

# How does the Nanopore technology meet the needs of our users ?

ONT – 6 Juin 2017

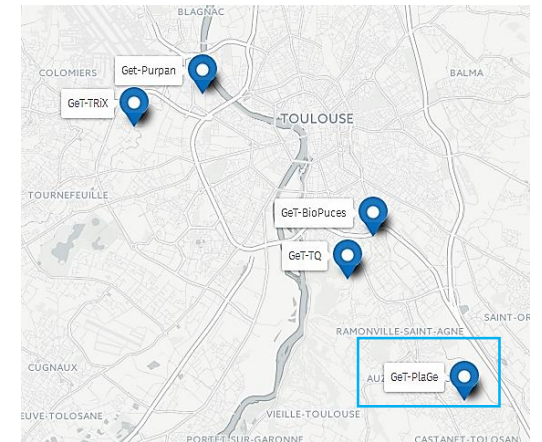
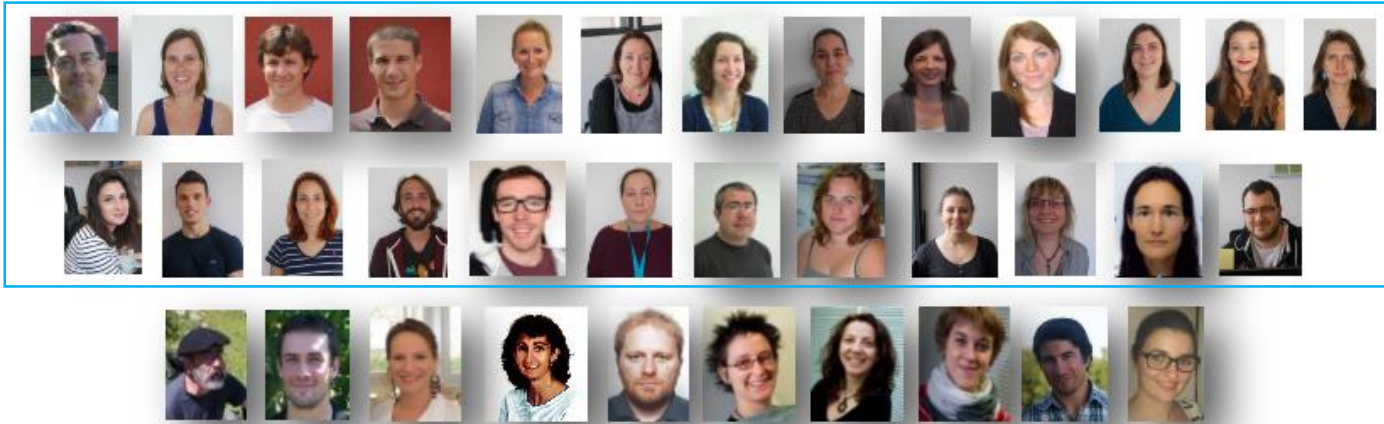
Catherine Zanchetta



*Team and expertise*

## ③ 35 people on 5 sites

- Experts in Agronomy, Environment, Microbiology, Health
- Competence in biology, bioinformatics, biostatistics



⑤ **More than 140 laboratories in 2016** (INRA, CNRS, INSA, INSERM, CHU, CIRAD ...)

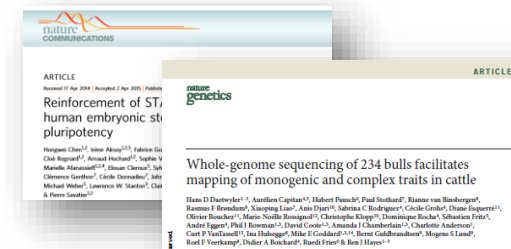
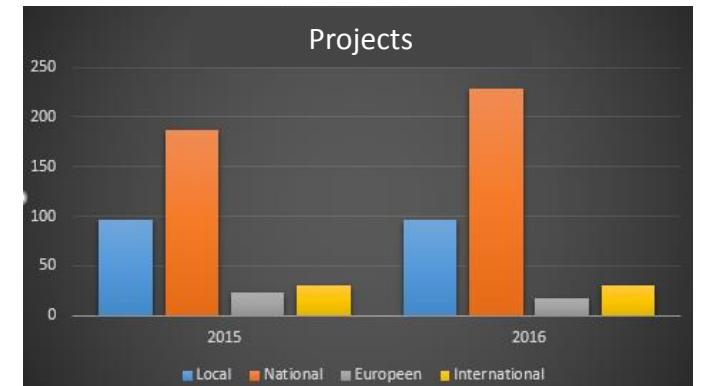
- More than 240 research teams
- More than 360 projects
- >2M€ of activity

⑤ **10 R&D projects**

- Chromium, MinION, ChIPseq, methylation, HiC...

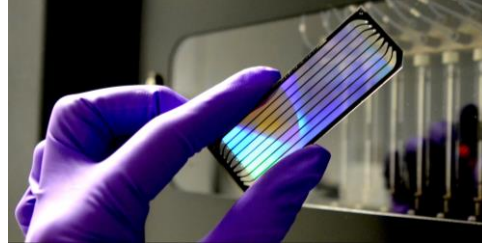
⑤ **19 Research projects (ANR, INCA, H2020 ...)**

⑤ **91 Publications as co-author since 2010**

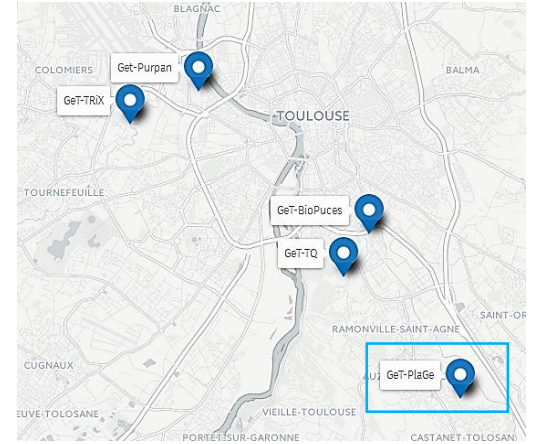




Short reads



Illumina : MiSeq + HiSeq 3000

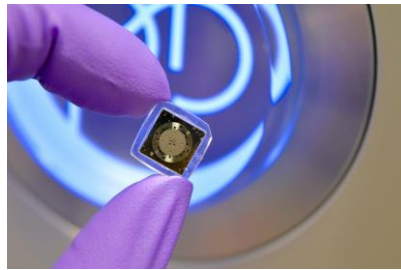


Synthetic long reads



10x genomics : Chromium

Long reads



Pacbio : RSII



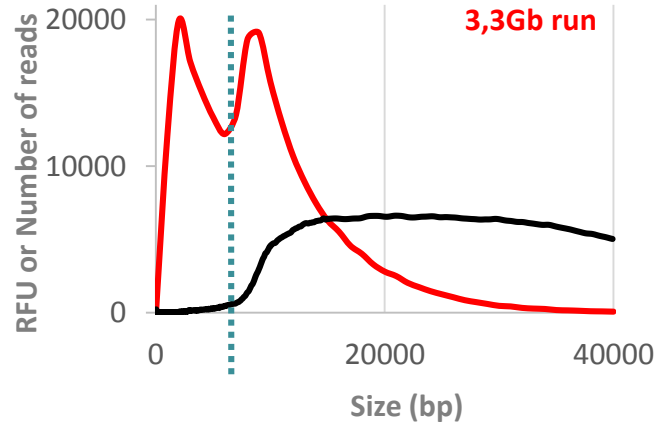
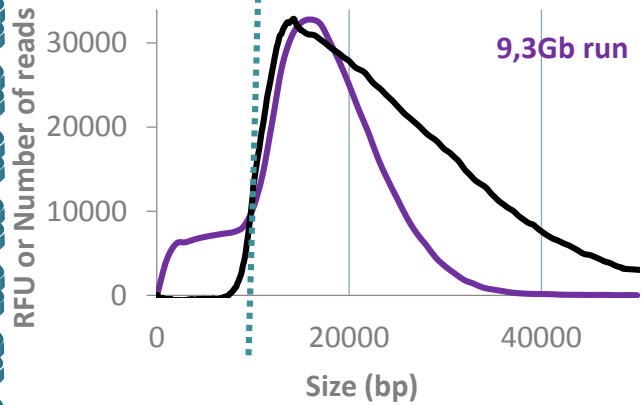
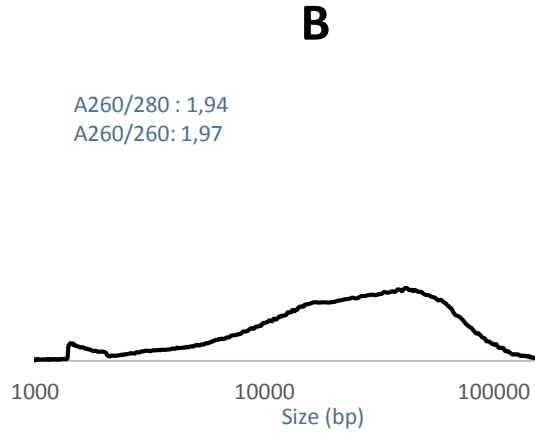
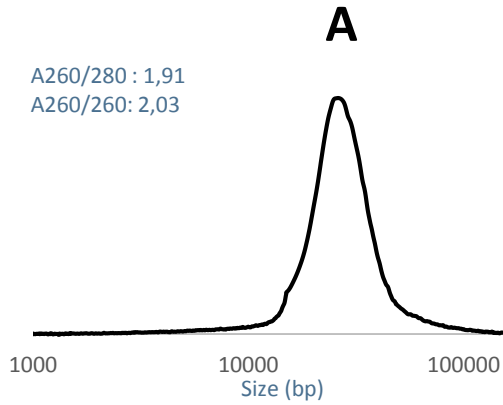
ONT : MinION

# What am I going to talk about for the next 20 minutes ?

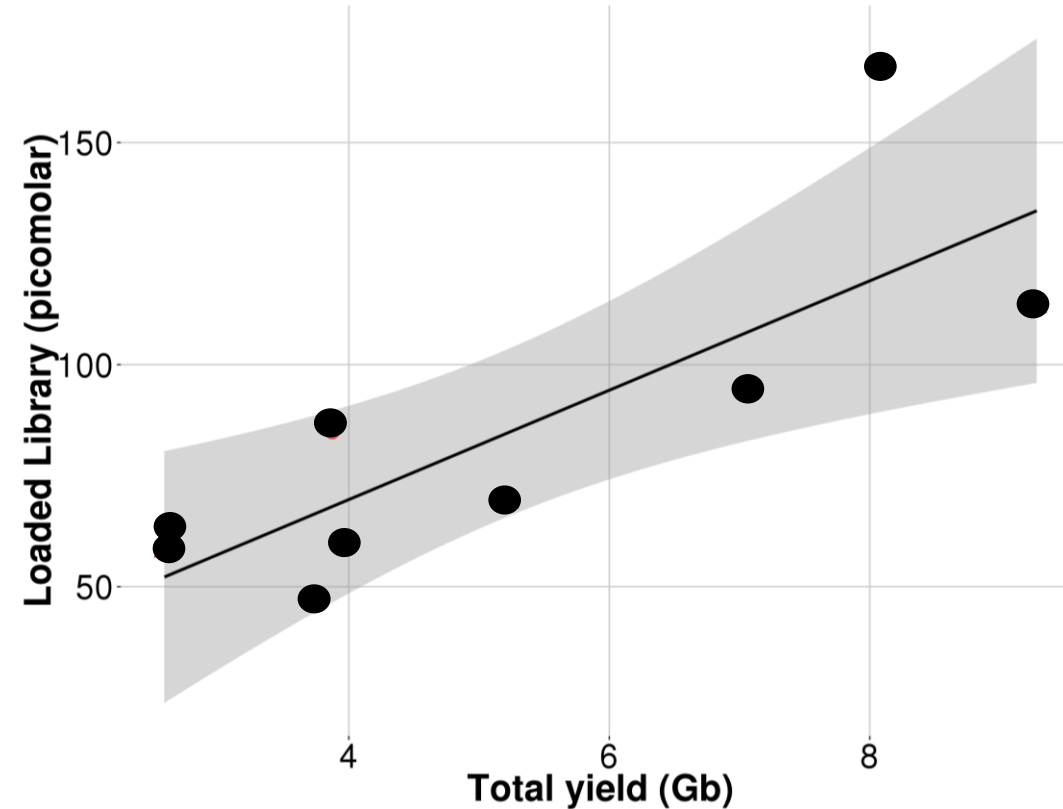
- ④ **Assesment of the technology**
  
- ④ **4 biological questions (gDNA)**
  - **Production of sequencing data**
  - **Veterinary diagnosis**
  - **Agronomics reasearch**
  - **Complexe genome, contaminant molecule**

# Assesment of the technology

*What can I expect with the MinION ?*



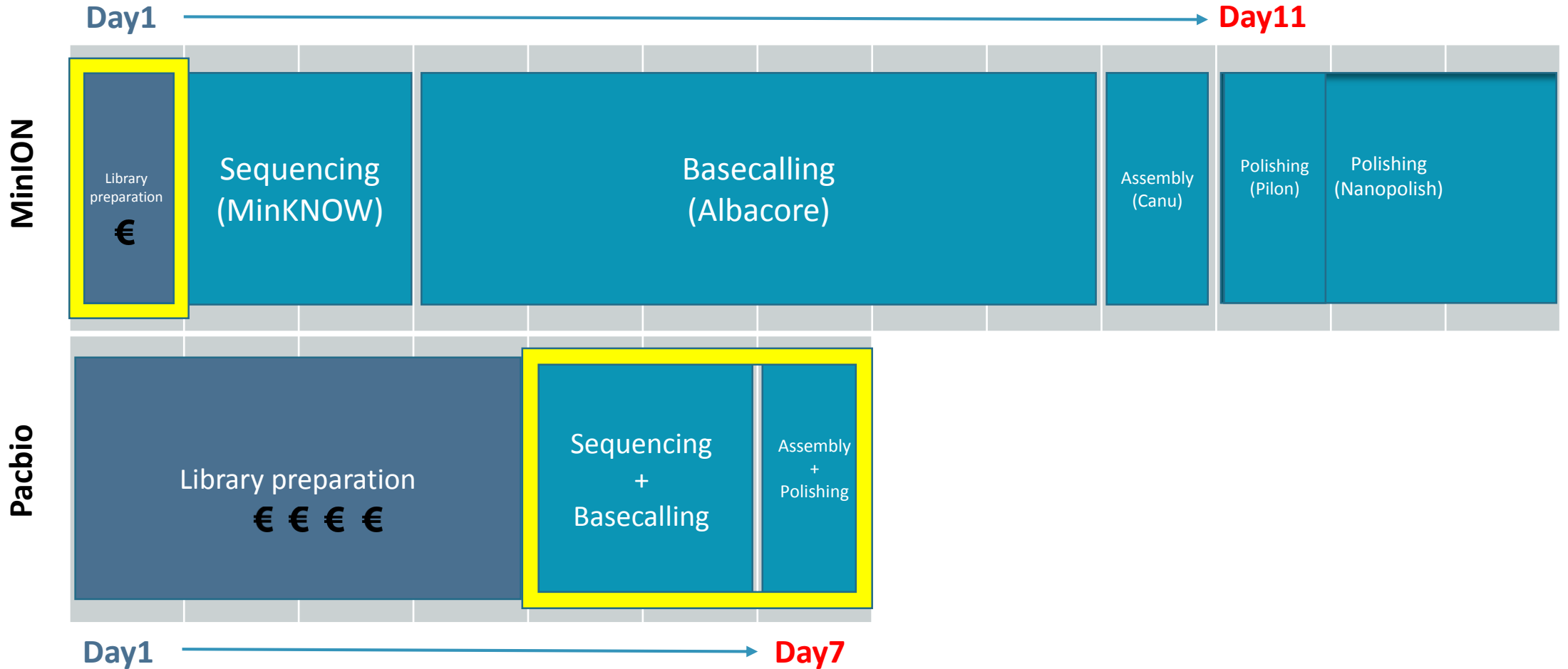
Library  
Reads  
Blue Pippin Cutoff



# Assesment of the rechnology

*When will I get my results ?*

**Example of a bacterial genome assembly :**  
Same quantity of data / Same informatics resources





# Assesment of the technology

*Accuracy on Raw Data and Qualitative analysis of an assembly what is the best option ?*

Example of a bacterial genome assembly

Technologie	Accuracy
Illumina	99.7 %
<b>Pacbio</b>	<b>81.3 %</b>
MinION Albacore V0	81.0 %
<b>MinION Albacore V1</b>	<b>86.3 %</b>

Assembly methods	% Complete genes	% Fragmented genes	% Missing genes
MinION	12.2	27	60.8
MinION-nanopolish	71.6	15.5	12.9
<b>MinION-pilon</b>	<b>95.3</b>	<b>0</b>	<b>4.7</b>
<b>Pacbio-hgap3</b>	<b>95.3</b>	<b>0</b>	<b>4.7</b>

# MinION and Users needs

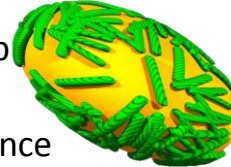
*Biological questions, technical answers*

250 kb

Production  
BAC clones sequencing  
**Same or better assemblies compared to the Pacbio**


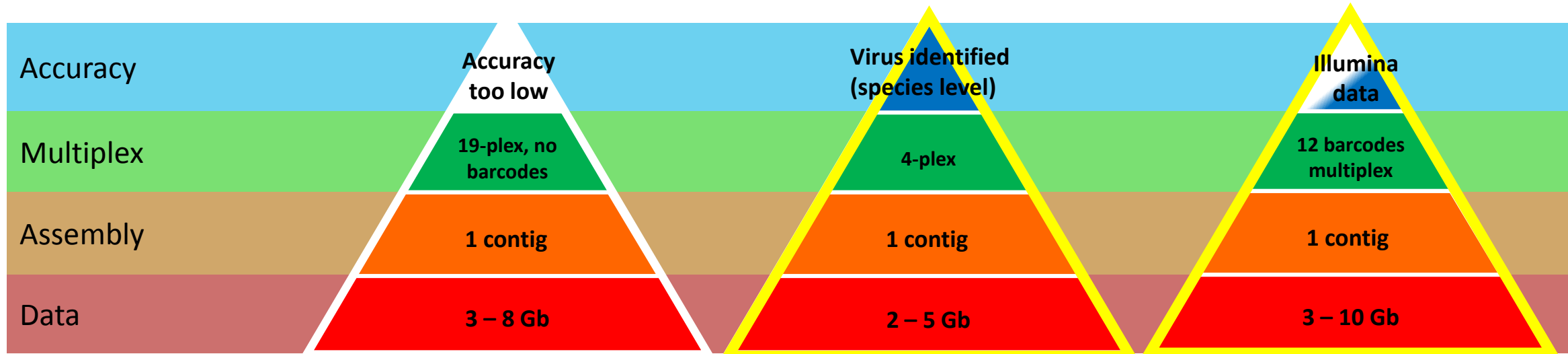
300 kb

Veterinary surveillance  
Support tool  
**Virus identification in a biological sample**



5 Mb

Plant-pathogen arm race  
Link Xcc genotypes to *A. thaliana* phenotypes  
**Identification of SNPs**

Caroline Callot, Stéphane Cauet, H  l  ne Berges

Guillaume Croville, Jean-luc Guerin

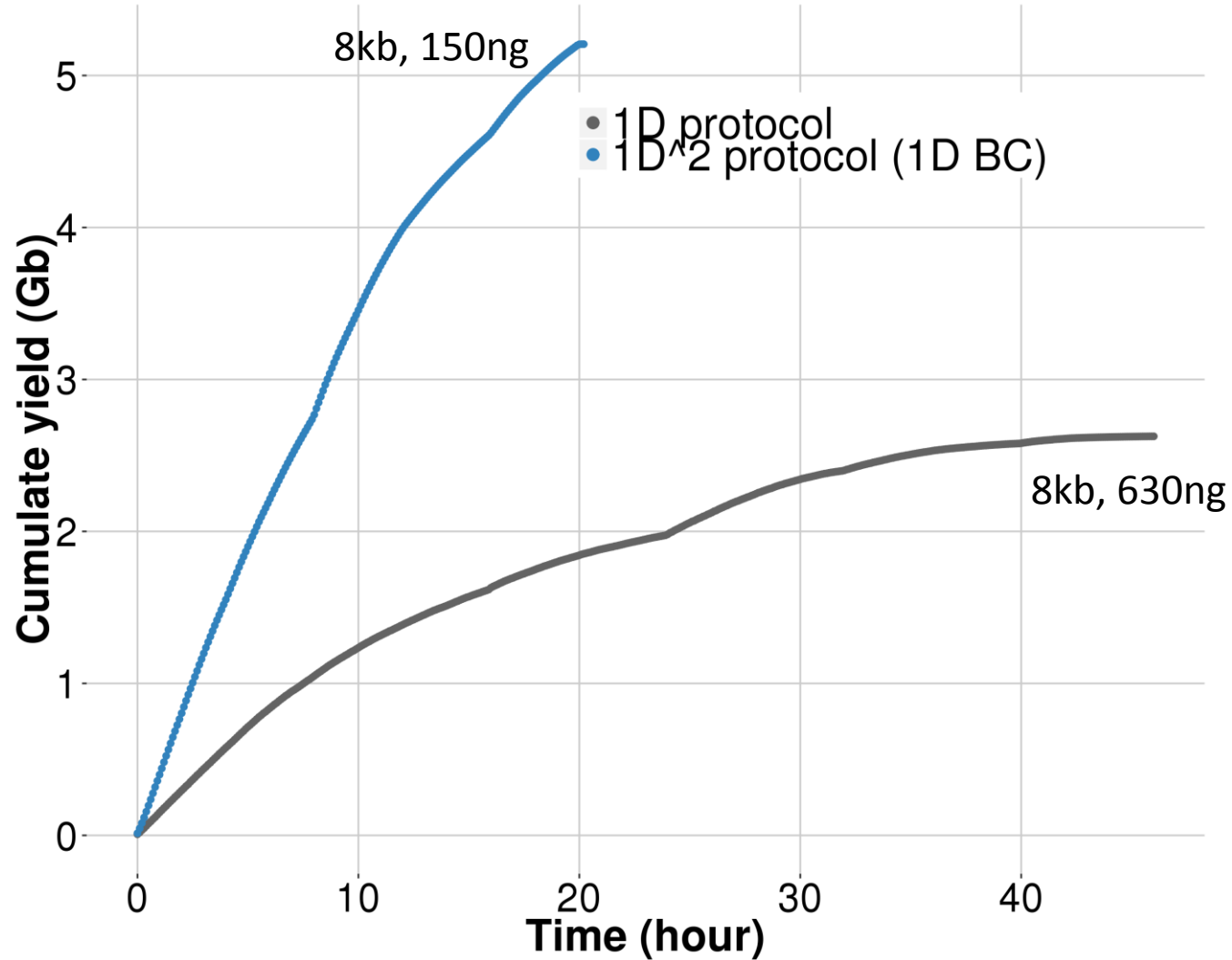
Baptiste Mayjonade, J  r  me Gouzy, Fabrice Roux





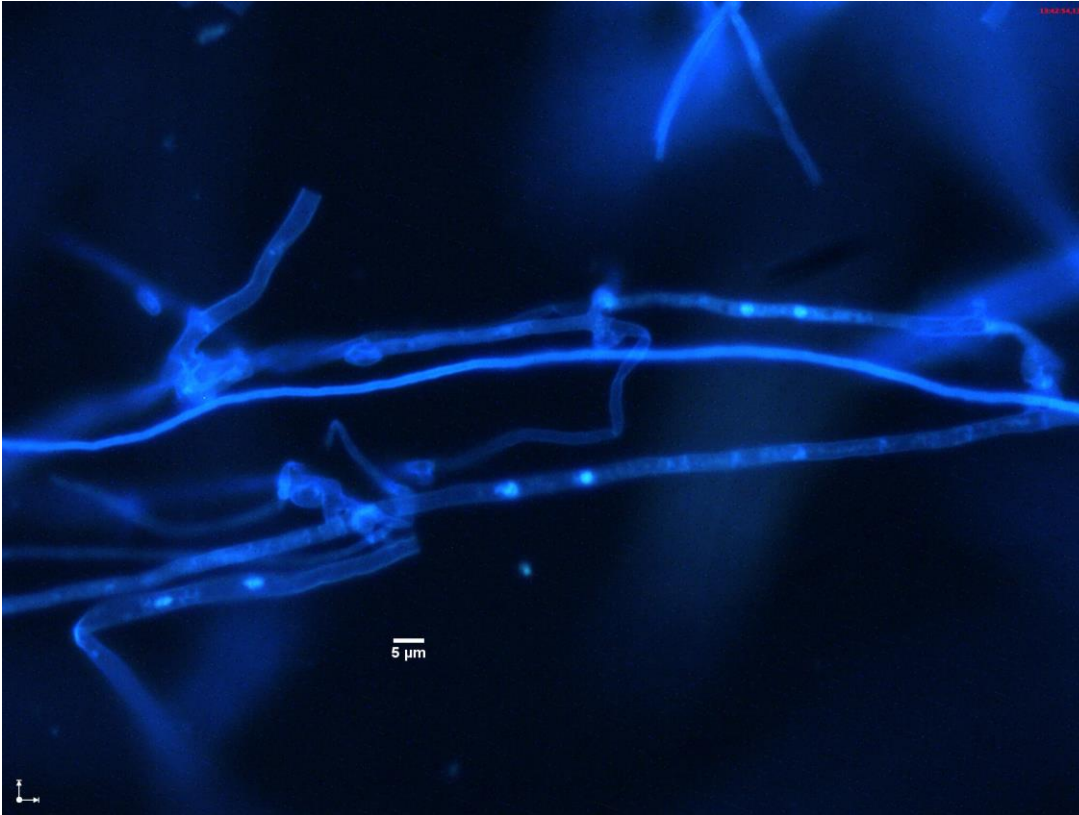
# MinION vs Users needs

New chemistry : 1D<sup>2</sup> / R9.5



# Complex genome

*Ganoderma boninense* – Oil palm pathogen



2 nucleus  
Heterozygous

Illumina results :

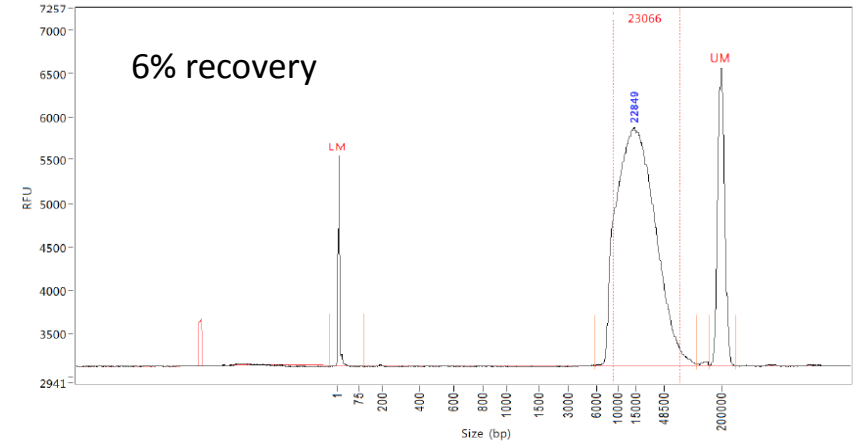
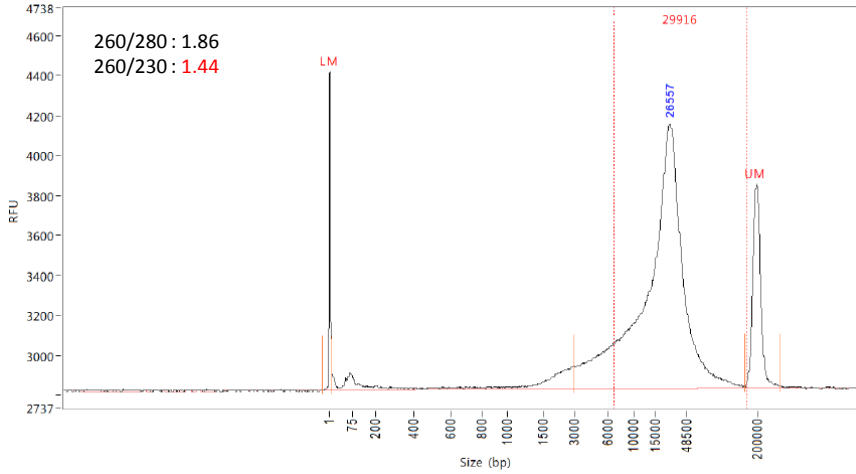
<b>Number of contigs</b>	<b>10164</b>
Longest contig	170 kb
<b>N50 contig length</b>	<b>12 Kb</b>
L50 contig count	1367

Repeated regions > 10 kb

50 Mb

# Pacbio vs Contaminated DNA

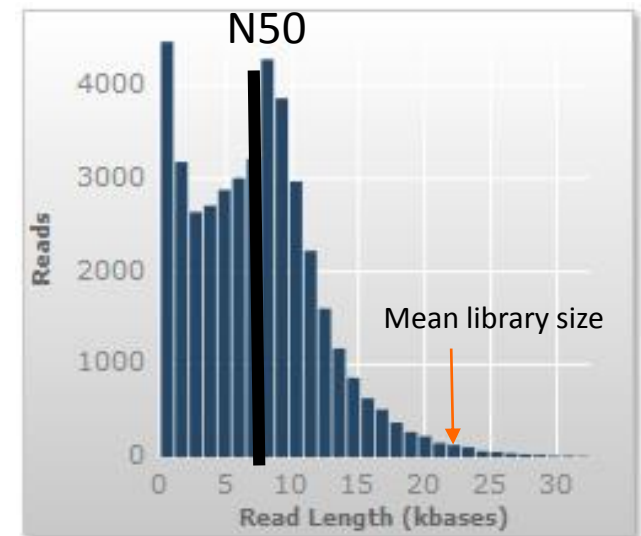
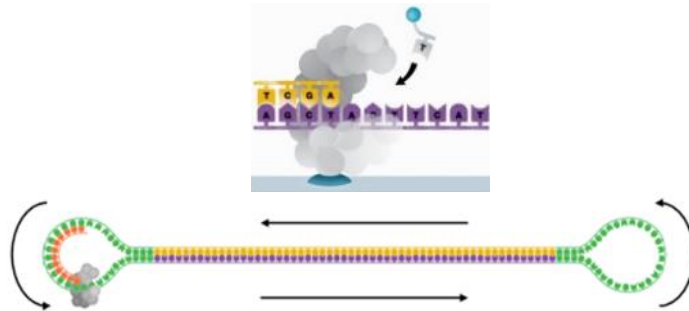
*The polymerase stops*



Pacbio

Polymerase Reads

Length
16956
17039
21560
15856
9910
15648
15479
16879



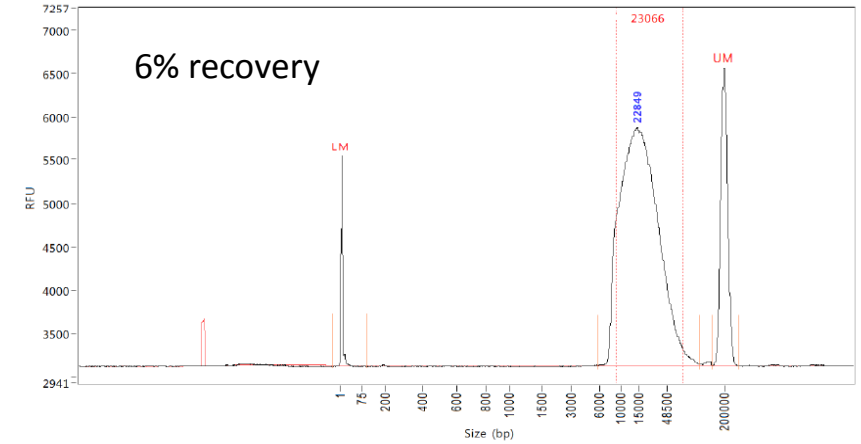
# Pacbio/MinION vs Contaminated DNA

*The MinION keep going when the Pacbio polymerase stops*

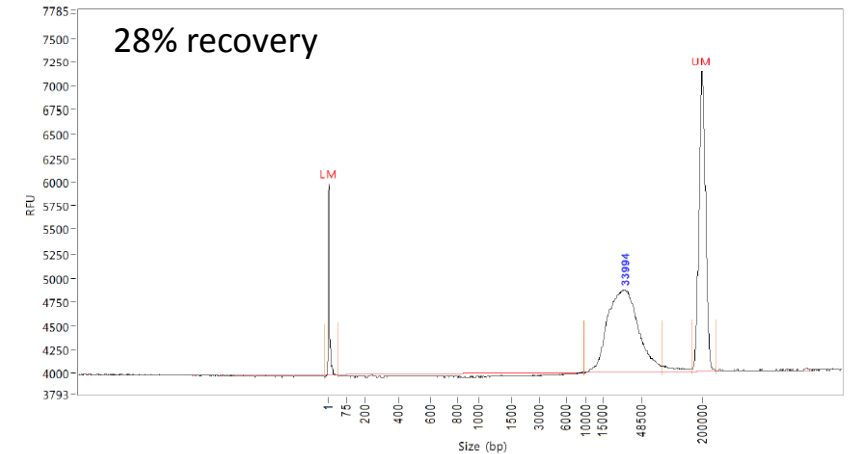
	Pacbio	Nanopore
Library prep	5 days (€€€€)	1 day (€)



Pacbio



MinION



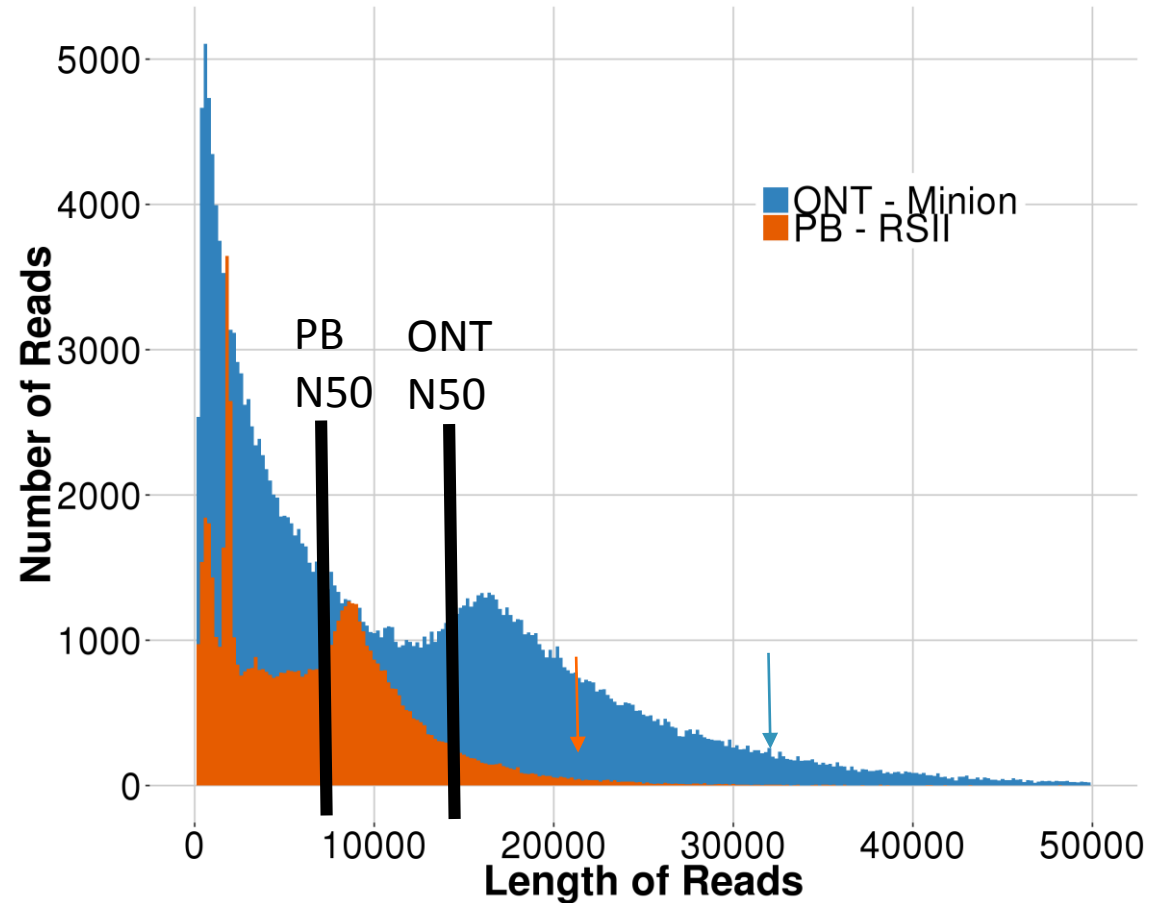
# Contaminated DNA + Complex genome

Technologies comparison

	Pacbio	Nanopore
Library prep	5 days (€€€€)	<b>1 day (€)</b>
Reagents	6 SMRT	<b>1 FC</b>
Quantity of data	4.4 Gb	4.3 Gb
WindowMaker Haplomerge 1x Pilon	<i>Ref + Alt</i>	<i>Ref + Alt</i>
Genes complete	92 %	89 %
Proteins found	89 %	86 %

## Close results

- 2 or 3 x Pilon ?
- Systematic errors ?
- Contaminant/Signal ?
- Tools ?





# Conclusion

§ MinION technology : Long DNA fragments = long reads



§ Depending on the biological question, the MinION can give a partial or a complete answer

§ Most part of the time, carry out study using multiple technologies will be the best option

§ To go further and improve genomes assembly :  
scaffolding technologies (10X Genomics, BioNano)...

Tomato genome

Technology	N50
PacBio (RSII 70 x)	3.2 Mb
+ Bionano (2 enzymes)	32 Mb
+ Chromium + Illumina (100x)	45 Mb





# Remerciements



Maxime MANNO, Céline Vandecasteele, Claire Kuchly,  
Cécile Donnadiou, Olivier Bouchez, Gérald Salin,  
Alain Roulet, Céline Roques



Christophe Klopp



Frédéric Breton, Alexandra Vaillant, Leatizia Camus-Kulandaivelu



Tan joon sheong, Sharifah Shahrul Rabiah Syed Alwee, Kwan Yen Yen



Baptiste Mayjonade, Jérôme Gouzy, Fabrice Roux



Guillaume Croville, Jean-luc Guerin



Caroline Callot, Stéphane Cauet, Hélène Berges



Mohammed Zouine, Pierre Frasse, Mondher Bouzayen

