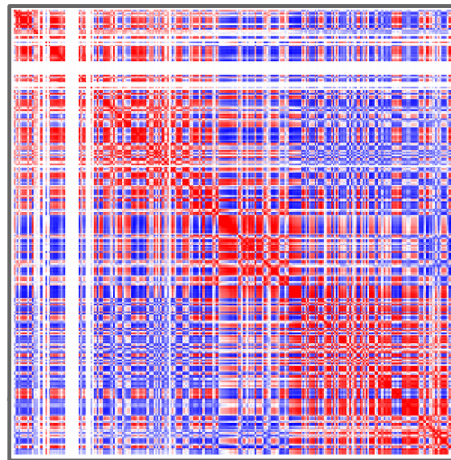
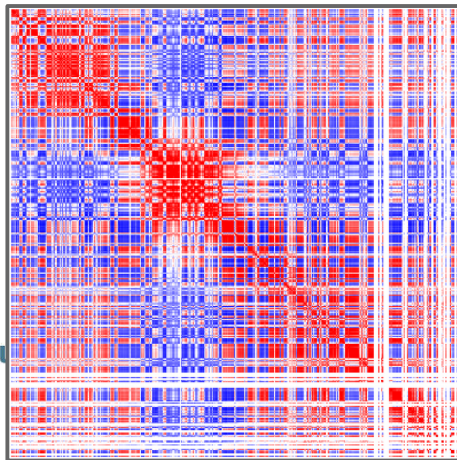
...

Goat



...

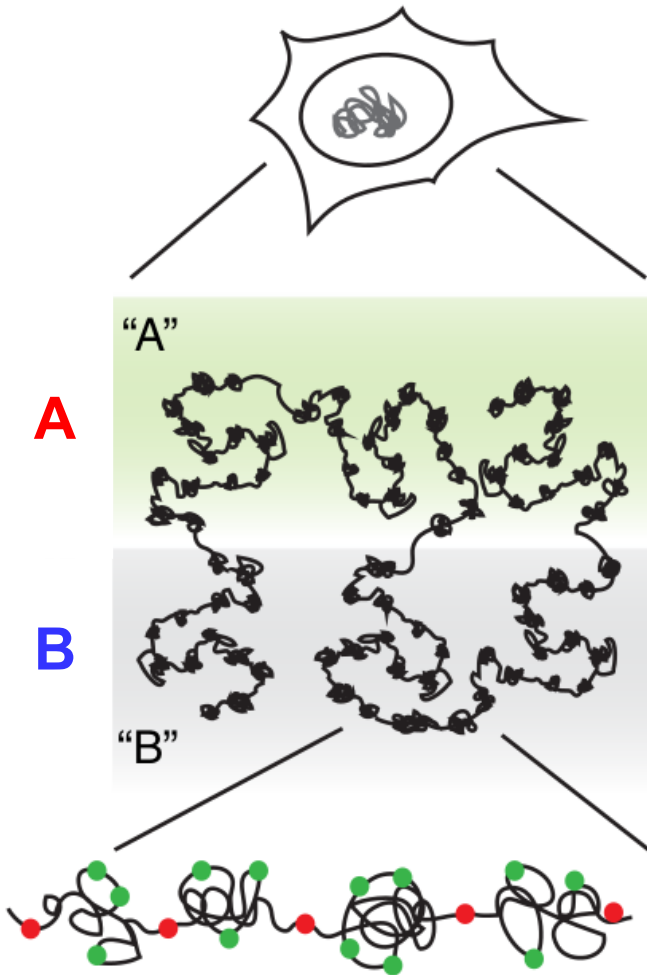
Chicken



...

Results consistency: Hi-C + RNA-seq + ATAC-seq

3D nuclear organization & features

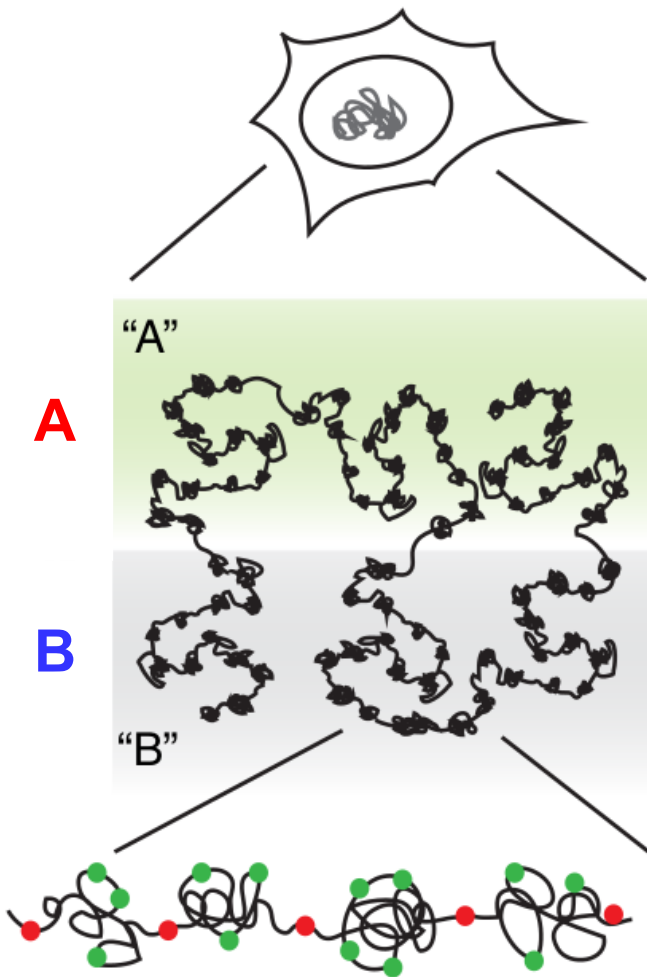


Compartments

- **A**: open, expressed
- **B**: closed, repressed

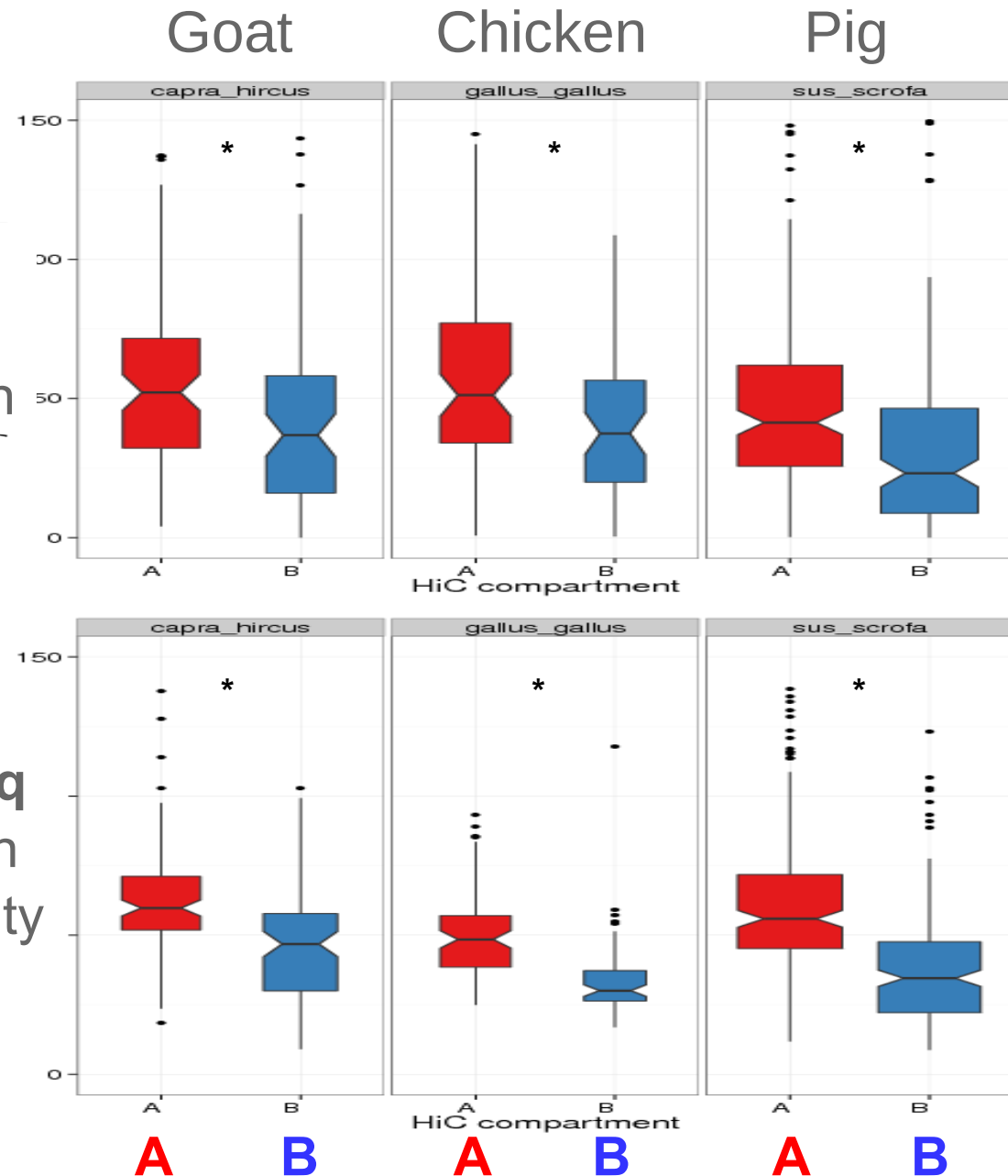
?

Results consistency: Hi-C + RNA-seq + ATAC-seq



RNA-seq
gene
expression

ATAC-seq
chromatin
accessibility



Conclusion

- Livestock genomes need a better annotation
- The 3D organization of the genome can be profiled with Hi-C from tissue samples in livestock species
- Chromosomes are structured in A/B compartments
- Hi-C results are consistent with RNA-seq and ATAC-seq data: compartments A > B for gene expression and chromatin accessibility
- The FR-AgENCODE project is a valuable contribution to the functional annotation effort of the FAANG consortium

Acknowledgements

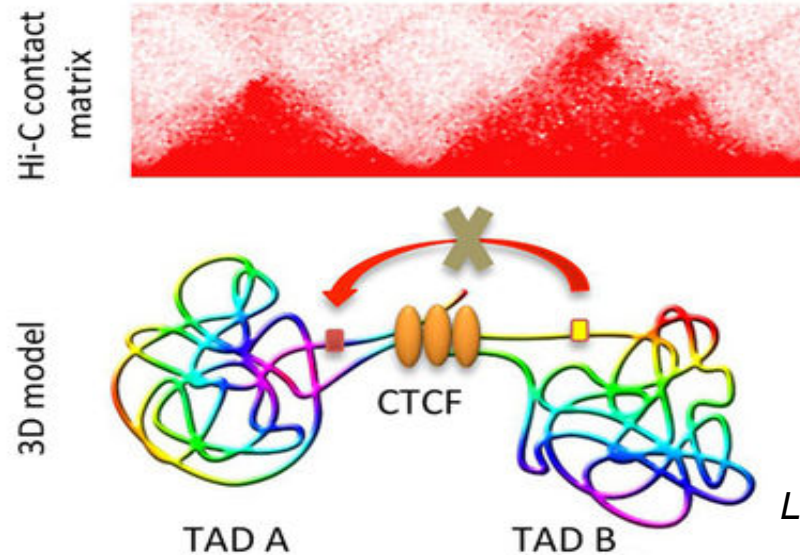
FR-AgENCODE

Acloque H.	Laurent F.
Bardou P.	Marthey S.
Blanc F.	Marti M.
Cabau C.	Mompart F.
Crisci E.	Munyard K.
Derrien T.	Muret K.
Djebali S.	Pinard-Van der Laan MH.
Drouet F.	Pollet S.
Esquerre D.	Queré P.
Fabre S.	Rau A.
Foissac, S	Robelin D.
Gaspin C.	San Cristobal M.
Giuffra E.	Tixier-Boichard M.
Gonzalez I.	Tosser-Klopp G.
Goubil A.	Villa-Vialaneix N.
Klopp C.	Vincent-Naulleau S.
Lagarrigue S.	Zytnicki M. [...]

Hi-C @ Toulouse

Maria Marti
Martine Yerle
Yvette Lahbib
Florence Mompart
Laurence Liaubet
Nathalie Villa-Vialaneix
Pierre Neuvial
Alain Vignal
Matthias Zytnicki
David Robelin
Hervé Acloque
Sarah Djebali

Predicted CTCF binding sites peak at Topologically Associating Domain (TAD) boundaries



Li et al, Scientific Reports, 2016

