

Analyse de communautés bactériennes complexes: du 454 au MiSeq

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Géraldine PASCAL

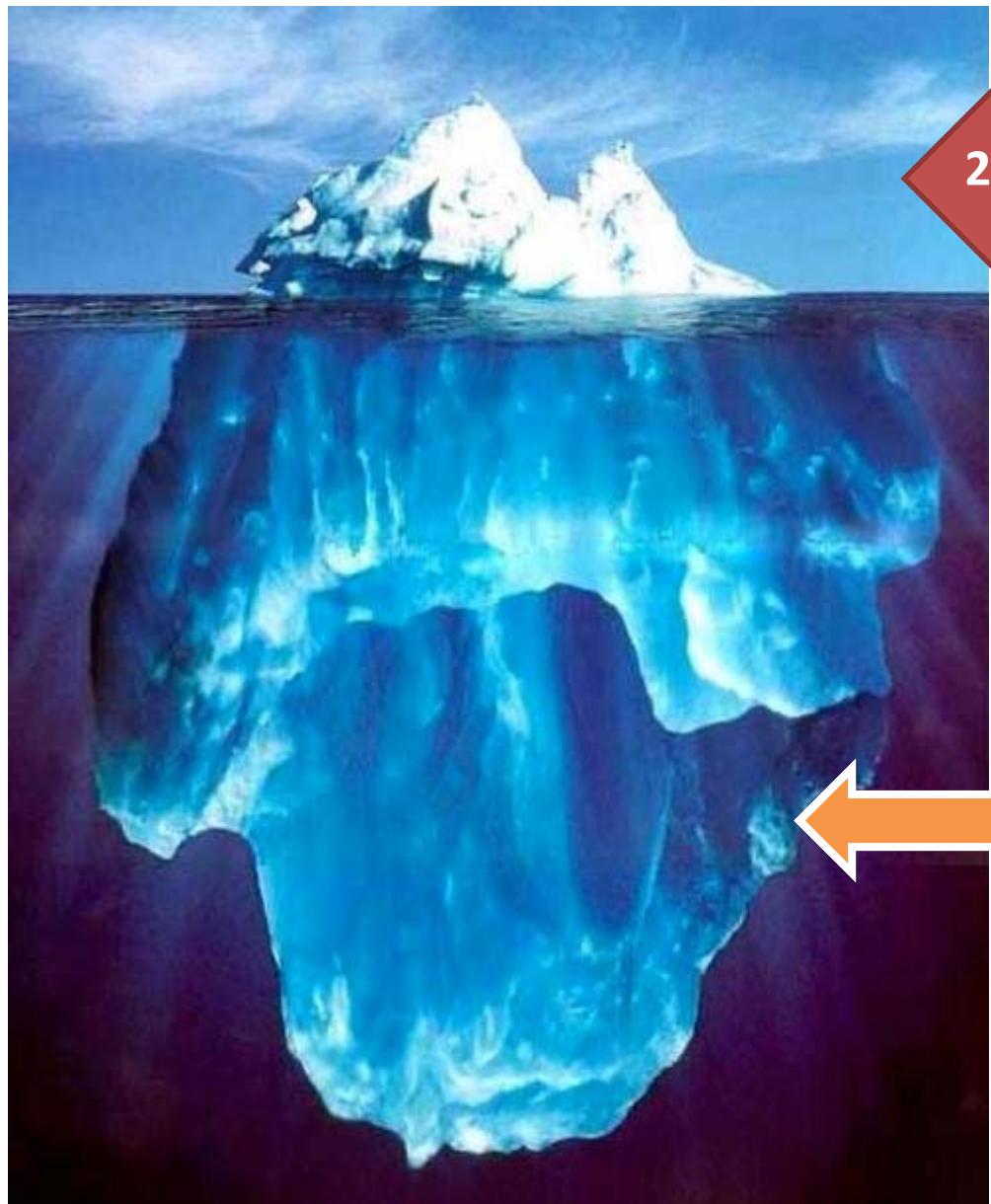


Olivier BOUCHEZ, Frédéric ESCUDIER, Sophie VALIERE



30 Sept 2013

Microbiote dominant

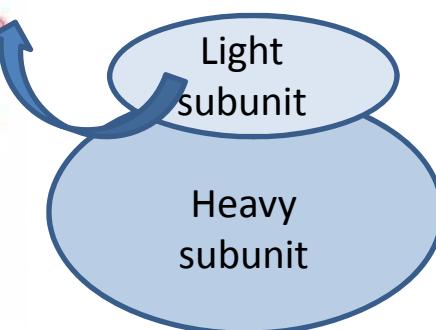
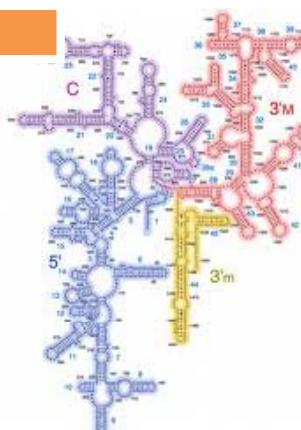


20 to 40%



Re-évaluation par
approches moléculaires
indépendantes de la culture

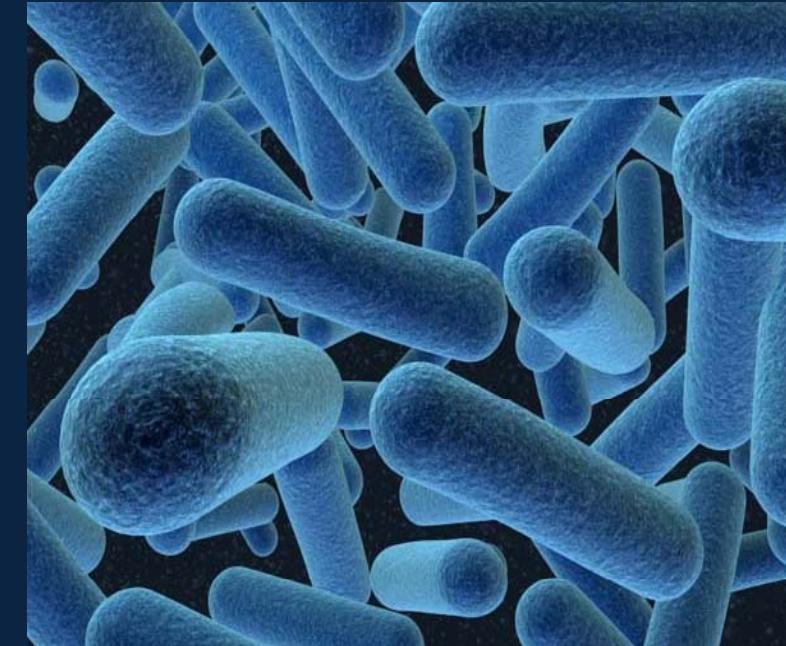
16S RNA



Light
subunit

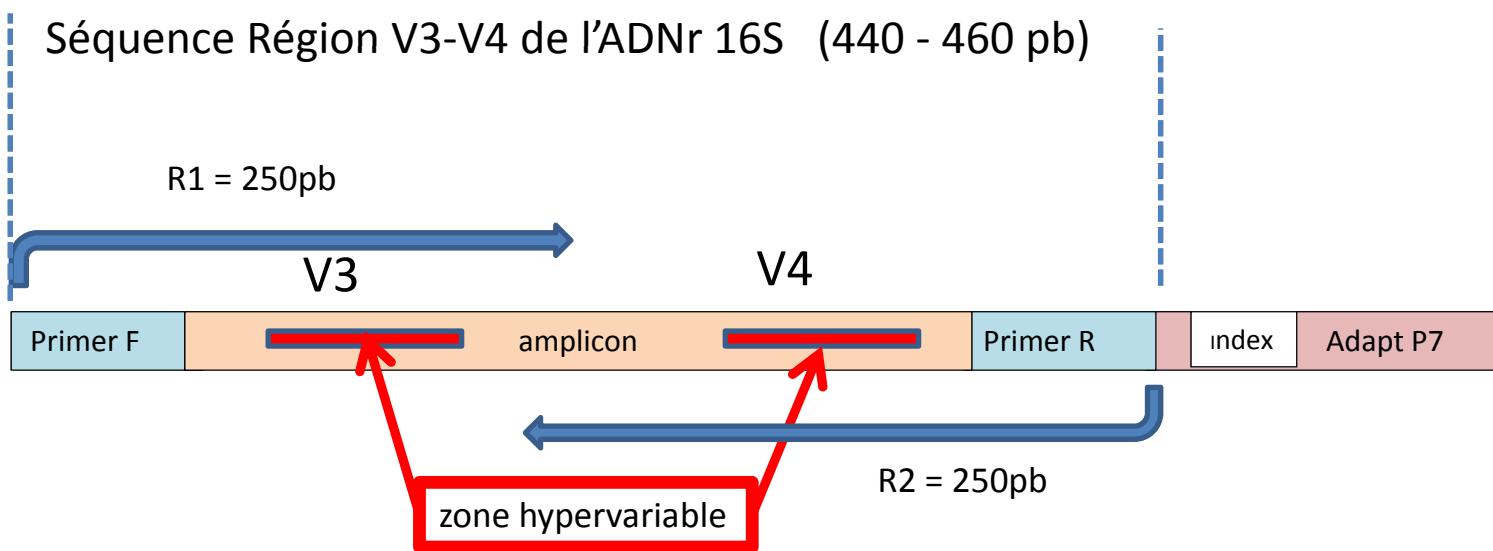
Heavy
subunit

Miseq: mode d'emploi



CONSTRUCTION

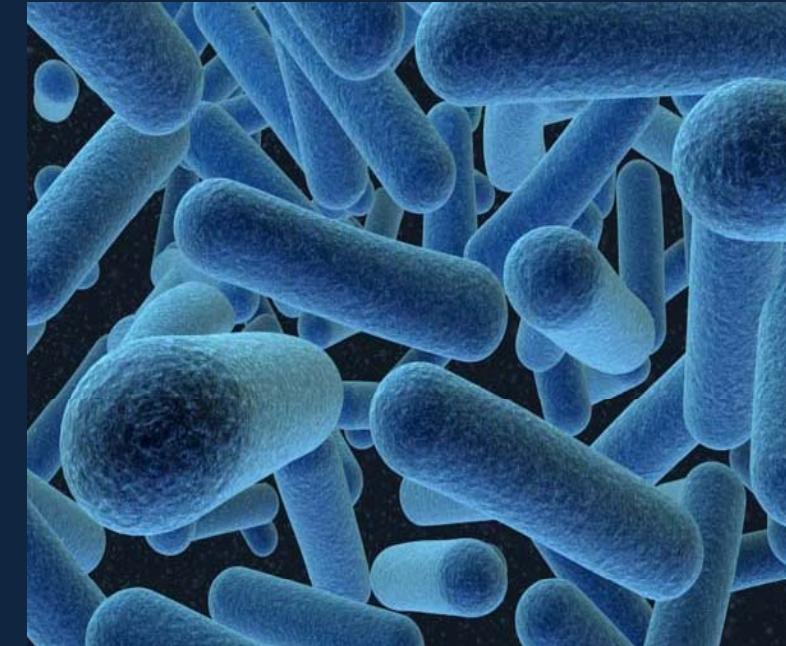
Séquençage paire-end 2x250nt
Géolocalisé (R1 & R2 associés)



Run de 104 échantillons (index de 6 nt):

- Communauté digestive lapin (n=80)
- Communauté digestive porc (n=21)
- Plasmides contenant la séquence ADNr 16S de bactérie connues (n=2)
- Mélange 4 bactéries marines (n=1)

Miseq: les données



Nombre de reads par échantillon

Nombre de reads après filtre Illumina: 24 854 086
(8 % de reads supprimées)

soit

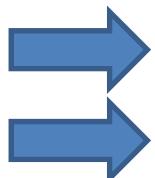
12 427 043 R1

12 427 043 R2

REPARTITION DES SEQUENCES DANS
LES 104 ECHANTILLONS

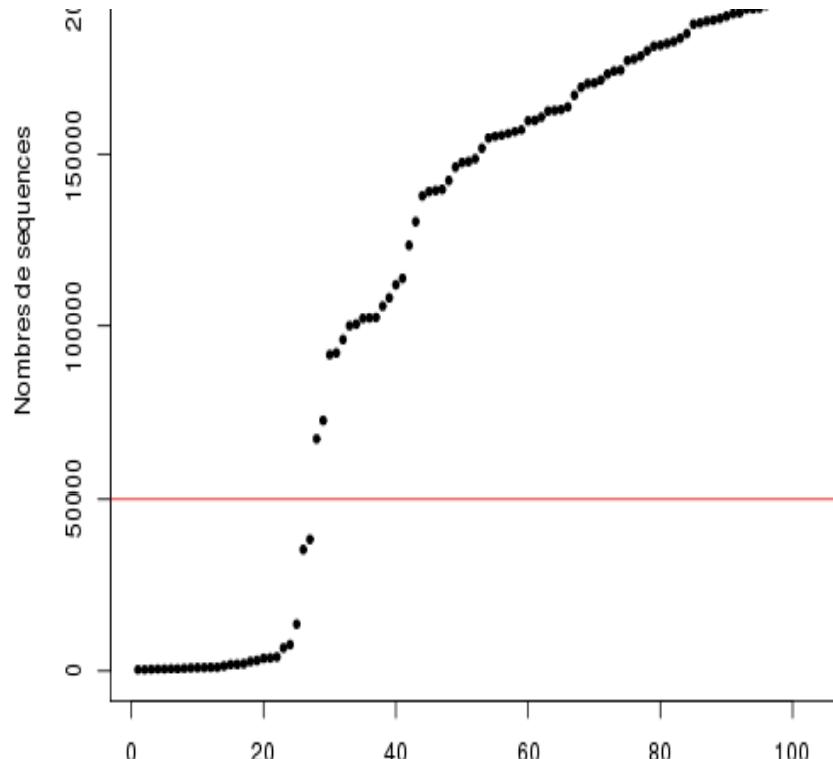
Problème technique:

27 échantillons ont moins de 50 000 séquences après filtre Illumina

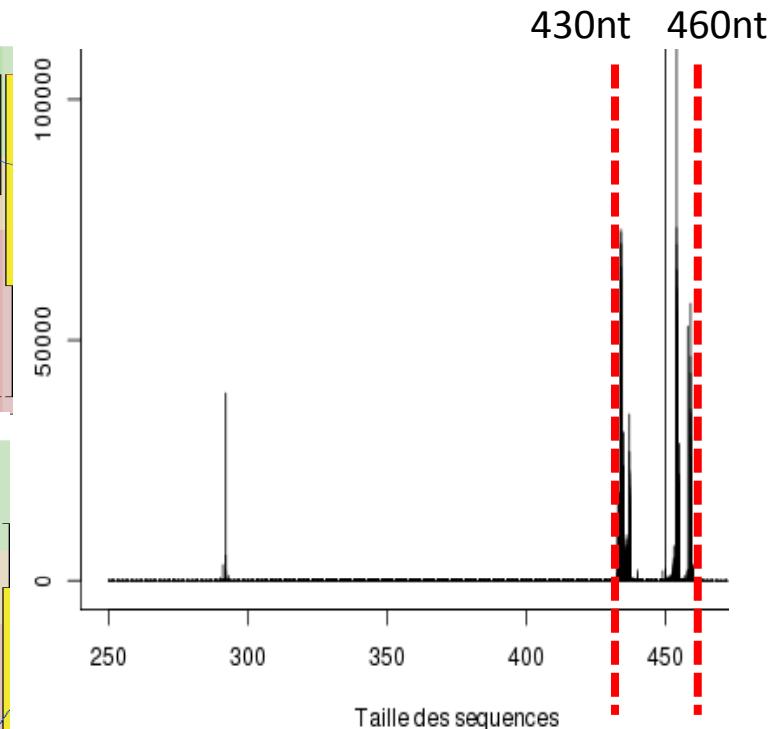
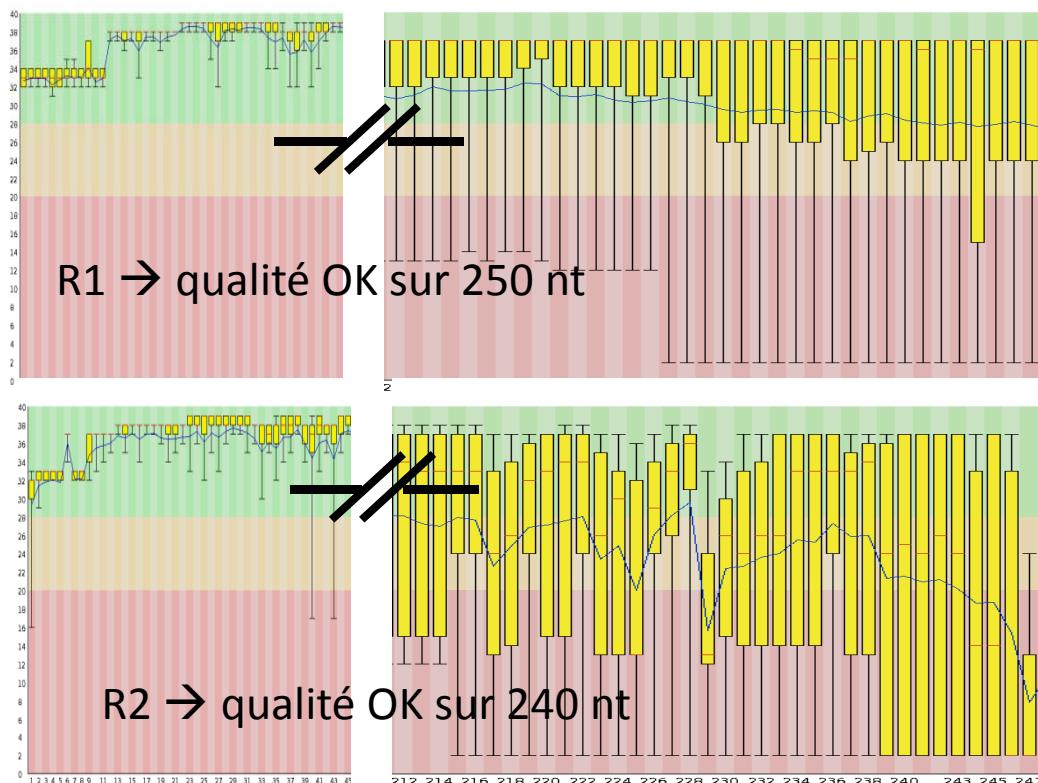


Problème d'amplification sur PCR1 → présence d'inhibiteur

Suite de la présentation sur $104 - 27 = 77$

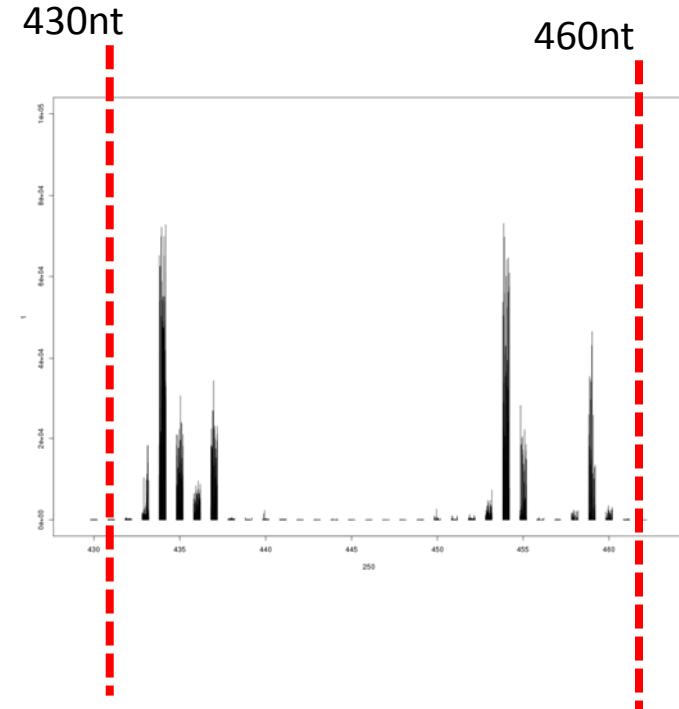
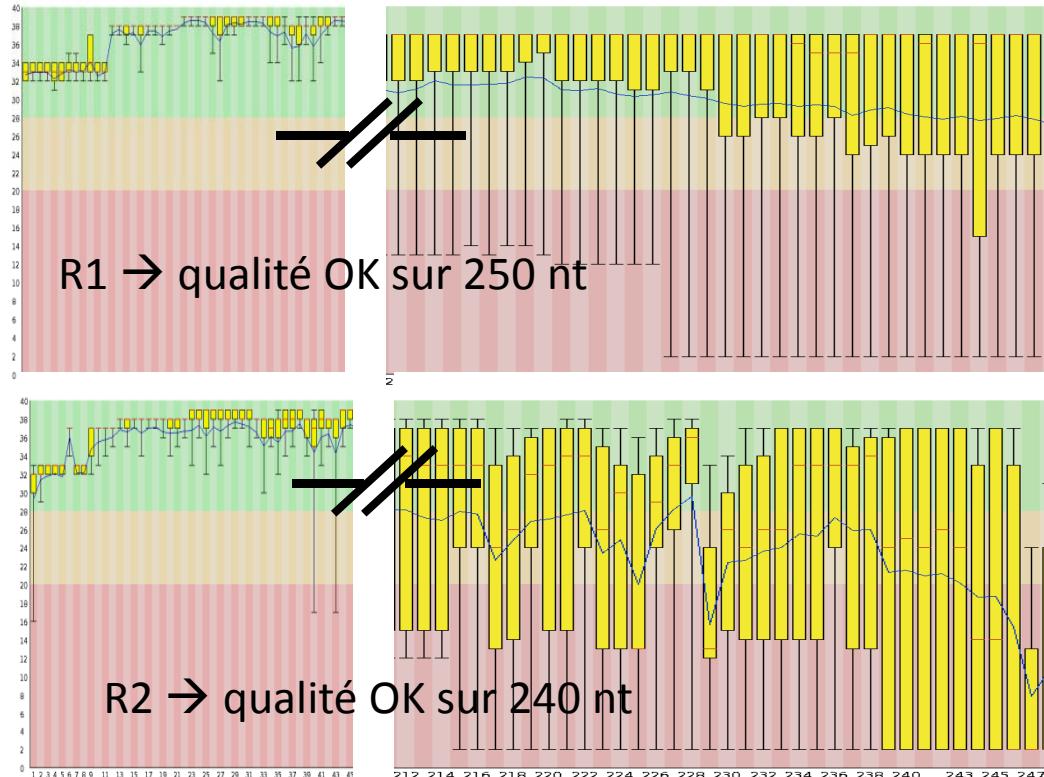


CONTIGAGE PAR FLASH



➤ overlap minimum 10 bases → notre construction nécessite un overlap de min 20 et max 35 nt

CONTIGAGE PAR FLASH



- overlap minimum 10 bases → notre construction nécessite un overlap de min 20 et max 35 nt
- élimine une grande partie des reads contenant des "N" mais pas tous

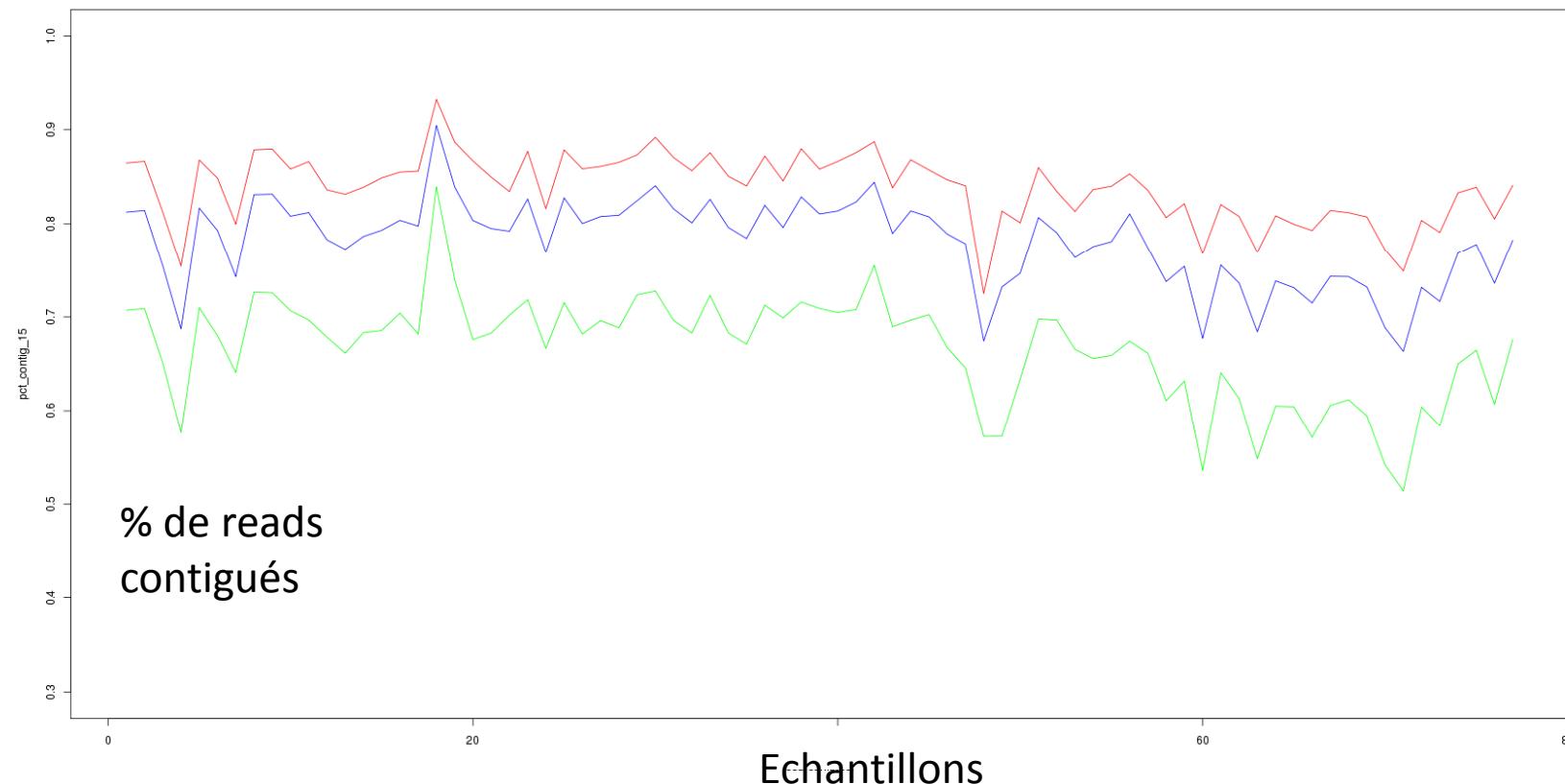
CONTIGAGE PAR FLASH

3 seuils de mismatch testés: 5%, 10%, 15%

A 5% → 65 % des couples R1 R2 sont contigus → 8 millions

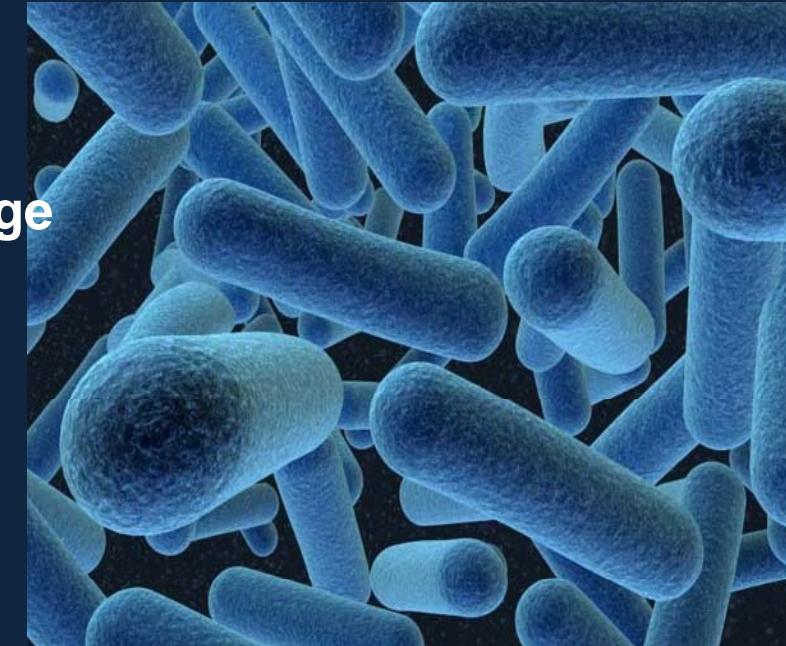
A 10% → 75 %

A 15% → 80 %



LES PLASMIDES

Vérifier
la qualité du contigage
et du processus de séquençage



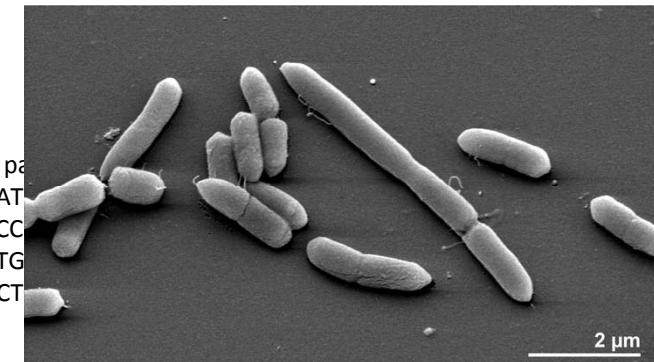
Plasmides

Prevotella bryantii (Bacteroides): NED4B6 (454nt)

>gi|129594480|gb|EF445235.1| Uncultured bacterium clone NED4B6 16S ribosomal RNA gene, partial sequence
TACGGGAGGCAGCAGTGAGGAATTGGTCAATGGACGGAAGTCTGAACCAGCCAAGTAGCGTCAGGAT
ATAAAGTTAGCCACGTGTTATTGCATGTACCTACGAATAAGGACGGCTAACCTGGCAGCAGCC
ATTGGGTTAAAGGGAGCGCAGGCCGTTGGTAAGCGTGTGAAATGTCCGGGCTAACCTGGGCACTG
GGCGGAATTCGTGGTAGCGGTGAAATGCTTAGATATCAGAAGAACTCCAATTGCGAAGGCAGCTCGCT
TCGAACAGGATTAGATACCCGGTA



208 824 R1 ou R2 brut
137 622 contigs (66%)

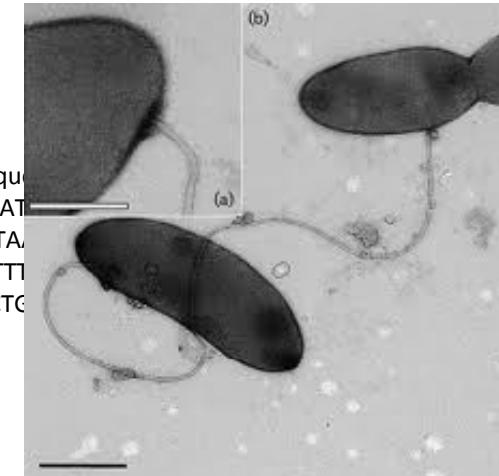


Butyrivibrio fibrisolvens (Firmicutes): NED4E2 (450nt)

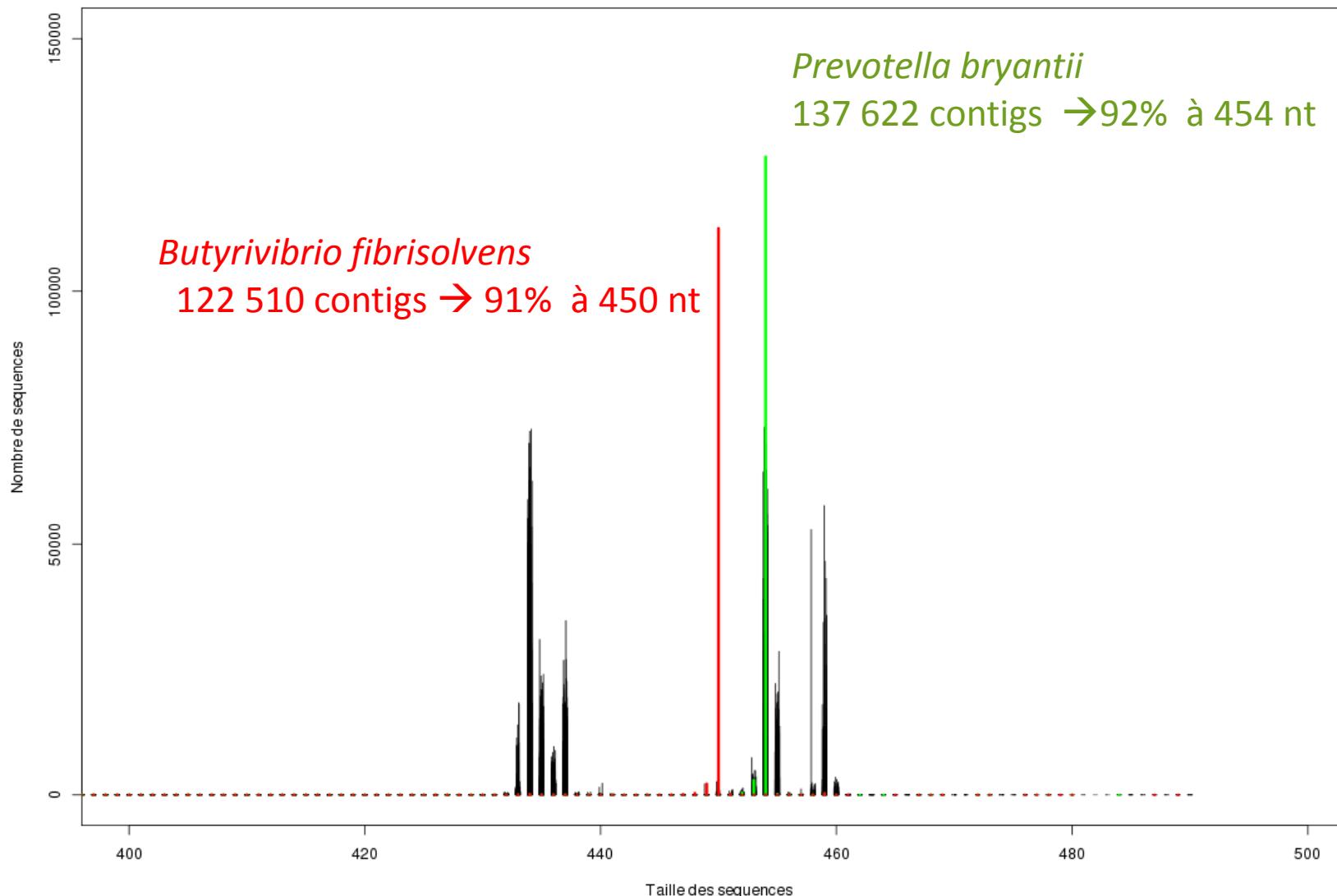
>gi|129594507|gb|EF445262.1| Uncultured bacterium clone NED4E2 16S ribosomal RNA gene, partial sequence
TACGGGAGGCAGCAGTGGGGATATTGCACAATGGCGAAAGCCTGATGCAGCGACGCCGTGAGTGAAGAAGTAT
GAAGAAAAGACCTCGTAAGAGGGGATGACGGTACCTGAGTAAGAAGCCCCGGCTAACTACGTGCCAGGCCGGTA
ATTACTGGGTGAAAGGGAGCGTAGACGGTGTATCAAGTCTGAAGTGAAGTAAACCCACGGCTAACCGTGGCTTGGCTT
GAGGTAAAGCGGAATTCTGGTAGTAGTGAATGCGTAGATATCAGGAAGAACATCAGTGGCGAAGGCAGCTTACTG
GGCGTGGGGAGCAAGATACCCGGT



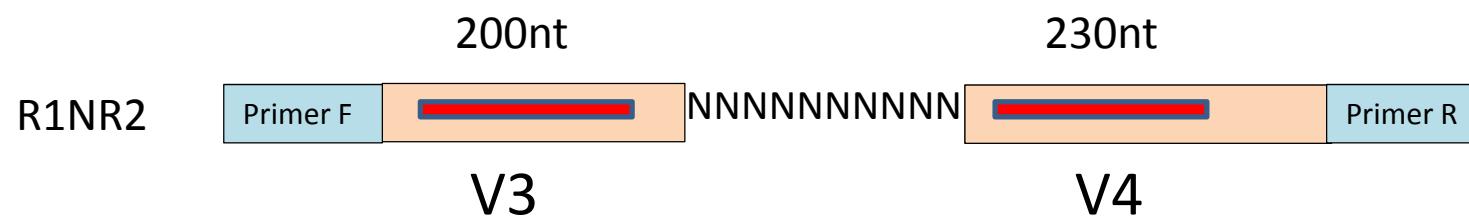
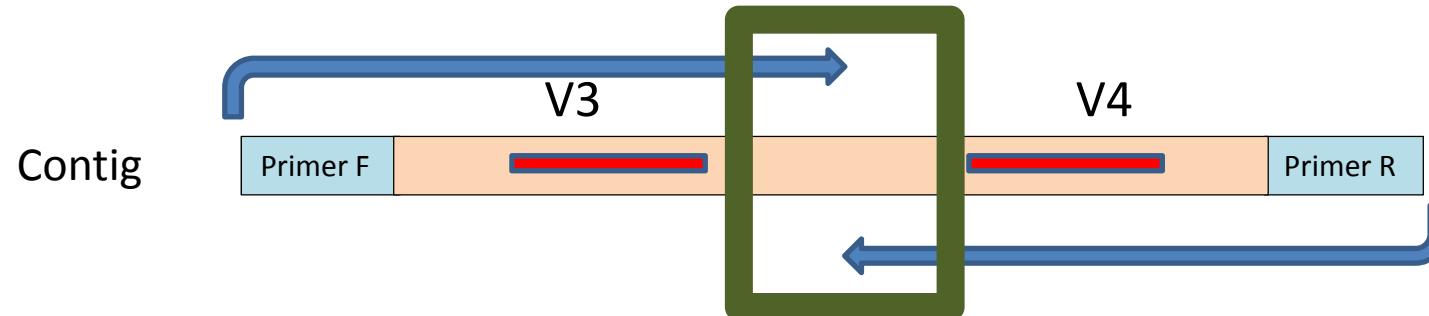
181 734 brut
122 510 contigs (67%)



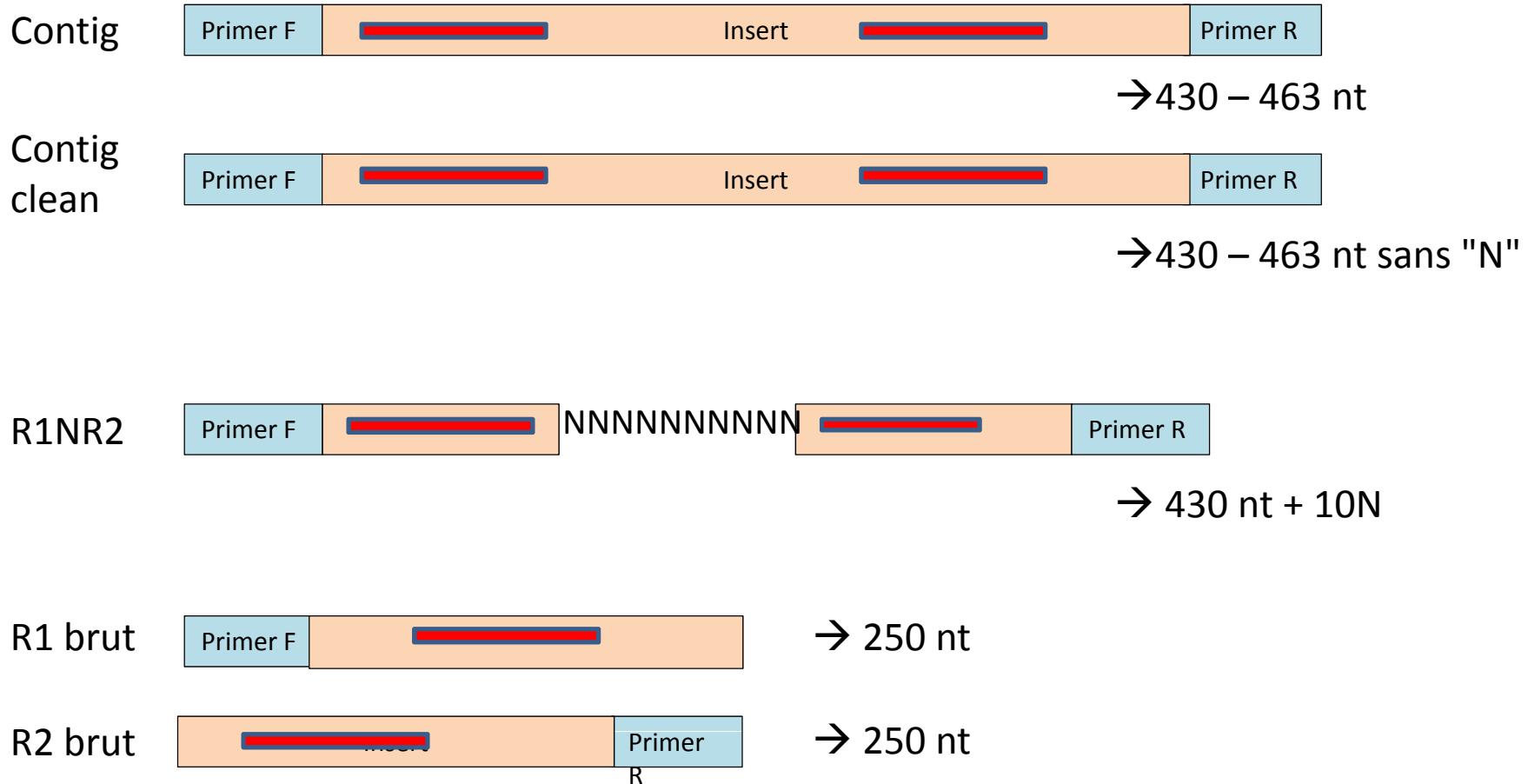
Plasmides



Plasmides les constructions comparées



Plasmides les constructions comparées



2 façons d'analyser les communautés

Reads

Assignation taxonomique

Mothur (RDP classifier) + Base LTP111

1 read = 1 taxon (bootstrap 85- 95 %)

Regroupement par chaîne de caractères

Clusterisation = regroupement en OTU

Mothur (RDP classifier) + Base LTP111

+ **Esprit Tree**

Chaque read est classé dans 1 OTU puis
assignation taxonomique des OTU

Table d'abondance

100 - 400 taxons connus ou «unclassified»

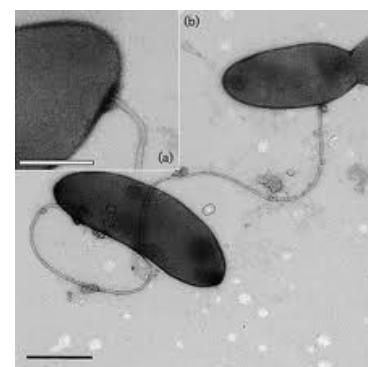
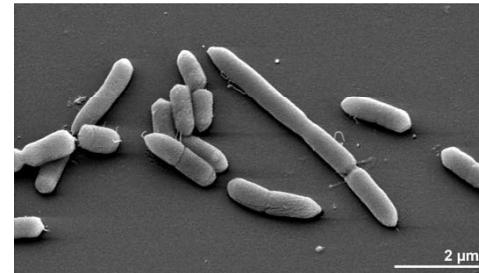
Y-matrix	Taxon 1	Taxon 2	Taxon 3	...	Taxon 196	Taxon n	TOTAL
sample 1	0	22	0	...	826	0	5399
sample 3	0	1	0	...	53	0	3155
sample 4	0	5	1	...	5694	0	7791
...
sample 100	0	2	4	...	5943	1	11421

90 000 OTU

Y-matrix	OTU 1	OTU 2	OTU3	...	OTU3176	OTU n	TOTAL
sample 1	0	12	0	...	2320	0	7928
sample 3	0	64	0	...	76	28	16616
sample 4	0	748	36	...	52	0	5436
...
sample 100	0	48	4	...	380	0	3248

Affiliation taxonomique (RDP classifier – LTP111)

		<i>Prevotella bryantii</i>			
Boot	Contig	ontig clean	R1NR2	R1 (V3)	R2 (V4)
95	94.5	99.2	93.4	91.2	88.4
100	94.4	99.0	90.9	86.0	83.0



		<i>Butyrivibrio fibrisolvens</i>			
Boot	Contig	ontig clean	R1NR2	R1 (V3)	R2 (V4)
95	94.2	99.1	90.4	88.9	80.8
100	88.2	91.2	74.1	72.4	31.0

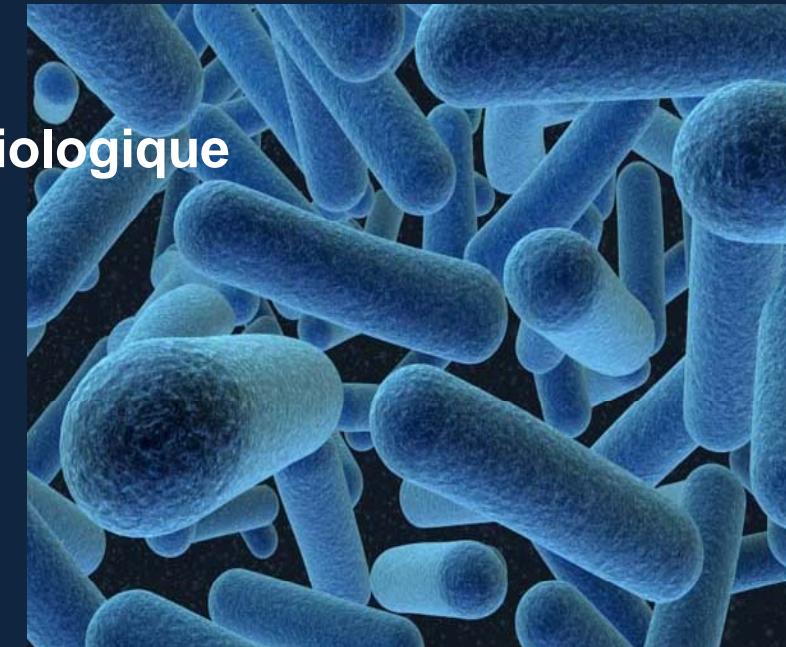
Clusterisation en OTU (Esprit-tree) sur contig

	<i>Prevotella bryantii</i>	<i>Butyrivibrio fibrisolvens</i>
Contig	137 622	122 510
Nb OTU attendu	1	1
Nb total OTU	1397	998
OTU 1 (maj)	93.6 %	93.6 %
OTU 2	0.2 %	0.2 %

To be continued...
- Contig clean
- R1NR2
- R1
- R2

Comparaison 454 vs MiSeq

Comparer les profondeurs
L'impact sur l'interprétation biologique



Comparaison de communautés digestives de porcs 454 vs Miseq

15 échantillons

3 lots : **propre, sale, t1** (n=5)

454 (USA)

61 158 reads

Miseq (PLAGE)

1 211 554 reads

Classification:

RDP classifier base LTP 111

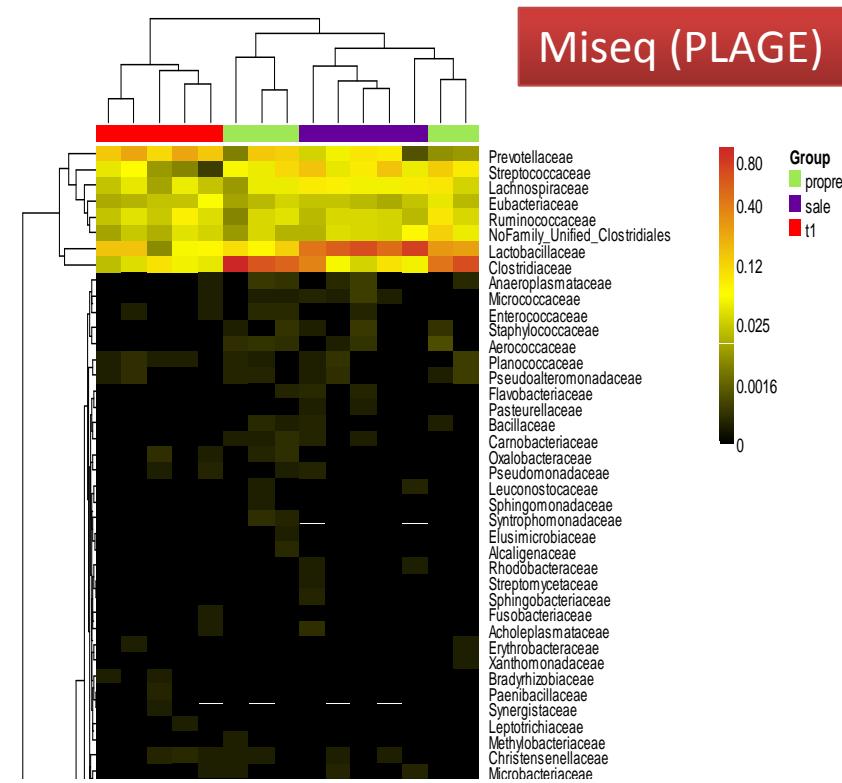
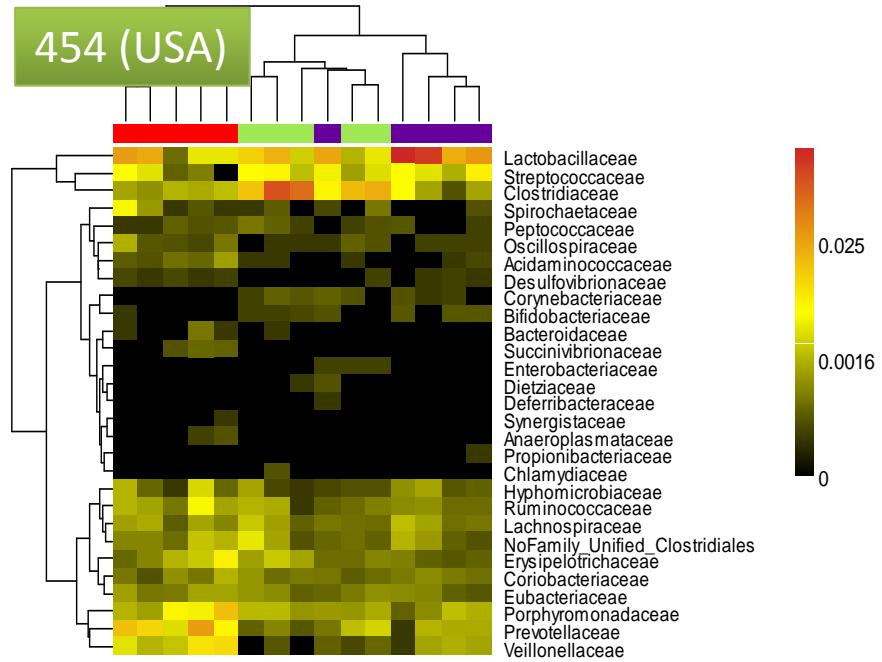
2 niveaux taxonomiques :

- famille (85%)
- genre (95%)

Clusterisation:

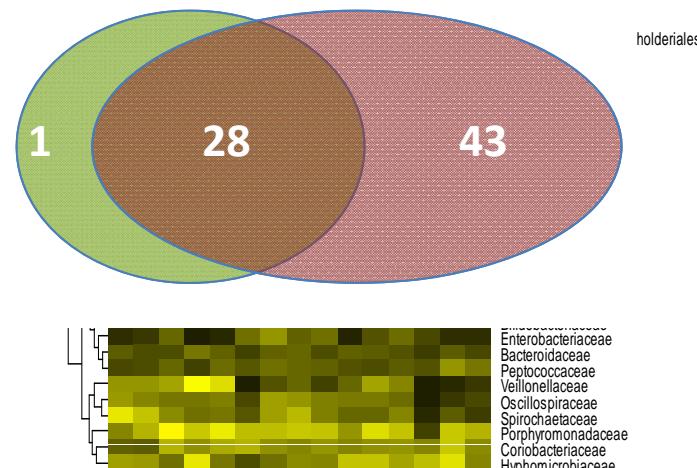
Esprit-tree

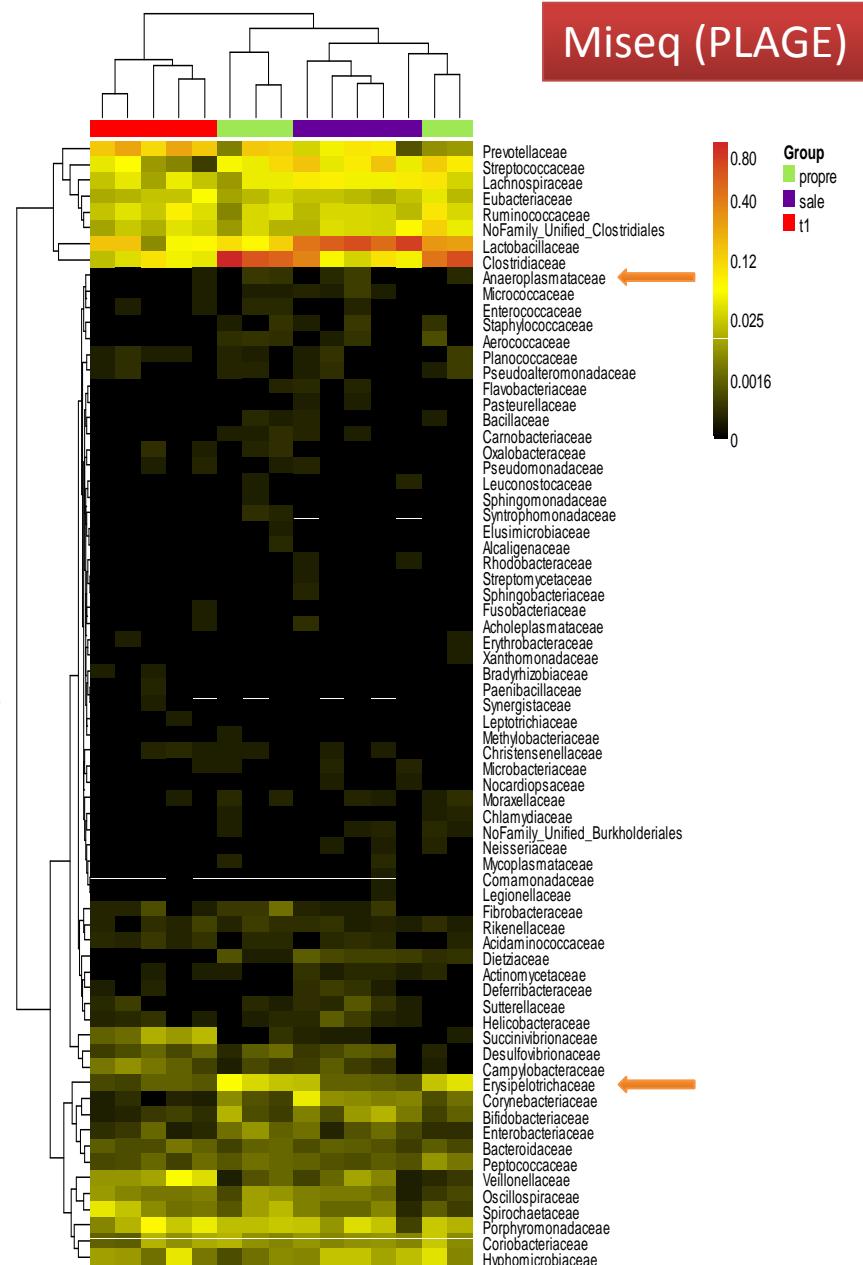
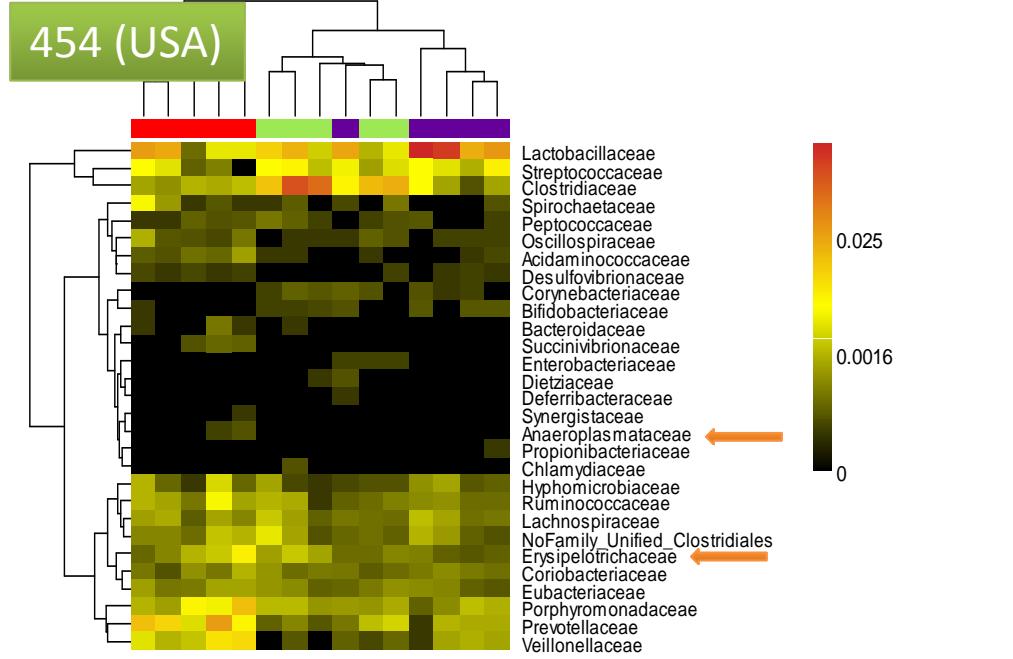
Avec préclusterisation par
classification RDP



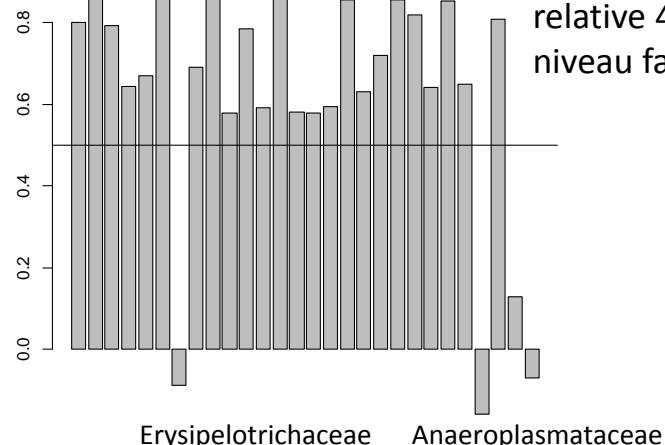
Classification (RDP classifier – LTP111) niv famille

	454	MISEQ
Nb seq tot	61158	1211554
Tx d'affiliation	85%	71%
Nb taxons	29 taxons	71 taxons



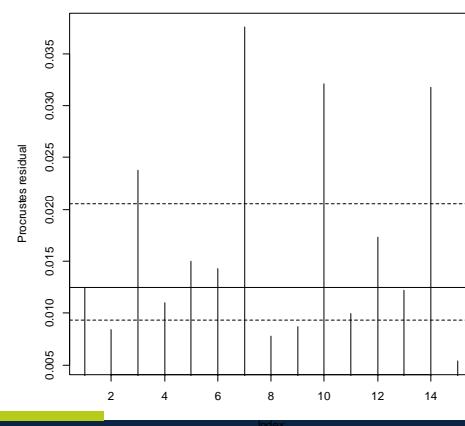
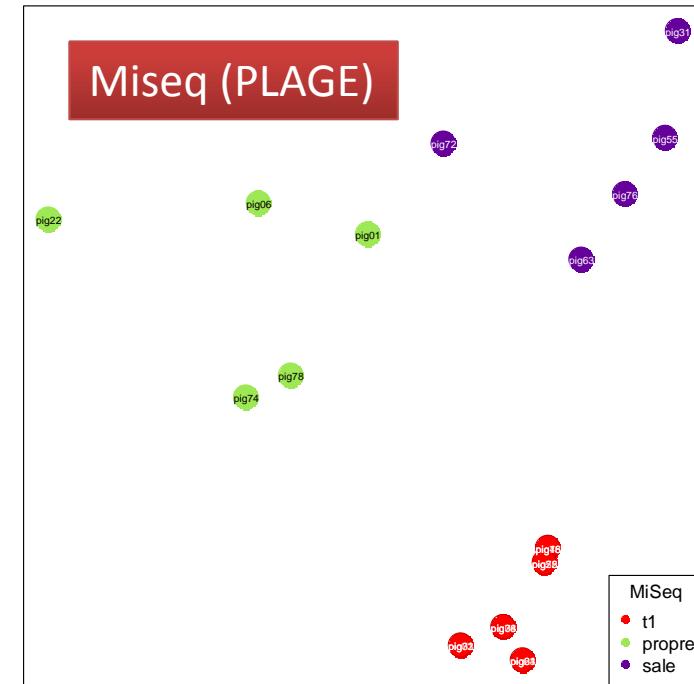
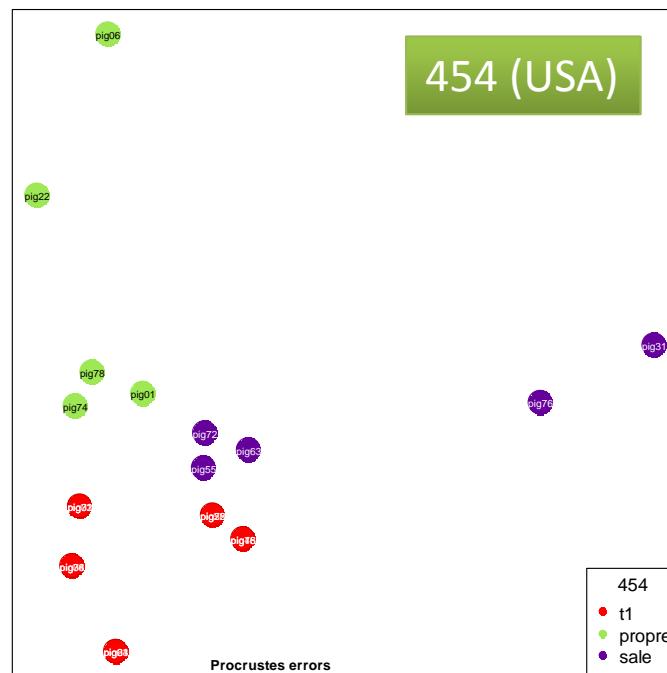


Corrélation abundance relative 454 vs MiSeq niveau famille



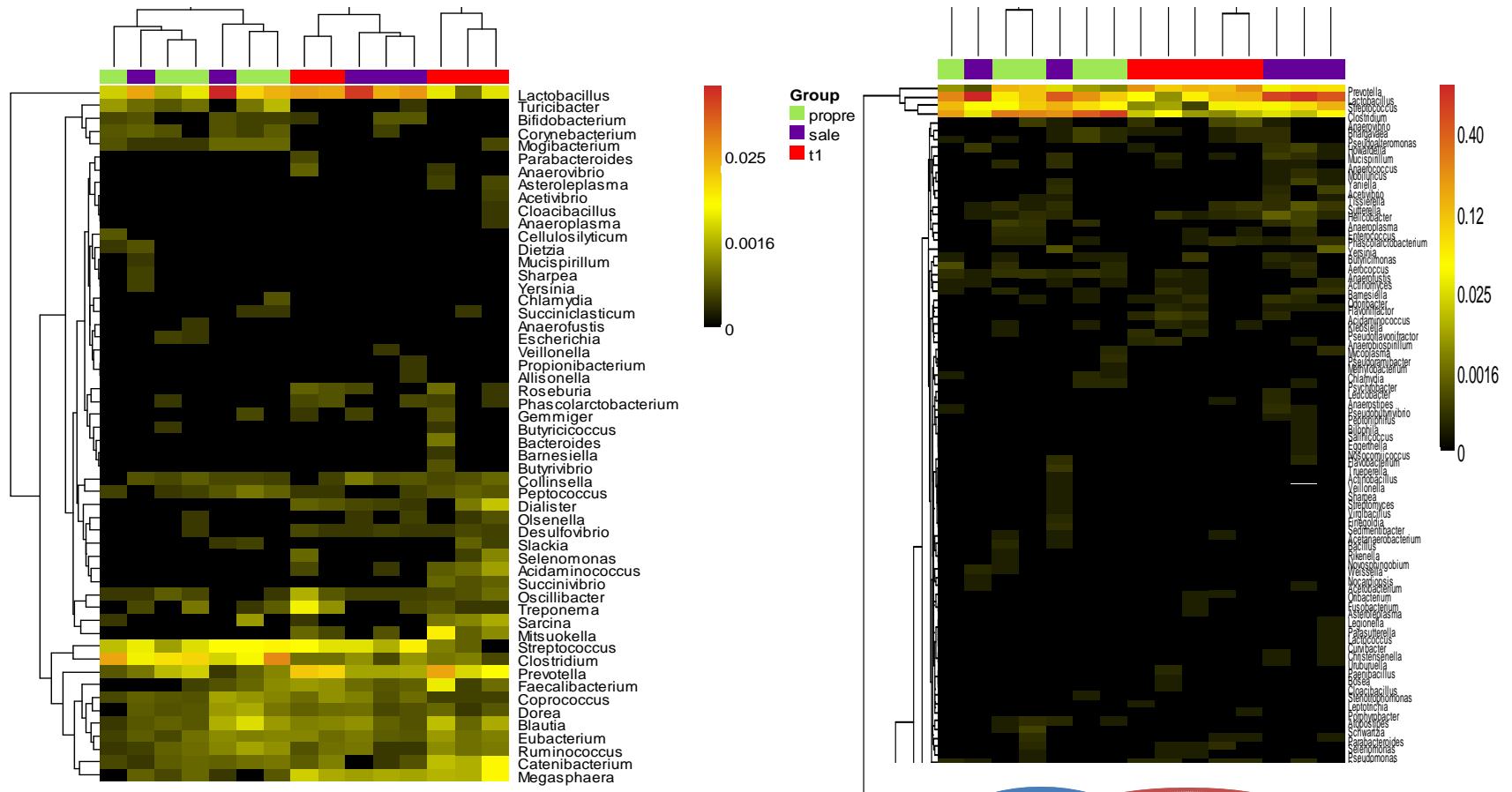
.021

Comparaison des profiles taxonomiques par nMDS



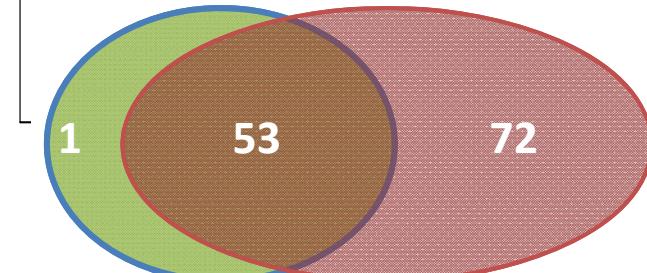
Classification (RDP classifier – LTP111)
niv famille

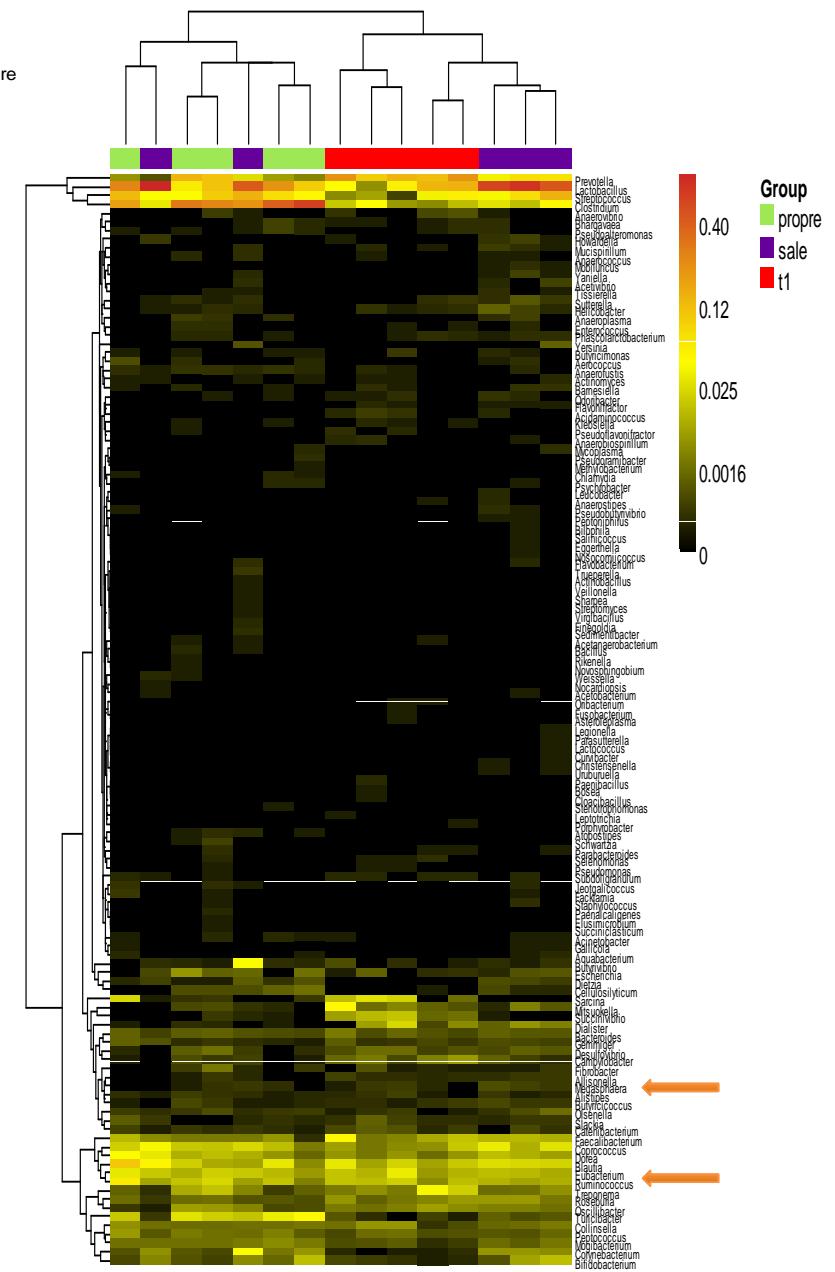
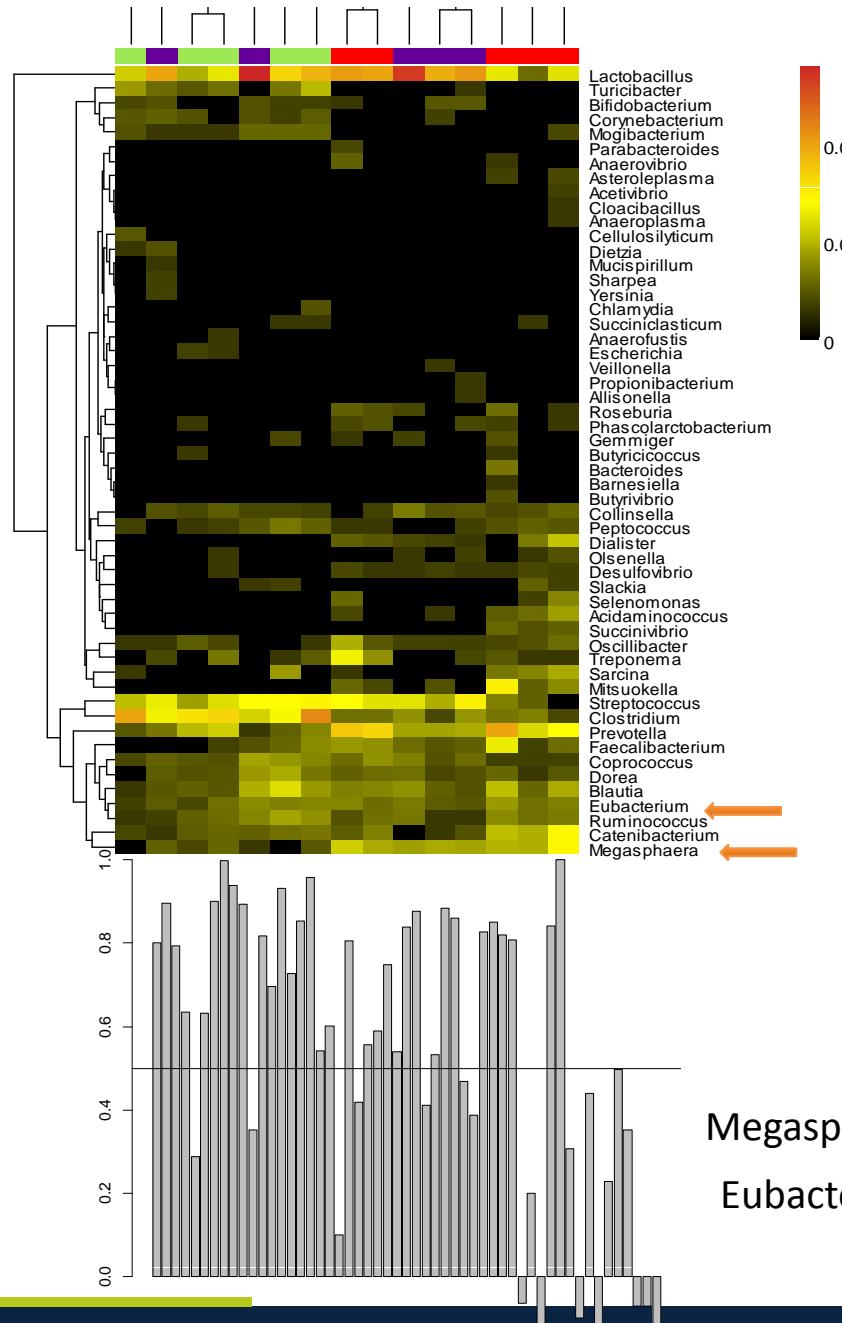
Représentation robuste
→ 4 individus ont un positionnement significativement différent



Classification (RDP classifier – LTP111) niv genre

	454	MISEQ
Nb reads	61 158	1 211 554
Tx d'affiliation	67 %	53 %
Nb taxons	54 taxons	125 taxons

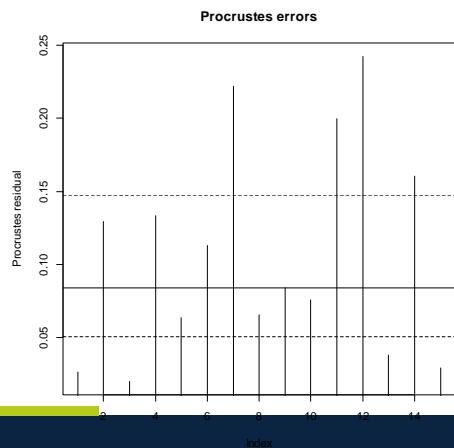
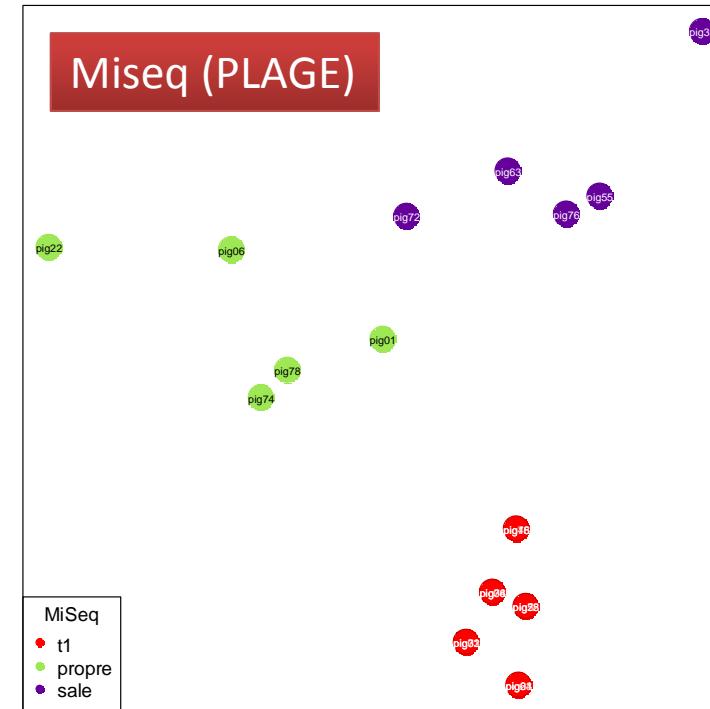
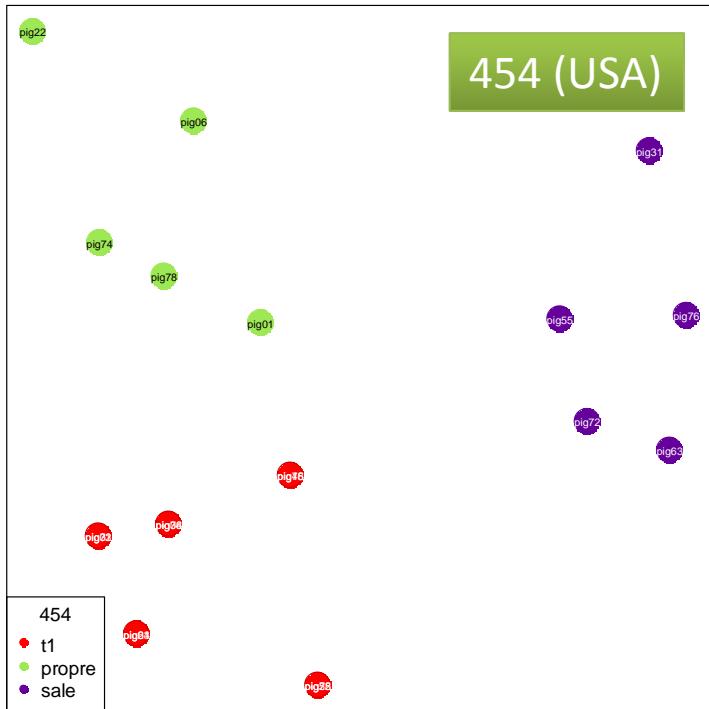




Clusterisation : Esprit Tree (0.03)

	454	MISEQ
Nb seq tot	61 392	1 227 651
Nb OTU	4 346	75 636
Singleton	56%	61%

Comparaison des profiles taxonomiques par nMDS



Clusterisation : Esprit Tree (0.03)

Représentation robuste
→ 4 individus ont un positionnement significativement différent

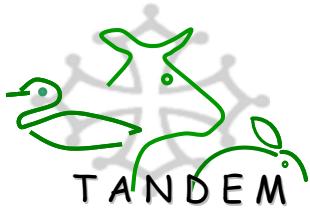
Conclusion

Miseq (PLAGE)

Qualité des séquences satisfaisantes
Contigage possible, suppression des bases N
Profondeur accrue
Moindre coût
Multiplexage plus important
Résultats cohérents d'un point de vue composition communauté

Mais

Difficulté de traitement dû à l'augmentation du nb de reads
→ Pipeline à revoir



Thanks for your attention

Merci pour votre attention

Thanks to each of my team members

